New Jersey Periphyton Bioassessment Development Projects

Trophic Diatom Inference Models and Index Development for New Jersey Wadeable Streams

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Understanding the Relationship Between Natural Conditions and Loadings on Eutrophication: Algal Indicators of Eutrophication for New Jersey Streams

Final Report Year 2

Report No. 03-04

Submitted to the

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Division of Science, Research and Technology

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Executive Summary

Nuisance levels of algae in New Jersey rivers and streams result primarily from high levels of nutrients coming from a variety of agricultural, residential and urban sources. This report presents results of the first two years of a project to develop algal indicators for streams and rivers in the Piedmont ecoregion of New Jersey. These indicators are designed to assess levels and causes of cultural eutrophication. All sites (37) studied for this project are part of the NJ Ambient Monitoring Network. They were sampled in 2000 and 2001 for diatoms, soft-algae and water chemistry. Measurements of algal biomass, algal species composition, physical stream conditions and water chemistry were used to develop models and metrics for quantifying algal biomass and inferring nutrient concentrations from diatoms and soft-algae.

The following summarizes findings of the research presented in this report :

- The relationships between algal biomass measures (Chl *a* and AFDM) and nutrient concentrations were not strong or significant, based on Spearman's rank-order correlations that included data from all the sites. However, variations in contents of Chl *a* can be explained through a combination of basin size (also reflecting river width and light conditions) and nitrogen (NO_3-N) (highly correlated with phosphorus).

- Three hundred and nine diatom taxa were found in the samples. Most were pollution-tolerant species. Only a few soft-algae species, the most common being *Cladophora*, a filamentous green alga, were found often in high abundance in nutrient enriched streams.

- Multivariate analysis of species and environmental variables shows that total phosphorus (TP), orthophosphate (O-P), nitrogen (NO_3-N) and ammonia (NH_3-N) explain significant differences in diatom assemblage composition. This finding provides statistical justification for developing diatom-based models and indices of nutrient conditions.

- Nutrient inference models and indices will be useful as water quality management tools. A model for inferring TP ($r^2_{(apparent)}$= 0.72; RMSE $_{(boot)}$ = 0.33 log µg TP) developed using the complete 2000 dataset (n=85), has good predictive ability with a bootstrapped r^2=0.55, and when tested on the samples collected in 2001 (r^2=0.61).

- Three indices developed for European rivers (Biological Diatom Index, the Polluosensitivity Index and the Trophic Diatom Index) all correlated relatively well with either O-P and/or TP. This suggests that all three methods would provide good nutrient monitoring tools for the rivers of the NJ Piedmont. Simple community metrics (e.g., species diversity) were generally not good indicators of nutrient conditions.

- A combination of indicators is best for monitoring nuisance levels of algae and nutrients in NJ rivers. For monitoring algal biomass, use the EPA Rapid Bioassessment Protocol and measure Chl *a*. To assess levels of phosphorus concentration and their influence on algae,

we recommend using diatom inference models and the European Trophic Diatom Index (TDI).

- In Year 3 of this project a larger data set will be used to further explore the relationships between biomass and nutrients, and to develop and test additional metrics and models. The roles of river size, light and nitrogen concentrations as influences on biomass-nutrient relationships will be further quantified and be accounted for in developing and applying models and metrics.

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List of Abbreviations

AFDM	ash free dry mass
AMNET	Ambient Biomonitoring Network
ANSP	The Academy of Natural Sciences, Philadelphia
boot	bootstrapped
CCA	Canonical Correspondence Analysis
Chl *a*	chlorophyll *a*
DCA	Detrended Correspondence Analysis
IBI	index of biotic integrity
NAWQA	National Water-Quality Assessment
NJ DEP	New Jersey Department of Environmental Protection
O-P	orthophosphate
PCA	Principal Component Analysis
PCER	Patrick Center for Environmental Research
RBA	rapid bioassessment
RMSEP	root mean square error of prediction
SWQS	Surface Water Quality Standards
TN	total nitrogen
TKN	total Kjeldahl nitrogen
TP	total phosphorus
USGS	United States Geological Survey
U.S. EPA	United States Environmental Protection Agency
WA	weighted averaging

1 Introduction

Nuisance levels of algae in New Jersey (NJ) rivers and streams result primarily from high levels of inorganic nutrients coming from a variety of natural, agricultural, residential and urban sources. Excessive algal growth can cause water quality problems and can harm the designated use of rivers and stream in different ways (Dodds and Welch 2000, U.S. EPA 2000b, ENSR 2001). Nutrient enrichment has been shown to increase benthic algal biomass in rivers through addition of both nitrogen and phosphorus (Blum 1956, Francoeur 2001). Many recent studies focus on understanding the effect of nutrient enrichment on excessive periphyton growth in rivers in order to develop management strategies for stream and river eutrophication (Dodds and Welch 2000, Dodds et al. 2002, Biggs 2000, Smith et al. 1999).

Nationwide there is a continuous discussion concerning the establishment of nutrient limits and thresholds; their implementation is different from state to state. The U.S. EPA technical guidance manual for rivers and streams (U.S. EPA 2000a) recommends three approaches for development of nutrient and algal criteria: (1) the use of reference streams, (2) applying predictive relationships to select nutrient concentrations that will result in appropriate levels of algal biomass and (3) developing criteria from thresholds established in the literature. Also, in the Ambient Water Quality recommendations for U.S. EPA Rivers and Streams Aggregate Nutrient Ecoregion IX (U.S. EPA 2000b), U.S. EPA recommends establishing nutrient reference conditions in rivers and streams, using two methods: 1) establishing reference conditions based on the upper 25th percentile (75th percentile) of all nutrient data from all reaches sampled, or 2) determining the lower 25th percentile of the population of all streams within a region. A review of this approach for the New England Interstate Water Pollution Control Commission revealed that the ranges of predicted biomass production responses to nutrients, as tested for the subecoregions 59 and 84, would be below consensus threshold values (ENSR 2001). Nevertheless, the establishment of reference conditions based on percentiles will set different threshold values in different regions, depending on the range of overall water quality in the rivers of each particular region. These thresholds will be too high in ecoregions with rivers having predominantly high nutrient concentrations as compared to ecoregions with mainly low nutrient rivers and vice versa. The applicability of this method to the NJ Piedmont ecoregion needs further review. In this study, we apply the proposed U.S. EPA percentile method to the NJ Northern Piedmont dataset, in order to calculate reference conditions. We compare our results to the suggested Level III Subecoregion 64 reference conditions, Ecoregion IX (U.S. EPA 2000b) (see Discussion).

The current NJ Surface Water Quality Standards (SWQS) N.J.A.C. 7:9B-1.14(c) state that "*phosphorus as total P shall not exceed 0.1 (mg/L) in any stream, unless it can be demonstrated that total P is not a limiting nutrient and will not otherwise render the waters unsuitable for the designated uses.*" (NJ DEP 2001). The SWQS further state as a nutrient policy, N.J.A.C. 7:9B-1.5(g)2: "*Except as due to natural conditions, nutrients shall not be allowed in concentrations that cause objectionable algal densities, nuisance aquatic vegetation, or otherwise render the waters unsuitable for the designated uses.*" Therefore, current NJ Surface Water Quality Standards are recommending a threshold of 0.1 mg/L TP in streams. Nevertheless,

the validity of this threshold value and the impact of nutrient inputs on algal growth in NJ rivers has not been studied in detail. The NJ DEP therefore needs a better understanding of the impact of the nutrients nitrogen and phosphorous on river and stream systems. Furthermore, nutrient-algal biomass relationships need to be investigated in more detail to develop alternative nutrient criteria and thresholds that can be applied to the state's rivers and streams.

The state of NJ monitors river quality through an extensive Surface Water Quality Monitoring Network, originally measuring chemistry parameters 5 times a year at 200 stations from 1976 to the mid 1990s. Since 1997, 115 stations are measured 4 times a year statewide (NJ DEP 2000). Also, biological indicators are used to monitor river health through the state's Ambient Biomonitoring Network (AMNET) established in 1992. AMNET is an extensive network of 820 stations statewide. Macroinvertebrates are used to assess the biological impairment and geomorphologic conditions of NJ rivers (NJ DEP 2000). Macroinvertebrates are widely used as indicators for organic pollution (Barbour et al. 1999), but they do not reflect inorganic nutrient levels well (Kelly and Whitton 1998, Schwoerbel 1999). Therefore the NJ DEP has a need for application of different additional biological indicators to assess eutrophication and relationships between nutrient conditions and related excessive algal growth in NJ streams.

Algae, especially diatoms, are known to be good indicators of water quality and have been used in the United States since the 1950s (Patrick 1951). Algae are important ecosystem components and they are widely distributed in many habitats. The main advantages of using diatoms as indicators are the following: taxa are numerous and large numbers of individuals can be collected easily; diatoms can be identified to the lowest taxonomic level and strongly correlate with environmental characteristics; they are sensitive to stress, and respond rapidly to environmental change; and finally, they can be stored efficiently. For all these reasons diatoms are valuable and cost-effective indicators for monitoring water quality (Barbour et al. 1999, Dixit et al.1992, Stevenson and Pan 1999).

This study was designed as a two-year project, initiated in July 2000. The purpose of this project was to develop algal indicators of stream and river eutrophication that can be applied in a regulatory context as secondary criteria for identifying nutrient impairment. These indicators are based on relationships between extant water quality criteria (e.g., phosphorus and nitrogen concentrations) and overt signs of eutrophication. They are based on an understanding of algal dynamics in New Jersey streams, and help to distinguish between situations in which nutrient concentrations are high due to natural environmental conditions and those that result from anthropogenic influences.

2 Study hypotheses, goals and approach

Hypotheses:

This study is based on the following working hypotheses:

1) nuisance levels of benthic algal growth in NJ Piedmont rivers are caused by high concentrations of nutrients, especially phosphorus and nitrogen;

2) benthic algal biomass and species composition can be used as indicators of levels and causes of ecological impairment, primarily those related to the nutrients phosphorus and nitrogen.

Goals:

To address these working hypotheses, the general objectives of this study were to:

1) explore the relationships between algal biomass as well as algal species composition and nutrients;

2) develop and test algal indicators of nutrients and water quality applicable to NJ Piedmont rivers;

3) make recommendations to the NJ DEP as to which indicators are best for monitoring nutrient impairment in NJ Piedmont rivers.

Approach:

In order to meet these objectives, the following approach was used in the analysis of the collected data.

1) First, all data were assembled in a database and files were created for data analysis and to present basic data in tables and appendices.

2) We examined site environmental data and created tables. We ran a PCA of environmental variables to understand the relative importance of major gradients and variability among sites.

3) The algal biomass data were summarized and characterized. Basic data were prepared in tables, graphics and appendices, and statistical programs were used to do correlations and regressions among the different measures and to determine how well they agree with each other.

4) The relationships among algal biomass measures and environmental characteristics, especially nutrients, were evaluated. We used ordination and correlation techniques to evaluate the potential for predictive relationships between nutrients, other environmental characteristics and algal biomass.

5) Biomass indicators of nutrient concentrations were developed and evaluated using simple and multiple regressions.

6) The relationships among algal species composition and environmental characteristics, especially nutrients, were evaluated using ordination methods and regressions with diatoms and soft-algae groups.

7) We developed and evaluated species composition-based indicators of nutrient concentrations. We used indicator taxa, simple metrics, European metrics, inference models, a Northern Piedmont TP model and other metrics/indicators.

8) Potential indicators for estimating algal biomass and for inferring relative phosphorus concentrations and overall water quality were compared.

9) Based on our results, we recommended the optimal set of available indicators for use in a monitoring program.

3 Study area

The study area was restricted to the Piedmont physiographic province in New Jersey. This limitation helps to minimize the natural variability in geochemistry, a major factor affecting algal species assemblage composition. In this study we refer to the "NJ Piedmont physiographic province" following the geophysical provinces concept based on traditional geological features (Wolfe 1977) used by the state (http://www.state.nj.us/dep/njgs/enviroed/physiog.htm). We decided to follow this concept, because the delimitation of the Piedmont area follows the bedrock geology, which in turn influences stream geochemistry. The NJ Piedmont physiographic province forms the northeastern extension of Omernik's Level III ecoregion 64, the "northern Piedmont" (Omernik 1987) (Fig. 1). All of our site selection was based on a NJ GIS ARC/INFO Geographic Information Systems (GIS) shapefile (geophysical.shp) received through the NJ DEP, and NJ DEP geological map (http://www.state.nj.us/dep/njgs/geodata) (NJ DEP 1999).

The geomorphology of the NJ Piedmont is characterized by irregular plains with low to moderately high hills and tableland, with elevations increasing to the northwest (US EPA 2000b, Tedrow 1986). The geology of the Piedmont is mainly late Triassic and Early Jurassic age sedimentary rocks, siltstone, shale, sandstone and conglomerate. Resistant gneiss and granites form a 200- to 800-ft (61- to 244-m) high escarpment in the Northwest Piedmont. Gray Sandstone (Stockton formation), red and gray argillite (Lockatong Formation), and red sandstone, including conglomerate (Brunswick Formation), cover most of the Northeast and the southern part of the Piedmont (Tedrow 1986). In the Northeast, volcanic activity associated with rifting of the rock layers of the Piedmont resulted in basalt and diabase intrusions interlayered with sandstone and shale. Both basalt and diabase, being more resistant to erosion, form ridges and uplands in the northeast (http://www.state.nj.us/dep/njgs/geodata).

The climate in the NJ Piedmont is temperate and continental (Tedrow 1986). The average annual precipitation ranges between 43 and 47 in (1092 to 1194 mm) (http://climate.rutgers.edu/stateclim/njclimoverview.html), and approximately half falls during the summer season. Annual snowfall averages 25 in (635 mm) (expressed as snow) in central New Jersey. The annual mean temperature is approximately 54˚F (12.2˚C) at Trenton (40-year average), with 15-20 days usually recording temperatures above 90°F (32.2°C) (Tedrow 1986). In the central Piedmont (Plainfield, NJ) the average July temperature is 75˚F (24°C) and the average January temperature is 30.0°F (-1°C) (ONJSC 1994-2002; http://climate.rutgers.edu/stateclim/norms/monthly/mean.html).

The average number of frost free days is 179 in the central and southern interior (ONJSC1994-2002, http://climate.rutgers.edu/stateclim/njclimoverview.html) and the growing season lasts from mid-March to October (Tedrow 1986).

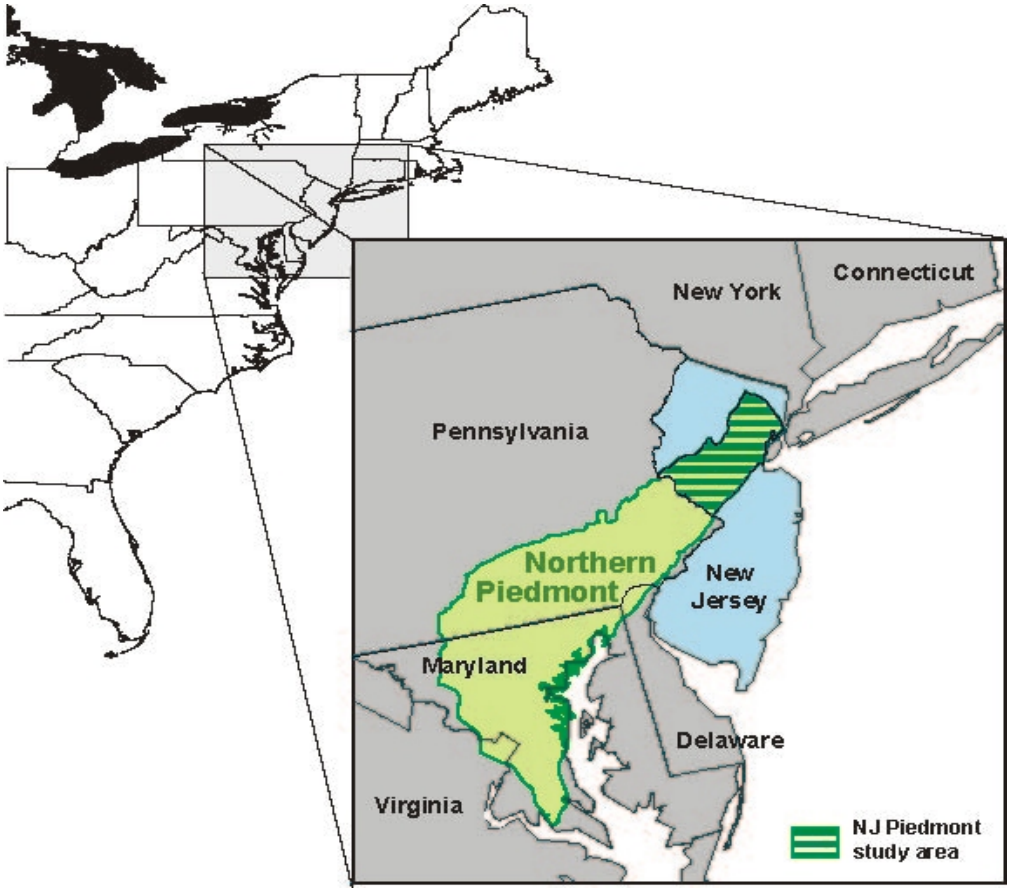

Figure 1: Location of the NJ Northern Piedmont physiographic province within Omernik's Level III ecoregion 64, the "northern Piedmont."

Landuse in the NJ Piedmont is primarily a mix of farmland and urban areas. The urban and industrial areas are concentrated in the northeastern, and to a lesser degree, the southwestern portion of the Piedmont. Nutrient inputs into the rivers of the NJ Piedmont come from a variety of natural, agricultural, residential and urban sources and make this area an ideal region to investigate impacts of nutrient input from different sources. The rivers and streams of this area fall within the U.S. EPA Rivers and Streams Aggregate Nutrient Ecoregion IX, the southeastern temperate forested plains and hills. This Aggregate Ecoregion contains 11 subecoregions, including the Northern Piedmont as subecoregion 64 (Omernik's Level III ecoregion) (US. EPA 2000b). The rivers in this subecoregion have relatively high nutrient concentrations. The median total phosphorous (TP) values, calculated over one decade, range from 2.5 to 1760 µg/L, with a summer mean of 150 µg/L and a median of 70 µg/L. Median total nitrogen (TN) values range from 0.5 to 12 mg/L with a summer mean of 4.8 mg/L and a median of 4.2 mg/L (U.S. EPA 2000b).

4 Methods

4.1 Site selection

We selected an initial set of 30 study sites to be sampled in fall 2000 in cooperation with NJ DEP staff, mainly Tom Belton. Because a goal of this study was to develop algal indicators of anthropogenic nutrient increases, it was important to select a suite of sites with relatively similar natural environmental conditions, but with a wide range of nutrient concentrations. The sites are restricted to the Piedmont physiographic province in northern New Jersey, and have a relatively limited range of hydrology, morphology and substrate type. This limitation helps to minimize the variability in geochemistry, a major factor affecting algal species composition. In addition, we used nutrient concentration data from the NJ DEP as an indication of watershed sources of anthropogenic phosphorus and nitrogen. For all sites, chemistry data were available either through the NJ monitoring network program or through the USGS for their monitoring stations. All sites are part of the NJ Ambient Monitoring Network. We selected sites with a range of impairment from no impairment to severe impairment, based on AMNET Macroinvertebrate classifications made in 1992/93 and 1998/99 (Table 1). About one-third of the selected sites were studied in the same year (2000) by the NJ DEP to develop a fish IBI. Three sites (AN0215, AN0318 and AN0321) sampled during 2000 were accidentally located in the Highlands and two sites (AN0382, AN0439) were sampled in the Inner Coastal Plains physiographic provinces (Fig. 2), due to initially inaccurate interpretation of the NJ Piedmont province delimitation. This error was corrected later, and the samples were excluded from development of indicator metrics.

During the second study year (2001) we selected 13 sites (in cooperation with NJ DEP staff), classified in three categories: 1) "new sites" to fill in data gaps in the gradient of phosphorus concentrations, and to supplement the "calibration" set chosen during the first year, 2) "test sites" to evaluate indicators developed during the first year and, 3) "duplicate sites" to investigate variation in algal biomass and diatom assemblage composition between years one and two. Selection criteria were the same as those used during year one with an altered focus within each category: "new sites" have high concentrations of TP (as recorded by the NJ DEP and/or USGS). "Test sites" cover a range from no- to severe-impairment based on AMNET results and have a USGS gaging station. We selected as "duplicate sites," AMNET sites with severe impairment in 1998 and/or that were planned to be Fish IBI sites in 2001 (see Table 2).

All rivers selected for both years are 1st to 6th order wadeable streams. The classification is based on information from the NJ DEP's GIS hydrography stream network line shapefiles for New Jersey counties, generated as line ArcInfo coverages from USGS 1:24,000 Digital Line Graph (DLG) files (http://www.state.nj.us/dep/gis/GIS maps). The sites sampled are located in the following USGS Watershed Management Areas: Central Delaware, Millstone, Lower Raritan, North and South Branch Raritan River, Upper Passaic, Whippany and Rockaway, Arthur Kill, Lower Passaic and Saddle, Hackensack and Packsack and Pompton, Wanaque and Ramapo. Most of them are located in Somerset, Morris and Bergen counties, and a lesser portion are distributed over Mercer, Hunterdon, Middlesex, Union, Passaic and Essex counties.

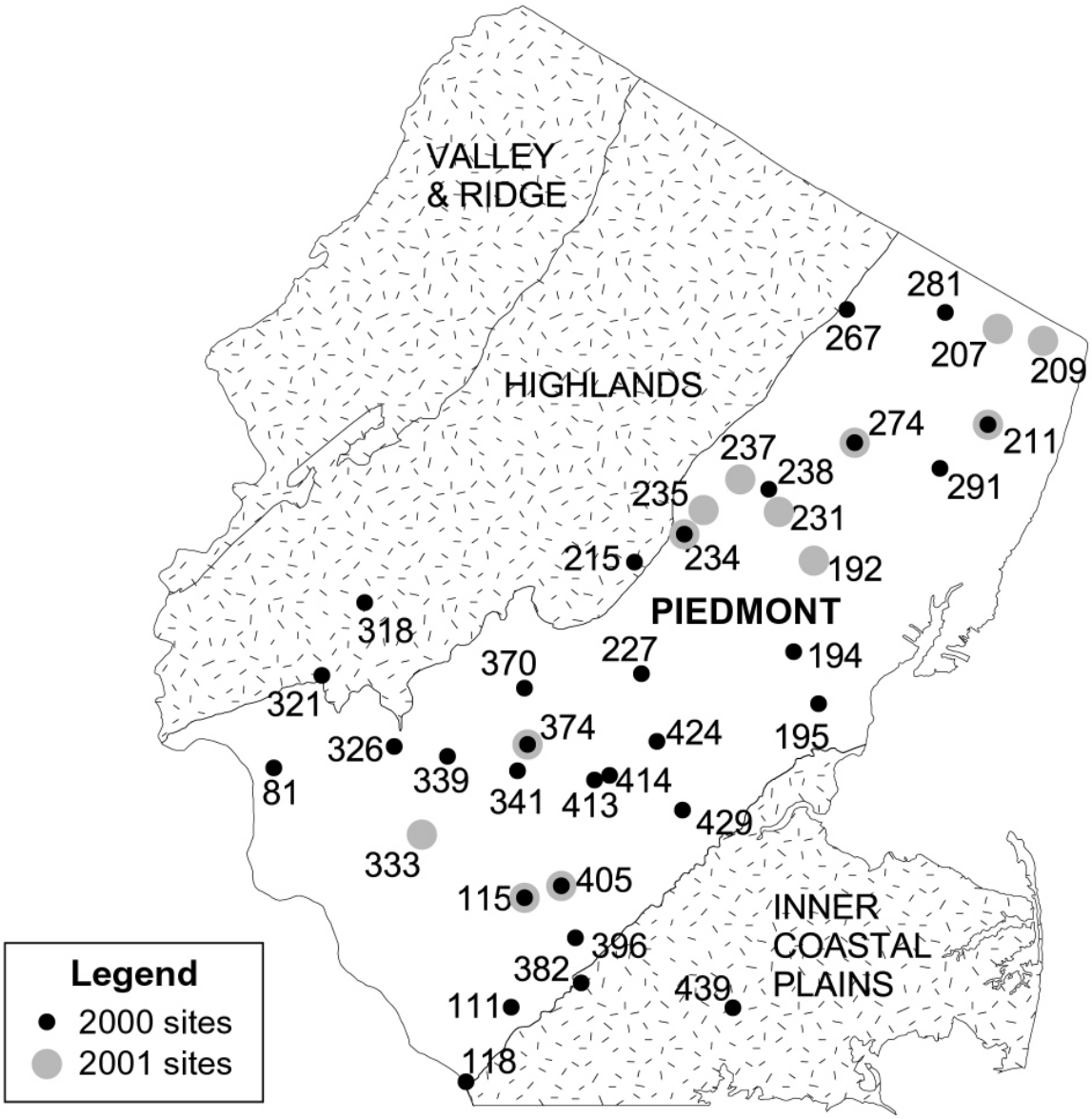

Figure 2: Site locations in the Piedmont physiographic province of New Jersey for sampling years 2000 and 2001. Site numbers correspond to New Jersey AMNET site location IDs. See Tables 1 and 2 for site names and locations.

Table 1: List of sites sampled in 2000.

NJ Site ID	Waterbody	Impairment 1992/93	Impairment 1998/99
AN0081	Nishisakawick Ck	Non-Impaired	Non-Impaired
AN0115	Miry Run	Moderate	Moderate
AN0118	Assunpink Ck	Moderate	Moderate
AN0194	Rahway R	Moderate	Severe
AN0195	Rahway R	Moderate	Severe
AN0211	Van Saun Bk	Moderate	Moderate
AN0215	Primrose Bk	Non-Impaired	Non-Impaired
AN0227	Dead R	Moderate	Moderate
AN0238	Whippany R	Moderate	Moderate
AN0274	Passaic R	Moderate	Non-Impaired
AN0318	Spruce Run	Non-Impaired	Non-Impaired
AN0321	Mulhockaway Ck	Non-Impaired	Non-Impaired
AN0341	Raritan R S Br	Moderate	Non-Impaired
AN0370	Lamington R	Non-Impaired	Non-Impaired
AN0374	Raritan R N Br	Non-Impaired	Non-Impaired
AN0382	Millstone R	Moderate	Moderate
AN0396	Heathcote Bk	Severe	Non-Impaired
AN0414	Millstone R	Moderate	Moderate
AN0424	Bound Bk	Moderate	Moderate
AN0439	Manalapan Bk	Severe	Moderate
AN0111	Shipetaukin Ck	Severe	Moderate
AN0234	Whippany River	Severe	Non-impaired
AN0267	Ramapo River	Moderate	Non-impaired
AN0281	Saddle River	Non-Impaired	Moderate
AN0291	Saddle River	Severe	Moderate
AN0326	S Br Raritan River	Non-Impaired	Moderate
AN0339	Pleasant Run	Moderate	Non-impaired
AN0405	Pike Run	Moderate	Severe
AN0413	Royce Bk	Moderate	Severe
AN0429	Mile Run	Moderate	Severe

Table 2: List of sites sampled in 2001.

NJ Site ID	Waterbody	Impairment 1992/93	Impairment 1998/99
AN0115	Miry Run	Moderate	Moderate
AN0192	Rahway River	Moderate	Moderate
AN0207	Pascack Bk	Moderate	Non-impaired
AN0209	Tenakill Bk	Severe	Severe
AN0211	Van Saun Bk	Moderate	Moderate
AN0231	Passaic River	Moderate	Severe
AN0235	Whippany River	Moderate	Moderate
AN0237	Troy Bk	Moderate	None
AN0274	Passaic River	Moderate	Non-impaired
AN0333	Neshanic River	Moderate	Moderate
AN0374	N Br Raritan River	Non-Impaired	Non-impaired
AN0405	Pike Run	Moderate	Severe

4.2 Sampling period

During both years, samples were collected by ANSP staff Mike Hoffmann, Diane Winter and Karin Ponader from August through October. The first year sites were sampled from 9 August through 3 October 2000. In the second year, sampling was completed between 20 and 26 August 2001. During the 2000 field season, sampling was suspended for two weeks to wait for rivers to recover from the scouring effect of high flow conditions caused by very heavy rainfall events during the second week in August. We chose to sample in late summer because the influence of higher streamflow velocity and discharge on algal assemblage composition is lowest during this period. Based on the average of monthly mean streamflow calculated for 77 years (since 1925), the lowest flow records in NJ rivers were measured in August, September and October (http://www.waterdata.usgs.gov). Samples collected during this time are also most directly comparable with sample data from other studies conducted in the area, such as the National Water-Quality Assessment Program (NAWQA), the EPA Riparian Reforestation Project and the Growing Greener Project all conducted at the PCER (http://www.acnatsci.org/research/pcer/projects.html). All these projects were conducted at the ANSP and follow USGS NAWQA Periphyton sampling protocols recommending sampling periods to be conducted during normal, low- or stable-flow periods (Moulton et al. 2002, Porter et al. 1993).

4.3 Collection of samples/data

4.3.1 Site characterization/establishment of sampling reaches

All sites sampled are located at NJ DEP AMNET monitoring stations, which are defined as the intersection of a road and the river to be sampled. According to NJ DEP field sampling protocols, we sampled on the upstream side of the bridge to minimize the effect of inputs from automobile use/traffic and street maintenance. Some exceptions were made at sites where conditions did not allow sampling upstream and where the downstream side was considered more representative of the river habitat. Prior to collection of water chemistry and algal samples, we took detailed notes on general physical site characteristics, geomorphology, weather conditions, overt signs of human impact, etc. The sampling area was divided into three sampling reaches, so that variability among different sections of the rivers could be assessed. The three sections were determined using the following criteria: each section should contain a minimum of 2 riffles and 2 pools and the length of each reach should be approximately 10 times the channel width. Commonly used guidelines (Fitzpatrick et al.1998) recommend a minimum reach length of 150 m for wadeable streams. We did not follow these guidelines and established shorter reaches because of the generally smaller width of the rivers sampled in the NJ Piedmont area. The average width of the rivers sampled was 13 m (range of 3-50 m) and the average length of the established sampling reaches was 44 m (see Appendix 1a). We believe our criteria were satisfactory for establishing reaches that represented the local variability within the river. Once the reaches were established, we recorded information on all three sections, made site drawings, and measured the physical characteristics of the sampling sites. Sites are documented with digital images (Sony MVC-CD1000), burnt on a CD and submitted to the NJ DEP. For each section, we made a visual

estimate of percent substrate type (boulder, cobble, gravel, sand, silt, bedrock) and flow velocity. Light conditions (percent open canopy cover) were measured using a spherical densiometer.

4.3.2 Water chemistry samples

Water chemistry samples were taken prior to algal sampling to avoid disturbance of the water column and sediments. Samples were taken using a plastic syringe with an attached filtration device. Laboratory analysis of NO_3-N, NH_3-N, O-P and TP was performed by the PCER Geochemistry Section (Velinsky 2000). In 2001, we took additional samples for analysis of chloride, total alkalinity, total hardness and conductivity. Samples were cooled immediately on ice in the field and shipped to the ANSP where samples for nutrient analysis were frozen immediately. Results of these analyses were used to supplement those collected by the NJ DEP. Samples collected directly by ANSP in the field better represent conditions near the time that algal samples were collected, and provide information of the nature and magnitude of variation in water chemistry.

4.3.3 Diatom and biomass/soft algae samples

Samples were collected from natural rock substrates using techniques consistent with those used in the USGS NAWQA program (Moulton et al. 2002) and the EPA Rapid Bioassessment protocols for periphyton (Barbour et al. 1999). All sampling procedures are documented in a PCER protocol (Charles et al. 2002). Two types of samples were taken. One, a composite diatom sample, was created by randomly selecting 4-5 rocks of ca. 5 cm diameter. The rocks were carefully selected from mid-stream and were free of visible filamentous algae. In 2000, samples from sticks, gravel or sand were collected at five sites where no rocks were available (AN0194, AN0227, AN0238, AN0382, AN0414). Algae were removed from the rocks by scraping and brushing, placed in plastic containers and preserved in the field by keeping them on ice in a cooler. The second type of sample was a quantitative composite biomass sample collected for measurement of chlorophyll *a* and ash-free dry mass (AFDM). These samples were analyzed by the Patrick Center for Environmental Research's (PCER) Geochemistry Section. Three bigger rocks (with an average diameter of ca.10 cm) were selected randomly to represent the distribution of algal coverage within each reach section. Rock surfaces were scraped, and outlines of rocks were drawn on waterproof paper. Surface area was measured using an aluminum foil method (Moulton et al. 2002, Ennis and Albright 1982). NJ DEP guidelines were followed for preservation and storage of Chl *a* samples. All samples (diatoms and Chl *a*) were preserved by keeping them on ice in a cooler and were shipped to the ANSP over night for immediate treatment in the laboratory the next morning. In total, 85 diatom samples were taken during 2000. Only 71 samples were collected for biomass in 2000; biomass samples were not collected at 6 sites with sandy substrate. In 2001, we only sampled rock substrate, collecting 35 diatom and biomass samples in total. Both year's datasets combined contain a total of 120 diatom samples and 106 biomass samples.

4.3.4 Visual biomass estimate (EPA rapid bioassessment protocol)

In addition to algal sample collection, the percent cover and thickness of algal growth was measured using the Rapid Periphyton Survey Method (EPA Rapid Bioassessment Protocol) developed by the U.S. EPA (Barbour et al. 1999). This method provides a quantitative estimate of

filamentous and other types of algae that often have patchy distributions and whose biomass is difficult to quantify. For each sampling section, we measured percent biomass cover for each algal group along three transects across the river. Length of filamentous strains and thickness of algal mats per algal group were also recorded. For each section, an average was calculated from the three transects and used in data analysis. We collected additional samples for algal identification and examination under the microscope when identification in the field was not possible.

4.3.5 Additional data (water chemistry, landuse)

In addition to the water chemistry and biomass data produced at the PCER, all other data were provided by the NJ DEP through Tom Belton. Landuse data for each watershed were assembled by Jack Pflaumer (NJ DEP). Also, Jack Pflaumer sent ANSP most of the additional chemistry data records collected by USGS and NJ DEP at the surface water monitoring stations. He assembled available data for the sampled periphyton sites for each sampling year. Additional data were retrieved by Karin Ponader through the USGS "water quality samples for USA" webpage (http://www.waterdata.usgs.gov/nwis/qwdata) and the NJ 2001 Water-Resources Data report (Reed et al. 2002). Because sampling was not necessarily done at the same time by USGS and PCER staff, all USGS/NJ DEP data used in our analysis were measured within a maximum of 4 weeks from algal sampling in the same year.

4.4 Algal sample preparation and analysis

Samples were prepared for algal analysis using standard protocols (Velinsky and DeAlteris 2000). Chlorophyll *a* and ash-free dry mass (AFDM) samples were analyzed by the PCER Geochemistry Section using methods as described in Standard Methods and US EPA method 445 (APHA, AWWA AND WPCF 1992, U.S. EPA 1992). Diatoms were permanently mounted on microscope slides following routine protocols (Charles et al. 2002). A total of 85 slides was prepared for year 1 and a total of 35 slides was prepared for year 2. Per slide, 600 valves were identified to lowest taxonomic level and counted using USGS NAWQA protocols (Charles et al. 2002). Identification was done using common taxonomic references available at the ANSP as well as type material from the ANSP Diatom Herbarium. Over 900 digital images were taken, recording nearly all identified and unidentified taxa. Taxonomic problems were discussed with PCER Phycology Section members, and problematic and unknown species were described and recorded in the ANSP Algae Image database (http://diatom.acnatsci.org). Also, the active participation of Karin Ponader in the Fourth through the Eighth NAWQA Taxonomy Workshops on Harmonization of Algal Taxonomy held at the Academy of Natural Sciences in October 2000, June and October 2001 and May and October 2002 helped in solving taxonomic issues in the NJ Piedmont diatom flora.

Diatoms were counted and recorded directly into a database using the computer program Tabulator, version 3.7.0 (Cotter 1999-2000, Cotter 2001). Count reports are created for each count, including information on assemblage composition, taxonomic notes, etc. The common filamentous algae were identified and semi-quantitative estimates were made of their abundance using a new count method developed specifically for this project (Ponader and Winter 2002). This semi-quantitative procedure is designed to provide percentage estimates of the most common

species of algae that make up the largest proportion of the algal biomass for each sample. The method consists of two steps. The first step involves identifying the most common genera/species and estimating the relative percentage of each of these in the algal assemblage. In the second step, the relative percentage that each genus/species contributes to the algal biovolume in the sample is estimated. Because this is a semi-quantitative method, cells are not counted or measured, but a general estimate is made, which describes the relative proportions of the common genera and species observed in the sample through examination of several transects.

4.5 Data storage and documentation

All data collected during this project were properly stored in the PCER Phycology section's database management system, the North American Diatom Ecological Database (NADED) using Microsoft Access 2000. The field sheets were scanned and all digital images of sites and samples were burnt on CDS. Copies are available on request. All image documentation and site information were archived in the database.

4.6 Data analysis

4.6.1 Water chemistry: PCA to explore gradients and variability among sites

Prior to examining the relationships among algal biomass, species composition and environmental variables, we performed a Principal Component Analysis (PCA) using Canoco for Windows version 4.02 (ter Braak and Prentice1998). The environmental variables were centered and standardized. The aim of running a PCA was to discover the principal patterns of variation within the environmental variables measured and how they relate to sampling sites. Outliers were defined as samples with scores falling outside the 95% confidence limit about the sample score means in a PCA of the environmental variables (Hall and Smol 1992, Birks et al. 1990b).

4.6.2 Algal biomass

4.6.2.1 *Spearman's rank-order correlation (correlations between nutrients, algal biomass and algal species composition)*

A Spearman's rank-order correlation was run using the program SPSS version 11.0 for Windows. We chose to run this analysis because many of the algal biomass variables listed below are not measured on a continuous scale and none of them had normal distribution (Dytham 1999). Included in this analysis were the following data for all 106 samples collected during both years: Chl *a* and AFDM data, nutrient measurements (TP, O-P, NH_3-N, NO_3-N), percent open canopy cover and substrate type, soft algal species composition data obtained through the semi-quantitative analysis for all 106 samples, as well as different measures of algal biomass. The latter were created through combination of different categories, e.g., different algae types and their abundances multiplied by estimated algal thickness and length rank. In total, 110 variables and combinations of variables/categories were used in the analysis. Because of the size of the complete report file (over 100 pages) we only list here (Appendix 3) the results of a reduced set of 67 selected variables, excluding the combinations (e.g., algal type multiplied with length rank

or thickness etc.). The results of the strongest and most significant relationships are listed in sections 5.2.1.1 and 5.2.2.2.

4.6.2.2 *Forward stepwise regression* (*analysis of principal factors influencing algal biomass)*

To help determine the principal factors influencing algal biomass, we examined correlations among algal biomass, nutrients, geomorphology and light conditions, running a Forward Stepwise regression with Sigma Stat 2.03. All chemical variables (except pH) were log10 transformed. The substrate categories were analyzed both separately and combined into different categories. We separated bigger hard substrate types into two categories, one including only bedrock and boulder and the other containing cobble and gravel. We created two other categories, one including all bigger substrate from bedrock to gravel and another combining all smaller and soft substrate (sand, silt and clay).

4.6.3 Diatom assemblages

Numerical analyses were performed to investigate the factors affecting diatom species composition, and to determine whether species composition was influenced by nutrients strongly enough to justify development of inference models. We used Canoco for Windows version 4.02 (ter Braak and Prentice1998) to perform these analyses. Because their distributions were skewed, we log10 transformed all water-quality variables included in the analysis, except pH. All diatom species identified in the counts from all the sites sampled in 2000 were included in the ordinations.

4.6.3.1 *Detrended Correspondence Analysis (DCA) to determine principal patterns of variation in diatom species composition*

A detrended correspondence analysis (DCA) was performed to determine the gradient length as a measure of the maximum amount of variation in the diatom data. The gradient length was 2.6 for the first axis, exceeding the value of 2 standard deviation (SD) units, recommended as the point above which unimodal techniques should be used for further analysis and development of calibration sets (Jongmann et al. 1995, ter Braak and Prentice 1998). In the same DCA of the species data, outliers were determined as samples with sample scores falling outside the 95% confidence limit about the sample score means (Hall and Smol 1992).

4.6.3.2 *Data screening*: *environmental variable with extreme influence on species composition*

All methods for screening data to remove outliers prior to developing diatom inference models follow standard procedures used in several publications (Fallu et al. 2000, Hall and Smol 1992, Winter and Duthie 2000). In our study, after outliers were determined in a PCA of the environmental variables and/or in a DCA of the species data (see sections 4.6.1. and 4.6.3.1), the second step was to delete samples that had an environmental variable with an extreme influence other than either TP, O-P, NO_3-N or NH_3-N on the diatom species composition (Birks et al. 1990b, Hall and Smol 1992). In this case, samples were deleted if their residual length on the environmental variable axis fell outside a 95% confidence limit as detected in a CCA constrained to the variable to be reconstructed (Hall and Smol 1992).

4.6.3.3 *Ordination analysis: CCA (influence of environmental variables on diatom species composition)*

To identify the variables that explained a significant amount of variation in diatom species composition and that had an independent influence on diatom species distribution, we ran a series of CCAs constrained to one variable at a time. We calculated the ratio of the sum of the first constrained eigenvalues (λ_1) to the sum of the second unconstrained eigenvalues (λ_2). The variables with highest values of λ_1 / λ_2 were selected as likely to have the most influence on diatom species distribution (Winter and Duthie 2000). Also, as part of the same CCAs that were constrained to one variable, the statistical significance of each variable on the first canonical ordination axis was evaluated using Monte Carlo permutation tests (199 permutations, $p \leq 0.05$) (Fallu et al. 2000). Variables that did not explain a significant amount of variation in diatom composition were excluded from the dataset used for development of inference models.

4.6.3.4 *WA-regression and calibration (development and testing of nutrient inference models)*

Nutrient inference models were developed with weighted averaging (WA) regression and calibration techniques using WACALIB version 3.5 (Birks 2001, Line et al. 1994). Diatom species optima and tolerances were calculated for the nutrient variables TP, O-P, NH_3-N and NO_3-N. The models included all diatom species. Species abundance (%) was transformed by calculating the square root of each value. Species tolerances were corrected by deshrinking with an inverse regression procedure (ter Braak and van Dam 1989). We used bootstrapping (1000x) (Birks et al. 1990b) to estimate the root mean square error of prediction (RMSEP) of each model developed. The predictive power of the developed models was assessed based on the $r^2_{(boot)}$ and the $RMSEP_{(boot)}$. The model with the highest predictive power and the lowest RMSEP is the best model calculated. To evaluate the performance of the TP model developed, we tested them on samples collected in 2001, performing WA-calibration using CALIBRATE version 0.61 (Juggins and ter Braak 1997, Juggins and ter Braak 2001). The performance of the model applied was assessed using statistics describing the correlation between the observed versus inferred values (Birks et al. 1990b).

4.6.4 Calculation of diatom metrics

4.6.4.1 *Diversity metrics and other simple metrics*

Diatom diversity indices and other simple metrics were calculated using 98 diatom samples from both years, following Barbour et al. (1999). We calculated the number of diatom taxa in the sample (# Taxa), the Shannon-Weiner diversity index (S-W Index), the percent of total diatom valves made up of taxa that occurred in >10% abundance (Percent Dominants), the percent of total diatom valves made up by the most abundant taxon (% Dominant Taxon), the ratio Centrales /Pennales C/P), and finally, the Siltation Index (% Siltation Index), which is the sum of the percent abundances of all species in the genera *Navicula, Nitzschia, Cylindrotheca,* and *Surirella*. These are common genera of predominantly motile taxa that are able to maintain their positions on the substrate surface in depositional environments (Bahls 1993). We evaluated the use of these

indices in conjunction with different types of landuse, running a Spearman's rank-order correlation using Sigma Stat 2.03.

4.6.4.2 *European diatom indices*

Twelve different diatom indices, widely used in Europe, were calculated for the NJ diatom dataset. In our study we were specifically interested in the results of the Trophic Diatom Index (TDI) (Kelly and Whitton 1995, Kelly 1998), mainly reflecting nutrient conditions (especially TN and TP), as well as in the Biological Diatom Index (IBD) (Prygiel and Coste 1999) and the Specific Polluosensitivity index (IPS) (Coste in Cemagref 1982), both reflecting overall impairment conditions. The calculations were done by Luc Ector (Centre de Recherche Gabriel Littmann, Luxembourg) with OMNIDIA, a program specifically designed for calculations of diatom indices (Lecointe et al. 1993).

5 Results

5.1 Environmental data

5.1.1 Water chemistry, biomass concentrations and summary of site characteristics

Table 3 summarizes the nutrient and biomass characteristics measured at all sites in both years. TN concentrations were calculated for 25 samples only, due to missing TKN measures in the available USGS data. For information on the full dataset used and all variables measured, see Appendices 1a and 1b.

Table 3: Statistical summary of nutrient and biomass concentrations at all sampling sites. Data include 2000 and 2001 samples. TP, O-P, NO_3-N, NH_3-N, Chl *a* and AFDM were measured at the PCER. TN is calculated combining PCER data (NO_3-N) and USGS data (TKN available from 25 stations only).

Variable	Mean	Median	Minimum	Maximum	*n* samples
TP (mg/L)	0.15	0.07	0.01	1.30	41
O-P (mg/L)	0.11	0.04	<0.01	1.14	41
TN (mg/L)	2.13	1.77	0.89	5.74	25
NO_3-N (mg/L)	1.92	1.33	0.23	7.55	41
NH_3-N (mg/L)	0.05	0.03	<0.01	0.18	41
Chl *a* (mg/m^2)	109	81.0	2.19	1115	106
AFDM (g/m^2)	20.5	12.7	3.80	153	106

Figure 3 shows TP and Chl *a* concentrations measured for both sampling years. The sites are ordered by increasing TP concentrations. In comparison, the sites sampled in 2001 have generally higher TP concentrations than in 2000, which was one of our goals when selecting sites for 2001. Comparison of TP and Chl *a* values at sites that were resampled in 2001 does not show significant differences between both sampling years. Figure 3 also shows that Chl *a* values do not increase significantly with increasing TP, reflecting challenges of using Chl *a* as an indicator of increased nutrient contents. This is discussed further in the statistical analysis and the discussion. In our dataset, 46% of the samples that were collected from sites with concentrations of 0.1 mg/L of TP in the water column show Chl *a* concentrations greater than 100 mg/m^2. The mean for all samples collected in 2001 and 2002 is 109 mg/m^2 Chl *a*. Observations in the field have shown that samples with Chl *a* >150 mg/m^2 were taken from sites with extreme algal growth based on visual estimates.

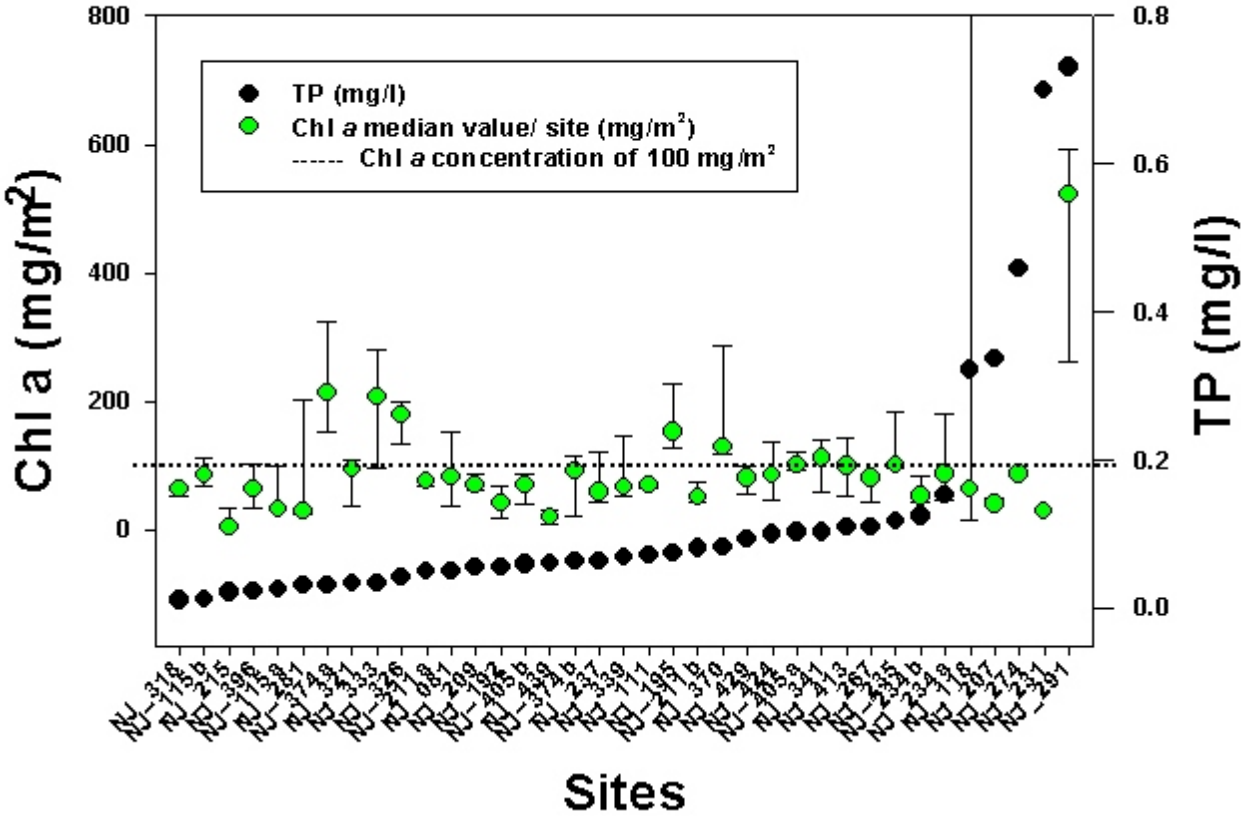

Figure 3: TP and Chl *a* concentrations measured in 2000 and 2001. Sites ordered by increasing TP (black circles). At each site three Chl *a* measurements were taken (one per reach). Green circles represent median Chl *a* concentrations and error bars indicate maximum and minimum Chl *a* concentrations per site. Site numbers a and b indicate that sites were sampled in both years (a=2000 and b=2001).

The visual estimates of algal cover along transects (see description of method under section 3.3.4) are summarized in Figure 4. To highlight the major trends, only samples with Chl *a* concentration exceeding 100 mg/m^2 are represented in this graph. Sites are ordered by increasing Chl *a* content. Estimates of filamentous algal cover showed that at most sites % estimated diatom (chain-forming) cover was most important, followed by % *Cladophora*. The third important group was % thin diatom cover, represented by thin diatom mats that do not form chains or filaments. Finally, % blue-green algae was the next most abundant cover, followed by % green algae cover. There was no clear correlation between percent visual estimate of algal cover of individual algal groups and measured Chl *a*. Therefore, we further investigated whether the visual estimate of total biomass shows any significant relationship with measured biomass (Chl *a* and AFDM) using correlation analyses as described in section 5.2.

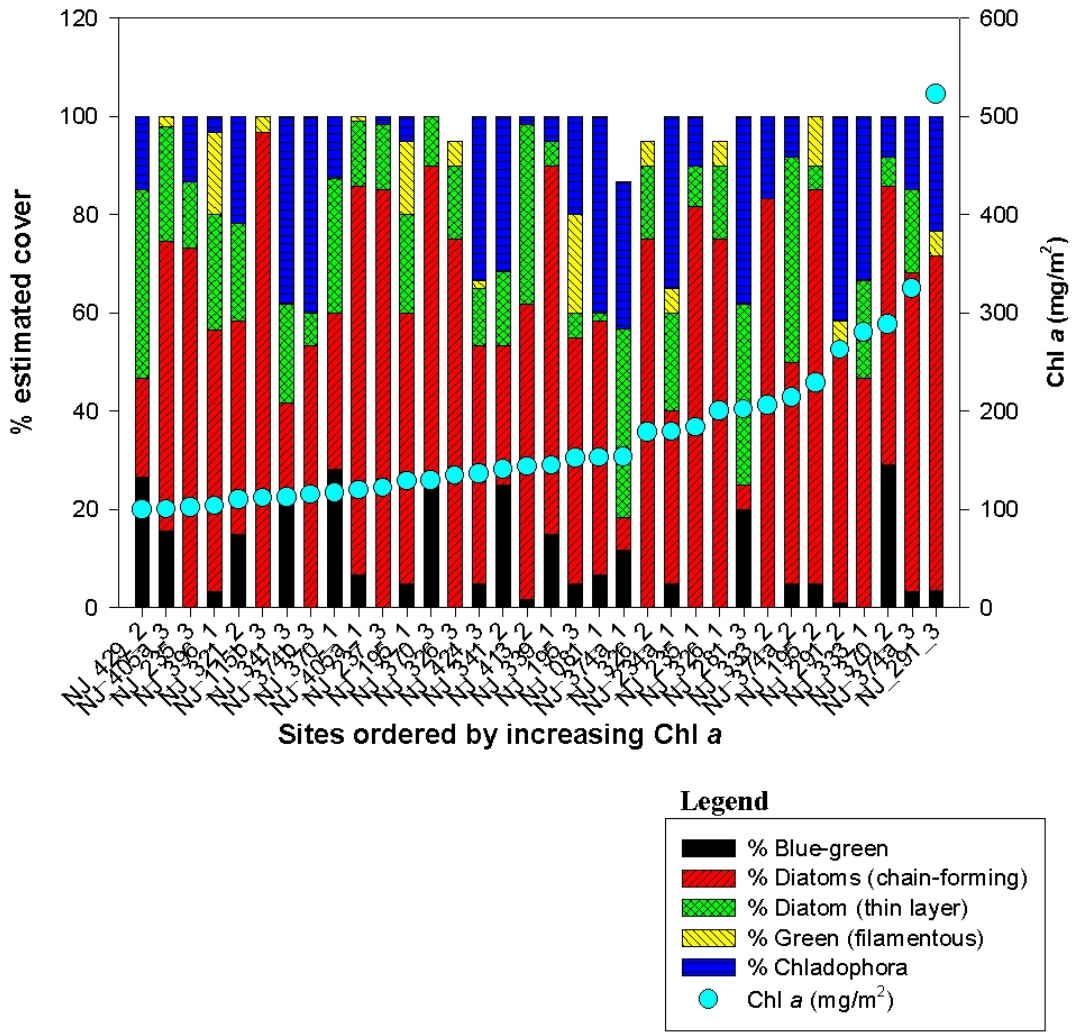

Figure 4: Main algal groups and their contribution to percent estimated algal cover, ordered by increasing Chl *a* content measured at the site. Only samples with Chl *a* contents exceeding 100 mg/m^2 are represented. Samples collected at the same site, but in different years are marked on the x-axis with "a" (for 2000) and and "b" (for 2001). The three different sections per site are indicated by 1, 2 and 3.

5.1.2 PCA: gradients and variability in environmental data

A PCA was run to identify principal patterns of variation among the measured environmental variables. We included the maximum number of variables in the analysis, but only a limited number of sites contained records for all variables. Therefore, 16 variables and a total of

24 samples in 2000 and 2001 were included. The PCA shows that the sites are distributed along three main axes (Fig. 5). The percentage of variance explained by the first two axes was 54%, with eigenvalues of λ_1= 0.34 and λ_2 =0.20, respectively. Axis 1 reflected a gradient of several nutrients (TKN, TN, O-P, TP, NO_3-N) and separated sites with low nutrient concentrations from sites with higher nutrient values. The second axis is mainly influenced by a combination of pH, basin size, DO, and DOC. This axis reflects mainly river width and related DOC loadings, separating narrower rivers with higher DOC loadings from wider rivers with lower DOC concentration. The third axis is influenced mainly by % urban, conductivity and % agriculture, showing that sites are mainly distributed along an urban gradient. Agriculture does not show a strong gradient, as most sites in the NJ Piedmont were sampled in urban areas. Generally, this analysis shows that the sites follow a strong nutrient gradient. The following samples were determined to be outliers: Sites AN0291 and AN0231 showed extreme O-P, TP and NO_3-N concentrations. Site AN0118 was identified to have extreme Chl *a* and nutrient values.

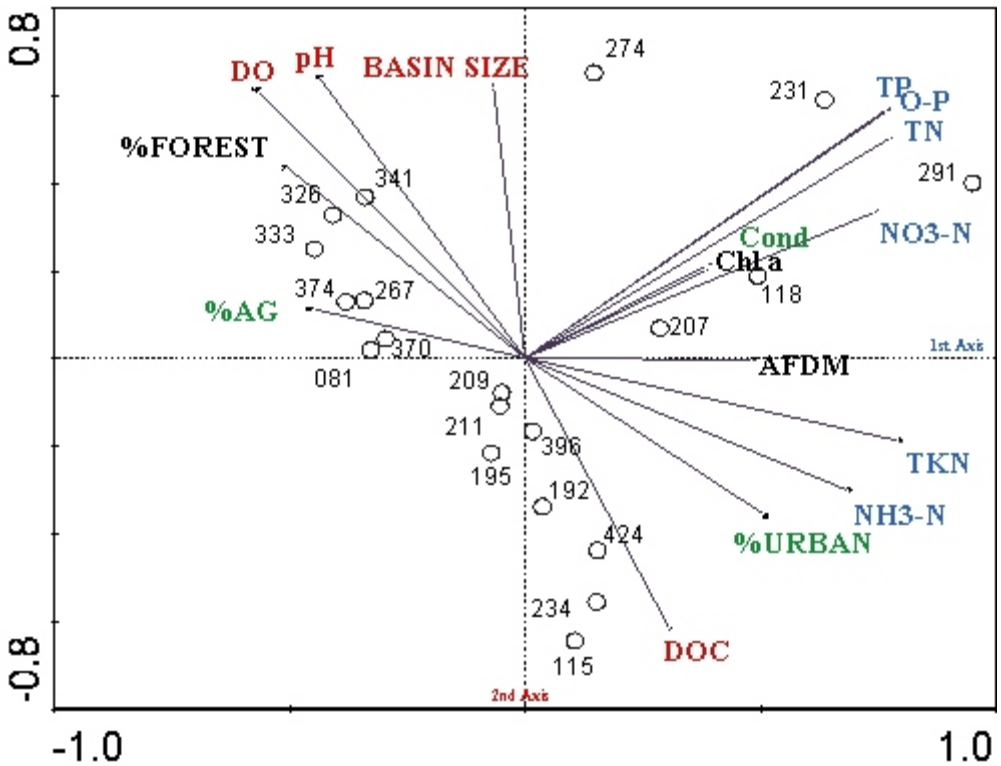

Figure 5: PCA including 24 sites (circles) and 16 variables (arrows) sampled in 2000 and 2001. Numbers represent the last three digits of the NJ site ID (see Table 1). The length of each arrow expresses the "strength" of the influence of the variable on site distribution. Each axis is determined by a combination of variables. The variables are color-coded corresponding to the axes: axis1= blue, axis 2 = red, axis 3 = green.

5.2. Algal biomass

The different methods of assessing biomass in the field and in the laboratory produced a multitude of variables, all expressing algal biomass in a different way. One of the main goals of this study was to identify the strength of nutrient-biomass relationships and their use for development of indicators. Therefore, we needed to know: a) How well do the different measures of biomass correlate? and b) How well does measured/estimated biomass reflect nutrient conditions? To answer both questions a Spearman's rank-order correlation was performed, as described below. The results are presented in sections 5.2.1.1 and 5.2.1.2, answering the above two questions. Finally, we explored how strongly other environmental factors influence algal biomass measures by running a Forward Stepwise regression.

5.2.1 Spearman's rank-order correlation

Relationships among all nutrient variables and biomass data for the full data set from both years' (n=106) data were explored using a Spearman's rank-order correlation matrix. To find out what type of correlation was appropriate to run, we needed to determine if variables in the dataset were normally distributed. We tested each variable using Kolmogorov-Smirnov (K-S) tests (Dytham 1999) using the procedure in Sigma-Stat 2.03. Normality tests failed for all variables when tested on untransformed data, showing that all data were skewed. After log transformation, another K-S normality test was run. The results showed that only Chl *a* (log) passed the test, and that the data were still skewed for most of the variables. Scatterplots using log transformed nutrient data (Fig. 6) show no significant trend in correlations of O-P or TP with Chl *a* concentrations. Therefore we decided to run a Spearman's rank-order correlation, to investigate if nutrients showed a significant influence on biomass concentrations.

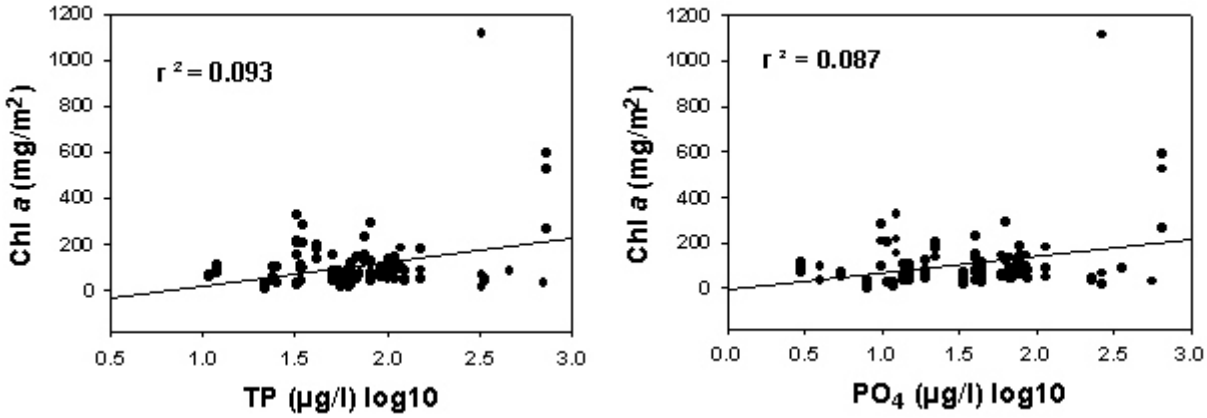

Figure 6: Correlations of PO_4 and TP with 3 Chl *a* concentrations. All samples from 2000 and 2001 are included. PO_4 and TP values are log transformed.

The variables in the Spearman's rank-order correlation included all measures of algal biomass (Chl *a* and AFDM data, RBA visual estimate and semi-quantitative count procedure) for all 106 samples collected during both years, and all nutrient measurements (TP, O-P, NH_3-N,

NO_3-N). For explanation of the different variables included in the correlation, and results of the correlation matrix, see Appendix 3. We used the following abbreviations to identify the method or analysis from which the data were derived: Rapid bioassessment (RBA) and semi-quantitative count method (SQCM).

5.2.1.1 *Comparison between results of different biomass measures/estimates (Chl a, AFDM , visual estimate (RBA) and % biomass estimate (semi-quantitative count method)*

The following correlations are significant at the 0.01 level using Spearman's rank-order correlation, two-tailed test. Chl *a* is significantly correlated with % visually estimated *Cladophora* sp. cover (RBA) (r= 0.40), with visually estimated % *Cladophora* sp. cover multiplied by its length rank (RBA) r = 0.41), and with visually estimated % blue-green cover multiplied by its length rank (RBA) r = 0.26). In contrast, AFDM is not significantly correlated at the 0.01 level (two-tailed) with any % biomass estimate. Correlations at the 0.05 level are more frequent but less strong showing the following results: Chl *a* correlates with estimated % *Cladophora* sp. biomass (SQCM) r =0.24), with % estimate blue-green algae cover (RBA) (r=0.25) and with estimated % cover of green algae (RBA) r =0.21). AFDM is correlated (at the 0.05 level, two-tailed) with estimated % *Oegodonium* sp. biomass (SQCM) (r=0.22), estimated % biomass green filamentous algae (RBA) (r= 0.19), and estimated % biomass green filamentous algae multiplied by its maximum length rank (RBA) r =0.19). AFDM is negatively correlated with diatom estimated biomass (SQCM) r = -0.20), estimate of % thin layer of diatoms (RBA) (r= -0.19) and estimate of % thin layer of diatoms multiplied by their thickness rank (RBA) (r= -0.23).

In summary, the results of the Spearman's rank-order correlation show that both, % estimate of *Cladophora* sp. biomass (RBA) and % estimate of *Cladophora* sp. biomass multiplied by its length rank (RBA), and visually estimated % blue-green cover multiplied by its length rank (RBA) are the two groups that are correlated strongest and most significantly (at the 0.01 level) with Chl *a*. AFDM correlates with mainly *Oegodonium* sp. biomass (RBA) and cell counts (SQCM), and green filamentous algae thickness and maximum length (RBA), but the correlations are weaker and less significant. In general, biomass measures (Chl *a* and AFDM) show stronger correlations with the results of the RBA than with the semi-quantitative count method. This study shows that the Rapid Bioassessment method is a good tool to estimate biomass impairment in rivers of NJ, and especially seems to reflect well extreme growths of *Cladophora* sp. Nevertheless, besides the correlation with *Cladophora* sp., none of the correlations is very strong and interpretations should be made with caution.

5.2.1.2 *Relationships between algal biomass measures and nutrient conditions*

We explored the relationships between algal biomass measures and nutrient conditions using Spearman's rank-order correlation, two-tailed test (Appendix 3). The only correlation that was significant at the 0.01 level was a positive relationship between AFDM and nitrate (NO_3-N) r = 0.26), and a negative relationship between estimated % blue-green cover (RBA) r =-0.31) and estimated % blue-green cover multiplied by its thickness rank (RBA) r =-0.29) and NH_4-N. In summary, except for AFDM and NO_3-N, we did not find significant trends or strong positive relationships between amount of Chl *a*, AFDM, visual biomass estimate and nutrient

measurements. The results of the data analysis performed during the first two years of our study reveal that nutrient concentrations measured in NJ Piedmont rivers do not show strong and significant correlations with any of the different biomass measures. However, the results of forward stepwise regression (section 5.2.2.) suggest that if the influence of river width (light conditions) and substrate are accounted for, nutrient concentrations will have a stronger relationship with algal biomass. We will perform detailed analysis of a bigger dataset (including year 3 data from this study) to investigate this relationship further.

5.2.2 Analysis of principal factors influencing algal biomass (forward stepwise regression)

We analyzed algal biomass (AFDM an Chl *a*) and its relationship with nutrients (TP, O-P, NH_3-N, NO_3-N), other chemical variables (dissolved oxygen, pH, conductivity), geomorphic variables (river basin size, river width, section length, percent type of substrate) and light conditions (percent open canopy cover) with Forward Stepwise regression using Sigma Stat version 2.03. Table 4 summarizes the results of the regression. The full results of the regression are attached in Appendix 4. The analysis was run twice, once each with either Chl *a* or AFDM as dependent variables. The results show that the dependent variables Chl *a* and AFDM can both be predicted from a linear combination of the independent variables NO_3-N and river basin size. In the case of Chl *a* only, size of substrate (sum of percent bedrock, boulder, cobble and gravel) had a significant influence on algal biomass. The correlations are significant at the 0.001 level for NO_3-N in both regressions, indicating that this variable shows the strongest influence. In the regression with Chl *a* as the dependent variable, NO_3-N is strongly correlated with TP and O-P, whereas in the second regression with AFDM as dependent variable, NO_3-N is independently having the strongest influence (strongest F-value of 18.62) in the dataset. In both regressions, basin size is strongly correlated with percent open canopy cover and average river width and section length (see results in Appendix 4). Basin size is correlated with light, river with, and section length and is therefore an indirect variable expressing light conditions. This shows that in our analysis, a bigger river basin reflects a wider river, with more light reaching the river bottom and therefore causing higher algal biomass.

5.3 <u>Algal flora- species composition</u>

5.3.1 Composition of soft-algae flora in biomass samples (soft-algae flora)

The soft algal flora is composed mainly of *Cladophora* sp. and *Audouinella* sp. Other algal groups like *Oscillatoria* sp., *Oegodonium* sp., *Rhizoclonium*, *Spirogyra* sp. and *Merismopedia* sp. are represented in much lower abundances and lower number of occurrences (see Table 5). The percentage estimates (or proportions) of the most common species of algae, identified though the semi-quantitative analysis (see section 4.4), showed the following composition (Table 5). The assemblages were strongly dominated by diatoms throughout the whole dataset. The second most important groups were *Cladophora* sp. and *Audouinella* sp. with *n* of 11 and 12 and median percentages of estimated biomass of 18 and 20 respectively. Finally *Oscillatoria* sp., *Oegodonium* sp., and *Rhizoclonium* sp. occurred less often (n = 2 to 4) but with relatively high medians. The least common groups, *Spirogyra* sp. and *Merismopedia* sp., were

Table 4: Variables significantly influencing algal biomass, as determined by Forward Stepwise regression.

Dependent Variable	Step	Variables Entered	F- to enter	*p*	*r*	r^2
Chl *a*	1	Basin size	31.019	<0.001	0.479	0.23
	2	NO_3-N	9.104	<0.001	0.541	0.292
	3	bigger substrate	10.18	0.002	0.597	0.357
AFDM	1	NO_3-N	18.62	<0.001	0.39	0.152
	2	Basin size	5.519	0.021	0.442	0.195

Table 5: Percentage estimates of the most common species of algae that make up the largest proportion of algal cells and algal biomass.

Algal Group	*n* (samples)	Max	Min	Mean	Median
Diatoms (% # cells)	106	85	0.5	83.0	85.0
Diatoms (%biomass)	106	100	60	95.2	100.0
Cladophora sp.(% # cells)	12	85	3	18.0	12.5
Cladophora sp.(%biomass)	12	40	4	16.1	18.0
Audouinella sp.(% # cells)	11	30	3	12.2	12.5
Audouinella sp.(%biomass)	11	40	4	18.6	20.0
Oscillatoria sp.(% # cells)	4	3	3	3.0	3.0
Oscillatoria sp.(%biomass)	4	4	4	4.0	4.0
Oedogonium sp.(% # cells)	3	12.5	0.5	5.3	3.0
Oedogonium sp.(%biomass)	3	19	1	8.0	4.0
Rhizoclonium sp.(% # cells)	2	12.5	3	7.8	7.8
Rhizoclonium sp.(%biomass)	2	20	3	11.5	11.5
Scenedesmus sp.(% # cells)	1	0.5	0.5	0.5	0.5
Scenedesmus sp.(%biomass)	1	1	1	1.0	1.0
Spirogyra sp.(% # cells)	1	12.5	12.5	12.5	12.5
Spirogyra sp.(%biomass)	1	18	18	18.0	18.0
Merismopedia sp.(% # cells)	1	0.5	0.5	0.5	0.5
Merismopedia sp.(%biomass)	1	1	1	1.0	1

observed in samples from one site each. Despite its low occurrence, *Spirogyra* sp. was estimated to contribute up to 18% of the estimate of biomass, in contrast to *Merismopedia* sp., with 0.5% estimated biomass.

5.3.2 Principal patterns in the variation of diatom assemblage composition (DCA)

The diatom flora is composed of 306 taxa (Appendix 2) dominated by pollution-tolerant species. The 10 most abundant species, determined by high abundances and high numbers of occurrences (see Appendix 2), are *Navicula minima* Grun., *Rhoicosphenia curvata*, (Kütz.) Grun. ex Rabh, *Nitzschia inconspicua* Grun., *Planothidium frequentissimum* (L-B) Round & Bukht., *Nitzschia amphibia* Grun., *Sellaphora seminulum* (Grun.) Mann, *Melosira varians* Ag., *Cocconeis placentula* var. *lineata* (Ehr.) V. H., *Navicula lanceolata* (Ag.) Ehr. and *Navicula gregaria* Donk.

The DCA analysis of species served to measure the maximum amount of variation in the diatom data, and also to help identify outliers (see section 4.6.3.1). Figure 7 shows samples with sample scores falling outside the 95% confidence limit about the sample score means on axis 1 and 2 (Hall and Smol 1992): site AN0115 section 1, 2 and 3, site AN0439 section 1, site AN0227 section 1, 2 and 3 and site AN0318 section 1, 2 and 3.

5.3.3 Relationship between species composition and environmental variables, especially nutrients

5.3.3.1 *Soft algae: Spearman's rank-order correlation*

Relationships among the semi-quantitative algal counts and nutrients and other environmental variables were examined by running a second Spearman's rank-order correlation (see section 4.6.2.1). Detailed results are given in Appendix 3. The following correlations were significant: the estimated number of *Cladophora* sp. cells correlate at the 0.05 level with Chl *a* r =0.25), average width of the river r =22) and sampling section length r =0.23). Furthermore, the estimated number of *Cladophora* sp. cells correlated significantly (at the 0.01 level) with dissolved oxygen r =0.25), pH r =0.33) and % open canopy (=light) r =0.29). For diatom cells, estimated numbers correlate significantly (at the 0.01 level) with amount of Chl *a* r =0.22) , the average width of the river r =27) and section length r =0.27), and at the (0.05 level) with % open canopy (=light) r =0.21). Furthermore, the estimated number of cells of *Oegodonium* sp. (r=0.22) is correlated significantly with AFDM (at the 0.05 level). Estimated number of cells of *Audouinella* sp. r =0.22) is correlated at the 0.05 level with NH_3-N.

In summary, the strongest and most significant correlations between soft algal species composition and environmental variables were found between abundance of *Cladophora* sp. cells and pH and % open canopy, and also between abundance of diatom cells and average width of the river and section length. Correlation between abundance of *Cladophora* sp. and pH might express high rates of photosynthesis, which influence the pH conditions in the water column, but could also be related to a preference by *Cladophora* sp for pH-neutral waters. Therefore, overall light conditions seem to have strongest influence on abundance of diatoms and *Cladophora* sp.

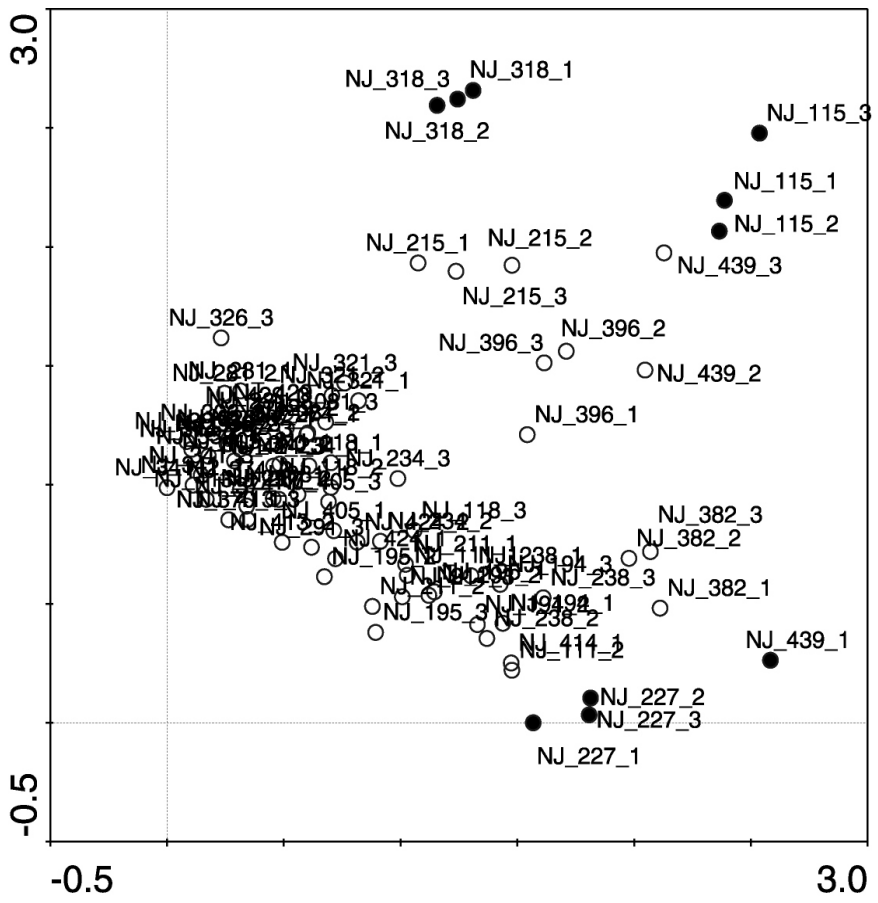

Figure 7: DCA showing site scores. Samples indicated with filled circles were identified as outliers.

Nevertheless, all correlations found are rather weak. Finally, except for *Audouinella* sp., which is correlated at the 0.05 level with NH_3-N, no significant correlations were found between soft algal species composition and nutrients.

5.3.3.2 *Diatoms: ordination analysis; influence of environmental variables on diatom species composition*

Development of nutrient inference models requires a strong statistical relationship between diatom species composition and the variable to be modeled (Winter and Duthie 2000, Birks et al. 1990a). Therefore, we identified the variables that explained a significant amount of variation in diatom species composition, running CCAs constrained to each variable separately (see section 4.6.3.3.). The λ_1/λ_2 ratio was high for TP (0.467) and O-P (0.474). These variables were therefore determined to have strong influence on diatom species distribution (Winter and Duthie 2000). The λ_1/λ_2 ratios for conductivity (0.331), NO_3-N (0.298), NH_3-N (0.264) and pH (0.263) showed some influence, but were weaker. Monte Carlo Permutation tests (199 permutations)

revealed that significant (p<0.05) amounts of the variability in diatom assemblage composition are explained by all measured inorganic nutrients (TP, O-P and NO_3-N, NH_3-N). A final CCA was produced to show the strength of the influence of each variable mentioned above on diatom species composition (Fig. 8).

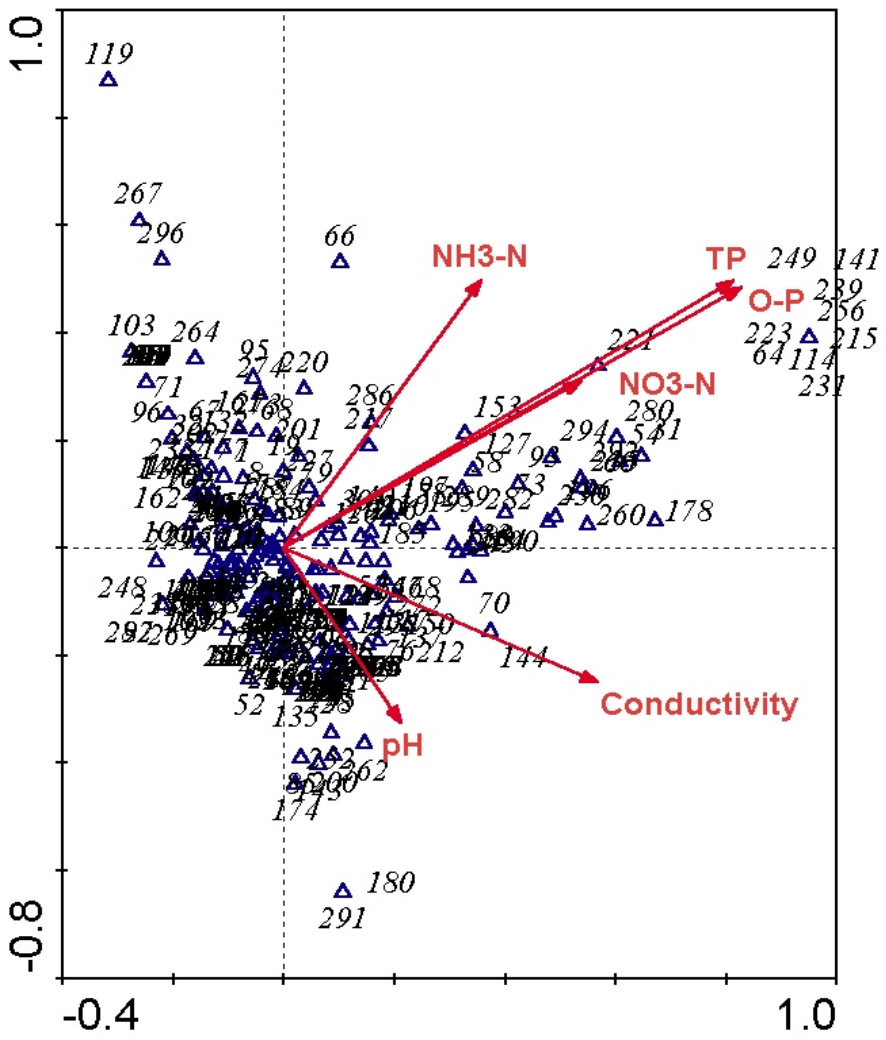

Figure 8: CCA of diatom assemblages, including 85 samples (outliers included) and 6 environmental variables having strong influence on the 306 the species included. Triangles represent samples and numbers represent species names as listed in Appendix 2.

5.4 Development of nutrient inference models based on diatom species composition

5.4.1 Data screening

In order to select the variables that have independent and significant influence on species composition, we ran a series of specialized analyses. The purpose of these analyses was to screen the data for unusual samples, and to remove those from the dataset to be used for development of models. Unusual samples ("rogues" or "outliers") were defined as samples that have either unusual diatom assemblages that are weakly related to the variable to be reconstructed, that have an unusual combination of environmental variables, or that have environmental variables that have a stronger influence than the variable to be reconstructed (Birks et al. 1990b, Hall and Smol 1992).

Because they were taken from sandy substrate, the following samples were deleted from the dataset used for diatom inference models: AN194, AN227, AN0238, AN0382 and AN0414. Furthermore, all samples from the five sites that were not located in the Piedmont were excluded from the dataset (AN0215, AN0318 and AN0321, N0382, AN0439). Due to extreme environmental variables and species scores, as previously described, the samples coming from site AN0291 sections 1, 2 and 3, from site AN0118 sections1, 2 and 3 and from site AN0115 section 1, 2 and 3 were deleted from the dataset used for inference models (see sections 5.1.2 and 5.3.2). Based on the above data screening process, 37 diatom samples were deleted from the original dataset. The final training set to be used for development of inference models contained 54 out of the 85 diatom samples, all collected in 2000. Nevertheless, as the screened dataset was reduced by nearly 50%, we decided to use both datasets, the full dataset (n= 85) and the reduced dataset (n= 54), as training sets for comparison of development of inference models.

5.4.2 Weighted averaging - nutrient inference models

In order to develop nutrient inference models, a strong statistical relationship between diatom species composition and the variable to be modeled is required (Winter and Duthie 2000, Birks et al. 1990a). As identified through CCAs constrained to each variable separately, we determined that O-P and TP have strong influence and that NO_3-N and NH_3-N have moderately strong influence on diatom species distribution. Based on the results of the Monte Carlo permutation tests, development of nutrient inference models was possible for all four variables (see section 5.3.3.2) . We developed inference models for the nutrient variables O-P, TP, NH_3-N and NO_3-N using weighted averaging regression and calibration on 2 different datasets: the first set included all 85 samples, and all species; the second set included only 54 samples (see section 4.5.2). The results indicate that all models have relatively high predictive power, and that the root mean square errors of prediction are relatively low (see Table 6). The two best models developed using the full dataset (n=84) are: the TP inference model (n=84), with an apparent r^2 of 0.72 and a RMSEP $_{(boot)}$ (log) of 0.33 µg/L and the NO_3-N inference model (n=84) with an apparent r^2 of 0.68 and an RMSEP $_{(boot)}$ (log) of 0.26 µg/L. The two best models developed using the reduced dataset (n=54) are: the TP inference model (n=54), with an apparent r^2 of 0.69 and a RMSEP $_{(boot)}$ (log) of 0.22 µg/L and the NO_3-N inference model (n=54) with an apparent r^2 of 0.64 and an RMSEP $_{(boot)}$ (log) of 0.21 µg/L.

Table 6: Predictive power of diatom inference models for TP, O-P NH_3-N and NO_3-N, as determined using WA-regression and calibration.

	***n*= 85**		***n*= 54**	
Parameter	r^2 **(apparent)**	**RMSEp$_{(boot)}$ (log) µg/L**	r^2 **(apparent)**	**RMSEp$_{(boot)}$ (log) µg/L**
TP	0.72	0.33	0.69	0.22
O-P	0.69	0.42	0.67	0.31
NH_3-N	0.71	0.36	0.68	0.34
NO_3-N	0.68	0.26	0.64	0.21

5.4.3 Evaluation of the performance of the TP model

The TP model (*n*=85) has relatively high predictive power $r^2_{(boot)}$= 0.55) (Fig. 9).

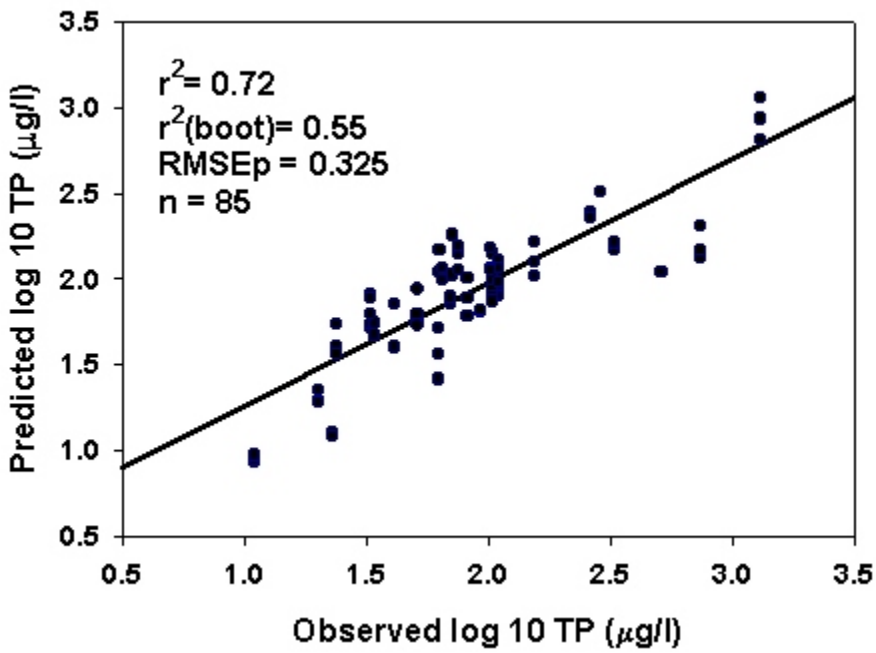

Figure 9: Observed versus predicted TP for the WA inference model developed based on 85 diatom samples collected in 2000.

5.5 Evaluation of diatom metrics, indices and inference models

Several different diatom indices and inference models were applied to the NJ Piedmont dataset to assess weather any of the existing indices could produce reliable results when applied to NJ river diatoms. The different indices and the results obtained are described below.

5.5.1 Test of the NJ Piedmont nutrient inference model on the year 2 samples

To evaluate the performance of the TP model developed, it was tested on the full year 2 dataset (12 sites) including the 5 duplicate sites AN0374, AN0274, AN0115, AN0211 and AN0405. To run the test, we performed WA-calibrations using CALIBRATE version 0.61 (Juggins and ter Braak 1997, 2001) and applied the model developed using sites sampled in 2000 to the samples collected in 2001. The performance of the TP model was assessed by evaluating the distribution of the observed versus inferred values (Birks et al. 1990b). With a correlation coefficient of 0.78 (r^2= 0.61) our test showed good results (Fig. 10). This analysis demonstrates that the developed TP model could be applied successfully to other diatom samples collected in the rivers of the NJ Piedmont to reliably predict TP concentrations.

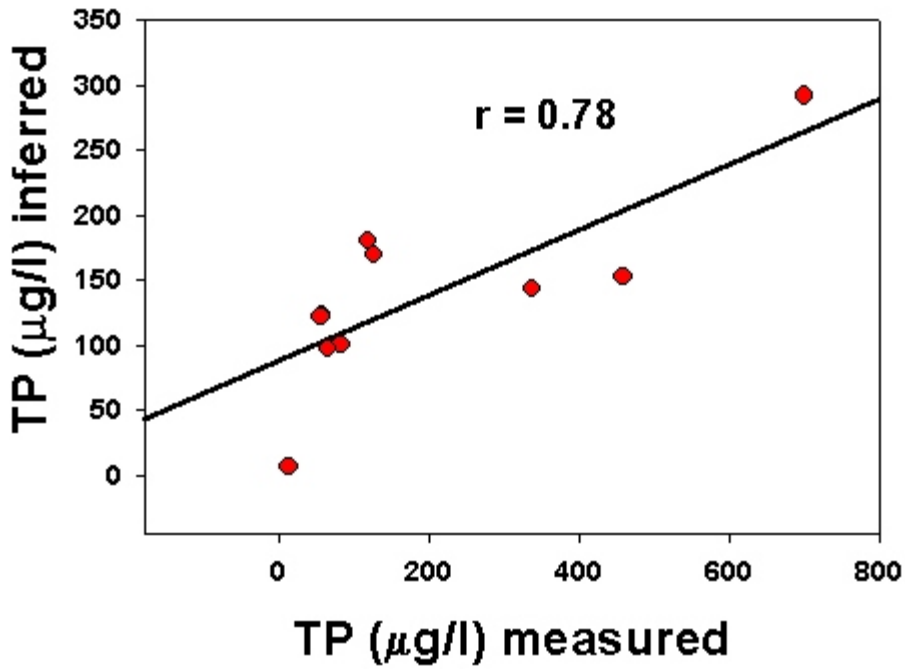

Figure 10: Test of TP inference model: plot of measured (samples taken in 2001) versus diatom- inferred TP (TP model based on samples taken in 2000).

5.5.2 Diversity metrics and other simple metrics

Six different diatom diversity indices and other simple metrics were calculated based on 98 diatom samples collected in 2000 and 2001. A Spearman's rank-order correlation was run to evaluate how strongly these indices were correlated with different environmental variables expressing river impairment, e.g. types of landuse, nutrient and biomass concentrations. The main goal of this analysis was to identify metrics that can be used to assess nutrient impairment in NJ Piedmont rivers. The main results are shown in Table 7; the different types of metrics are explained in section 4.6.4.1. Considering the indirect relationship between the metrics and the variables they were correlated with, we consider that any correlations with an r greater than 0.4 are relatively strong, that correlations between $r = 0.2$ and 0.4 are moderate and that any correlations below an r of 0.2 are weak. The following indices showed significant (at 0.01 level) and strong correlations. Number of taxa was strongly correlated with NO_3-N and TP, and moderately correlated with basin size and NH_3-N and O-P. S-W diversity was moderately correlated with basin size, O-P and TP. Centrics/Pennates is strongly correlated with O-P and T-P and moderately correlated with basin size, NO_3-N and NH_3-N and % urban landuse. The Siltation Index is strongly correlated with basin size, O-P and T-P and moderately correlated with Chl *a.* In summary, the Siltation Index, the Centrics/Pennates and number of taxa, showed the strongest correlations with basin size, NO_3-N, O-P and TP ($p < 0.01$). Therefore, these diatom indicators could be used to monitor river impairment, especially of the nutrients NO_3-N, O-P and TP.

5.5.3 European indices (TDI, IBD and IPS)

The results of the Trophic Diatom Index (TDI), the Biological Diatom Index (IBD) and the Specific Polluosensitivity index (IPS) were calculated for samples taken in 2000 and 2001 and the results were tested for correlation with nutrient measurements. The European diatom flora used for the development of the indices differs from the NJ flora in species composition, hence we could only include 80% of the diatom species contained in our counts in the calculation of the European diatom indices. In particular, three North American species, *Gomphonema kobayassii*, *Gomphonema patrickii* and *Achnanthes* sp.1 were abundant and reached high numbers of occurrences in our dataset, but are not included in the European index. We used a Pearson's product-moment correlation matrix to assess how well the different indices reflect nutrient impairment (O-P, TP, Chl a, AFDM , % Urban, % Agriculture) in NJ Piedmont rivers (Table 8). The strongest correlation r =-0.65) was obtained for the IPS versus measured O-P, also presented in a scatterplot (Fig. 11). Because the TDI was developed mainly to reflect trophic conditions, it shows strong correlation with O-P r =0.64) and TP r =0.54), but a rather weak correlation with NO_3-N r =0.27) and NH_3-N r =0.08).

The IPS and the IBD were both developed to reflect overall impairment conditions, using the same approach. The difference between the two indices is that the IPS is based on a bigger dataset of diatom species (Prygiel and Coste 1999). When comparing the results of both indices, we found that the IPS shows stronger correlation with O-P and TP and NH_3-N but the IBD showed slightly stronger correlation with NO_3-N. Figure 12 shows that, in general, the IBD gives higher ratings for NJ river quality than the IPS.

Table 7: Spearman's rank-order correlation between diatom metrics and different variables expressing nutrient impairment. For explanation of metrics see section 4.6.4.1. p-value:** significant at 0.01 level and *significant at 0.05 level.

Diatom Metric/ Environmen-tal Variable	# Taxa	S-W Index	% Dominants	% Dominant Taxon	C/P	Siltation Index	
% Urban	0.049 0.629	-0.019 0.852	-0.021 0.837	-0.002 0.983	0.270** 0.007	0.158 0.120	***r*** ***p*-value**
% Agriculture	-0.056 0.585	0.004 0.970	0.002 0.985	-0.052 0.613	-0.176 0.0821	0.018 0.862	***r*** ***p*-value**
% Forest	0.006 0.951	0.091 0.374	-0.059 0.571	-0.167 0.099	-0.050 0.624	0.110 0.280	***r*** ***p*-value**
Basin size	0.324** 0.0012	0.306** 0.002	-0.259** 0.0011	-0.240* 0.0176	0.344** 0.000	0.413** 0.000	***r*** ***p*-value**
NO_3-N	0.423** 0.000	0.319** 0.002	-0.250* 0.014	-0.138 0.175	0.222* 0.028	0.168 0.099	***r*** ***p*-value**
NH_3-N	0.280** 0.005	0.0872 0.393	-0.066 0.523	0.0308 0.763	0.395** 0.000	0.017 0.866	***r*** ***p*-value**
O-P	0.366** 0.000	0.333** 0.000	-0.244* 0.017	-0.340 ** 0.000	0.431** 0.000	0.445** 0.000	***r*** ***p*-value**
TP	0.403** 0.000	0.326** 0.001	-0.235* 0.022	-0.326** 0.001	0.482** 0.000	0.449** 0.000	***r*** ***p*-value**
Chl *a*	-0.057 0.609	0.068 0.538	-0.087 0.437	-0.117 0.286	0.112 0.309	0.471** 0.000	***r*** ***p*-value**
AFDM	-0.001 0.991	-0.012 0.911	0.001 0.953	0.055 0.619	0.141 0.200	0.215 0.050	***r*** ***p*-value**

Table 8: Pearson's product-moment correlation matrix comparing European diatom indices with different measures of nutrient impairment.

	NO_3-N	**NH_3-N**	**O-P**	**TP**	**Chl *a***	**AFDM**	**% URB**	**%AGR**
TDI	0.27	0.1	0.64	0.54	0.58	0.17	0.24	-0.3
IPS	-0.3	-0.3	-0.65	-0.6	-0.5	-0.3	-0.4	0.21
IBD	-0.46	-0.17	-0.55	-0.48	-0.55	-0.35	-0.4	0.24

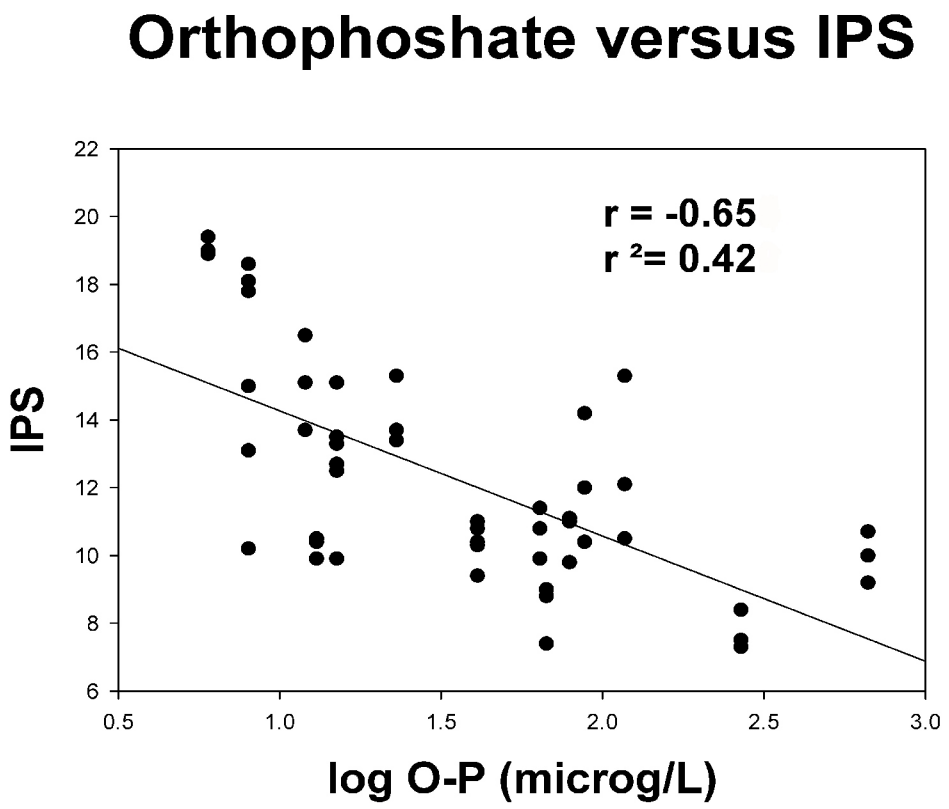

Figure 11: Scatterplot of measured O-P versus the indices calculated by Specific Polluosensitivity index (IPS) for all 2000 and 2001 samples.

In summary, the three European diatom indices were good predictors of orthophosphate and total phosphorus. Nevertheless, these results should be interpreted with caution due to differences between the European diatom flora used for the development of the indices and the diatom flora in NJ. Overall the results show that there is potential for expanding the existing European diatom models by including data for North American species.

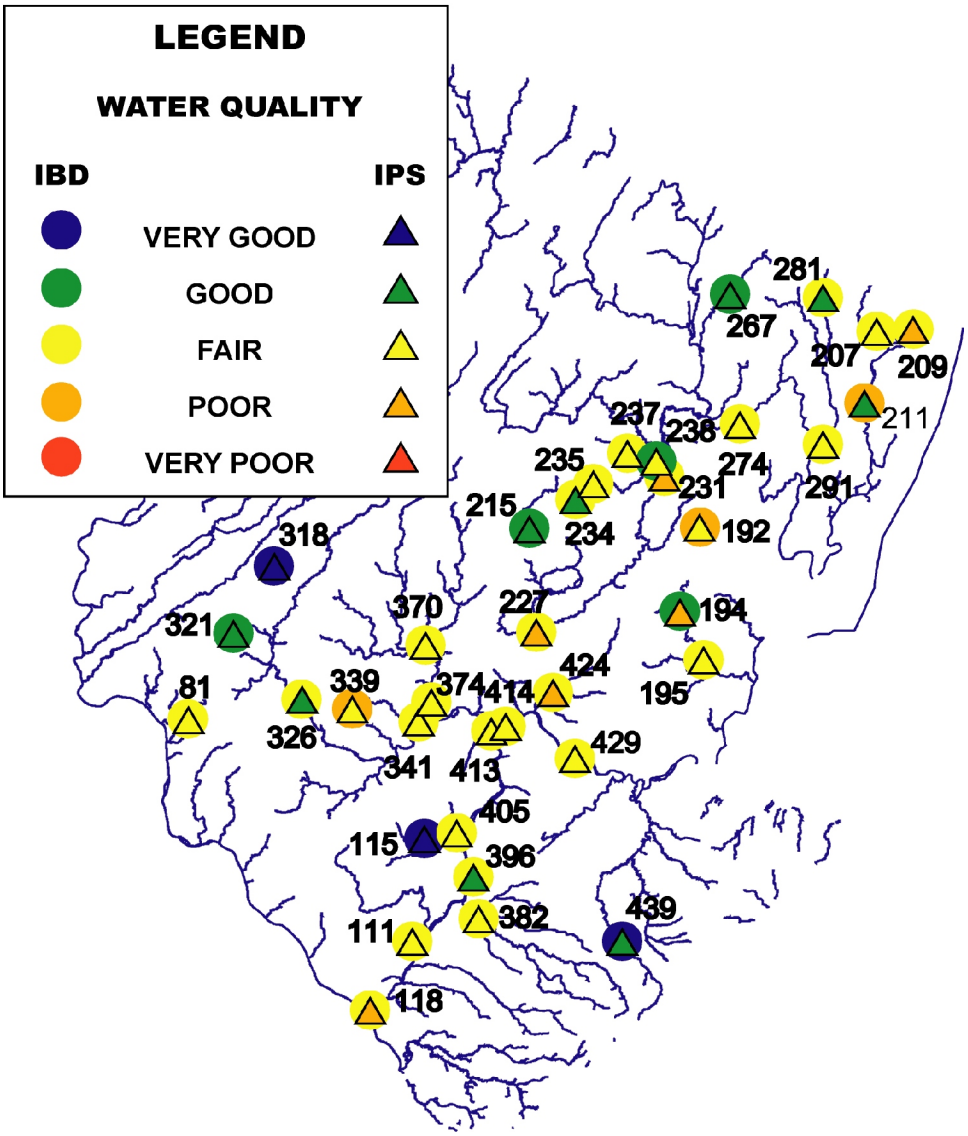

Figure 12: Map showing the difference in the ratings of river quality as calculated by two European diatom indices, the Specific Polluosensitivity index (IPS) and the Biological Diatom Index (IBD). The numbers correspond to NJ site ID's (see Tables 1 and 2).

6 Discussion

A main goal of this study was to explore the relationships among algal biomass, algal species composition and nutrients. A further goal was to identify the most promising indicators for assessing excess nutrients in NJ rivers and streams using biomass and algal species composition (soft algae and diatoms). In the following paragraphs we summarize nutrient-algal relationships identified from this study and discuss the outcome of all different methods used. Finally, we discuss applicability of the indicators developed and provide recommendations towards their use and further development.

6.1 Principal factors influencing algal biomass

6.1.1 Principal variables influencing algal biomass

A multitude of recent studies have been conducted to understand which combination of factors determines algal biomass. A large amount of literature is available on this subject and a comprehensive review is published in a report presented by the New England Interstate Water Pollution Control Commission (ENSR 2001). The main factors influencing biomass accrual are nutrients, light, temperature, substrate availability and stream velocity (Biggs 1996). To determine the principal variables influencing algal biomass in our study, we analyzed all variables measured in the field and in the lab using forward stepwise regression. All factors mentioned above were included in this analysis, except for temperature measurements. To include temperature measurements we would need a long-term record of average daily temperatures for weeks before each sampling date. No such detailed record was available for most sites sampled. The results of a stepwise regression showed that the dependent variables Chl *a* and AFDM can both be predicted from a linear combination of the independent variables NO_3-N and river basin size. In the case of Chl *a* only, bigger substrate size (sum of percent bedrock, boulder, cobble and gravel) also has a significant influence on algal biomass. These results show that algal biomass in the NJ dataset is influenced by a combination of light conditions (reflected through basin size) nitrogen (NO_3-N) and factors closely associated with these. In our dataset TP and O-P are strongly correlated with NO_3-N, therefore correlation of Chl *a* with NO_3-N also reflects correlation with phosphorus. Also, in the case of Chl *a* , the size of the type of substrate, and the availability of algae to attach is an additional important factor. In summary, we can deduce from this analysis that in the NJ Piedmont rivers a combination of three factors together – high light levels, high nutrient concentration, and high proportion of larger-sized substrate – lead to the greatest quantities of algal biomass in NJ Piedmont rivers. These results suggest that there is potential for development of a metric for biomass combining all of these factors. We will explore the possibility of developing such a metric further, especially in combination with the third year data of this study.

6.1.2 Nutrient-biomass relationships as assessed by correlations

After determining the main factors influencing algal biomass, we investigated the strength of the relationship of individual nutrients on different measures of biomass. Spearman's correlation analyses showed that there is no significant and strong relationship between measured biomass concentrations of Chl *a* , AFDM or estimate of biomass as assessed through the RBA with the nutrients TP, O-P, NO_3-N and NH_3-N. Only the correlation between AFDM and nitrate

(NO_3-N) shows a significant trend, but the correlation is weak $r = 0.26$). As shown in section 6.1.1, biomass can only be explained through a combination of factors including nutrients. There is need for more detailed analysis of this dataset including the larger dataset with year 3 samples to explore the strength of the relationship between biomass and nutrients. A recent study on large datasets (national and international) on temperate streams revealed that a significant portion of variance in annual mean and maximum biomass can be explained by total nitrogen and total phosphorus concentrations (Dodds et al. 2002). The same study also shows that such relationships are very weak at the regional scale. Our study is consistent with this observation, and demonstrates the challenges of finding clear relationships at smaller (regional) scales, such as the NJ Piedmont. It demonstrates that a strong correlation between nutrient and biomass is at best difficult to establish, and that other factors such as light (as a function of river basin size) and substrate must be taken into account when estimating nutrient-biomass relationships (see section6.1.1). Therefore, commonly used biomass measures (Chl *a*, AFDM) must be interpreted with caution, and inferences of nutrient levels in rivers based on these measures should be made only in conjunction with analyzing other variables.

6.2 Comparison and evaluation of methods for estimating algal biomass

Algal biomass was measured in three different ways: a) we measured the contents of Chl *a* and AFDM contained in the composite biomass samples collected in the field; b) we estimated the proportion of cells making up most of the biomass in the composite biomass samples, using a specially designed semi-quantitative analysis and finally c) we visually estimated proportions of abundance and thickness of algal cover in the rivers as assessed through the Rapid Bioassessment (RBA). Spearman's rank-order correlation between the different measures of biomass shows significant correlations between Chl *a* and with RBA estimates of biomass, with the strongest correlation between Chl *a* and *Cladophora* sp.($r = 40$). In contrast, both the measures of AFDM and the estimates of biomass through the semi-quantitative analysis do not show significant and strong correlations with any other measures of biomass. Therefore, we do not recommend the use of the semi-quantitative analysis method for estimating biomass. Also, when given the choice between measuring either Chl *a* or AFDM as a variable to be used for biomass-algal group relationships, Chl *a* should be given priority.

Our study shows that a combination of measuring Chl *a* from composite diatom samples and using the RBA method seems to assess the amount of algal biomass best. When using those two approaches in combination, a good assessment of the biomass impairment in rivers of the NJ Piedmont can be achieved. *Cladophora* sp. is, based on our results, the species that correlates most closely with Chl *a* and is therefore the algal group that needs to be monitored most. The assessment through the RBA in combination with measuring Chl *a* from composite biomass samples provides a good tool for monitoring growth of *Cladophora* sp. Nevertheless, none of the correlations found between the different measures of biomass is very strong and therefore a combination of different methods should be used to assess biomass.

6.3 Comparison and evaluation of diatom metrics and models

In the following sections we compare the results of the diatom metrics and models with the objective of determining the best method for estimating phosphorus concentrations and overall water quality in NJ Piedmont rivers. The relative advantages and disadvantages of each method are discussed. Performance of all are potentially limited by how well the environmental measurements, especially nutrient concentrations, represent the variability of conditions to which the algae are exposed. It would be useful for evaluating metric effectiveness if the variability of these conditions could be quantified.

6.3.1 Nutrient-inference models

The diatom species composition found in the NJ Piedmont dataset was strongly influenced by the measured nutrient variables (O-P, TP, NO_3-N and NH4-N). Therefore, development of WA - inference models for nutrients was possible. We developed 4 different nutrient inference models based on the full (n=85) and a reduced dataset (n= 54). All models have relatively high predictive power. The best models developed are the TP and the NO_3-N inference models for the full (n=85) and a reduced dataset (n= 54). For both variables, the model developed with the full dataset (n=85) has a higher r^2, but the model based on the reduced dataset (n= 54) has a lower $RMSEP_{(boot)}$, respectively. This means that the errors obtained for the values inferred using the TP and NO_3-N models developed for the full dataset will be higher than those based on the reduced dataset. Nevertheless, when we tested observed versus predicted TP for the full dataset inference model (n=85), we found that the model has relatively high predictive power $r^2_{(boot)} = 0.55$. Also, testing of the same model on the year 2 samples produced reliable results ($r^2 = 0.61$). In the future, different techniques will be applied to improve the model to increase its predictive power. Also, we plan on including samples taken during the second (2001) and the third year (2002) of this project to increase the predictive power of these two WA-inference models for NO_3-N and TP.

In summary, in contrast to soft algal species composition, we found good correlation between diatom assemblage composition and nutrients. The WA nutrient-inference models developed showed reliable results when tested, and we intend to improve them further. The diatom inference models seem to be better indicators for nutrients and river eutrophication than biomass. In general, species composition based indicators correlate better with nutrient concentrations, because the ecological information from diatoms is not as variable as biomass (e.g., Chl *a*) concentrations over time. The application of diatom inference models to NJ Piedmont rivers and streams is highly recommended as a tool for monitoring eutrophication. Nutrient inference models are relatively easy to apply as a regular monitoring tool. Nevertheless, their use requires appropriate software and expertise.

6.3.2 Simple metrics

The usefulness of the of the six diversity and simple metrics was evaluated by comparing their correlations with nutrient impairment measures. Siltation Index, the Centrics/Pennates and number of taxa index, showed moderate, but significant correlations with NO_3-N, O-P and TP. Therefore, it is possible to use these diatom indicators to monitor river impairment, especially of

the nutrients NO_3-N, O-P and TP. The formulas for the indices are simple and calculation is easy. They could be included in the list of metrics in the EDAS database and analysis system used by the NJ DEP. Nevertheless the correlations obtained between the indices and the nutrient variables are not very strong and the significance of the results is questionable. We consider the results obtained through the nutrient inference models by far more reliable and recommend using simple metrics only as complementary method.

6.3.3 European indices

The three European diatom indices (TDI, IBD, IPS) showed good results and relatively strong correlation with orthophosphate and total phosphorus in the NJ Piedmont dataset. It is interesting to note that the strength of the correlation of the Northern Piedmont Diatom TP index and the British Trophic Diatom Index with Orthophosphate is similar $r = 0.66$ and $r = 0.64$ respectively). Nevertheless, we do not recommend using the European indices solely, because of important differences between the European diatom flora used for the development of the indices and the diatom flora in the NJ Piedmont.

6.4 Comparison of diatom inferred TP and impairment classifications based on macroinvertebrate metrics (AMNET)

We compared TP calculated using the diatom TP diatom inference model (n=85) to macroinvertebrate impairment ratings based on the 1998/99 assessment. The results (Fig. 13) show that the diatom inferred TP values do not correspond strongly to the macroinvertebrate impairment groups of "non-impaired," "moderately impaired" and " severely impaired." No significant separation is found between ratings, as all groups' 10th and 90th percentiles overlap to a high degree with the adjacent category. The"moderately impaired" category in particular includes a large number of sites that are indicated by the diatoms to have a wide range of TP values. Nevertheless, the sites rated "non-impaired" and "severely impaired" do show a trend of having lower and higher diatom inferred TP, respectively. More detailed analysis comparing diatom inferred nutrient concentrations with macroinvertebrate metrics that best reflect nutrient (e.g., % EPT etc.) will be performed in collaboration with the NJ Integrated Assessment (Horwitz and Flinders 2003) including Year 3 data of this study.

6.5 Evaluation of the EPA percentile method for determining reference conditions

We applied the proposed U.S. EPA percentile method (U.S. EPA 2000b) to the NJ Northern Piedmont dataset to calculate reference conditions. In our study, the 25th percentile was calculated using all nutrient data from all reaches (Appendix 1b). We compared our value to those for the aggregate of all Level III Subecoregions of Nutrient Ecoregion IX, and to the Northern Piedmont subecoregion (64) only, as given in the Ambient Water Quality Criteria Recommendations (U.S. EPA 2000b). The range of TP reference conditions given for all subecoregions is 22.5-100 µg/l; for the Northern Piedmont subecoregion it is 40 µg/L. We calculated a reference condition of 51 µg/L for the NJ Northern Piedmont, which is only 10 µg/L greater than the one proposed in the EPA document. The range of EPA total nitrogen reference conditions is 0.07-1.00 mg/L; it is 1.30 for the Northern Piedmont. Our dataset shows a

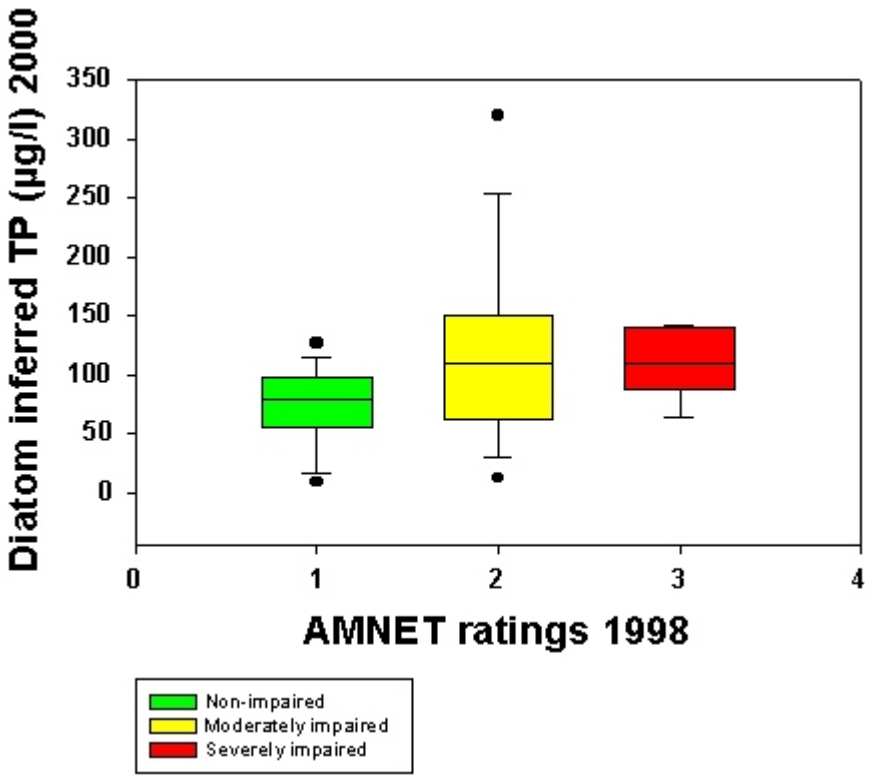

Figure 13: Box-plots comparing diatom inferred TP to AMNET macroinvertebrate impairment ratings. Upper limit of error bars indicate the 10th and the 90th percentile. Filled circles indicate outliers.

25th percentile of total nitrogen of 1.28 mg/L, which is relatively near EPA's value for the Northern Piedmont. Also, the periphyton chlorophyll *a* measured in our dataset shows a 25th percentile of 48.07 mg/m^2 which substantially exceeds the range given by EPA of 3.13-20.35 mg/m^2 and the value for the Northern Piedmont of 20.35. This comparison shows that, using the percentile method, there is reasonable good agreement between TP and TN reference values derived from data in the EPA study and this study, but that there is a substantial difference for Chl *a* values.

7 Conclusion: Recommendation for use of the ideal algal indicator monitoring program for the NJ Piedmont

Based on our study, we recommend the use of a combination of algal indicators and metrics for monitoring nutrients and biomass. For monitoring biomass, we recommend using the EPA Rapid bioassessment Protocol in combination with measuring chlorophyll *a*. Our results show that chlorophyll *a* correlates especially well with *Cladophora* sp. cover and that measuring both of these variables provides a good monitoring tool. Biomass can be explained by a combination of factors, such as nutrients and light conditions and there is potential for developing a biomass metric that incorporates these factors. More detailed analysis of our dataset is needed to develop such a metric, especially in combination with the third year data of this study.

Diatom assemblage composition is strongly influenced by nutrients, especially phosphorus (O-P and TP), we were therefore able to develop phosphorus inference models and indices. Inferred values and metrics were tested by comparing them with measured phosphorus values. The best results were achieved with the TP diatom inference models developed for the NJ Piedmont. We compared the results obtained for the European indices, the Biological Diatom Index, the Polluosensitivity Index and the Trophic Diatom index. All three indices showed relatively good correlations with either O-P and/or TP, suggesting that all three methods could potentially be applied and the results compared, when using diatoms as indicators of river phosphorus in the NJ Piedmont. The biggest limitation to further development of models and metrics is probably the representativeness of nutrient values that are based on very few samples per site. Increasing the number of samples per site is recommended for future studies.

In summary, this study shows that algae can be used as indicators of nutrient impairment for the NJ Piedmont. Diatoms especially show good response to nutrients and their use as monitoring tool is highly recommended. Biomass metrics need further analysis, the possibility of developing metrics combining different factors influencing algal growth, especially nutrients is promising.

References

American Public Health Association, American Water Works Association and Water Pollution Control Federation (APHA, AWWA and WPCF) 1992. Standard Methods for the Examination of Water and Wastewater, 19th ed. Washington, DC.

Bahls, L. L. 1993. Periphyton bioassessment methods for Montana streams. Montana Water Quality Bureau, Department of Health and Environmental Science, Helena, MT.

Barbour, M. T., J. Gerritsen, B. D. Snyder, and J. B. Stribling. 1999. Rapid bioassessment protocols for use in streams and wadeable rivers: Periphyton, benthic macroinvertebrates, and fish, Second Edition. EPA 841-B-99-002, Washington, D.C.,U.S. Environmental Protection Agency, Office of Water.

Biggs, B. J. F. 1996. Patterns in benthic algae of streams. In Algal Ecology, Freshwater Benthic Ecosystems. R. J. Stevenson, M. L. Bothwell, and R. L. Lowe (eds.). Academic Press, NY.

Biggs, B. J. F. 2000. Eutrophication of streams and rivers: Dissolved nutrient-chlorophyll relationships for benthic algae. Journal of North American Benthological Society 19: 17-31.

Birks, H. J. B., S. Juggins, and J. M. Line. 1990a. Lake surface-water chemistry reconstruction from paleolimnological data. In: The Surface Water Acidification Programme. Edited by B.J. Mason. Cambridge Press, Cambridge. Pp.: 301-311.

Birks, H. J. B., J. M. Line, S. Juggins, A. C. Stevenson, and C. J. F. ter Braak. 1990b. Diatoms and pH reconstructions. Philosophical Transactions of the Royal Society of London, series B 327: 263-278.

Birks, H. J. B. 2001. Maximum likelihood calibration and the compute program WACALIB a correction. Journal of Paleolimnology 25(1): 111-115.

Blum, J. L. 1956. The ecology of river algae. Botanical Review 22: 291-331.

Charles, D., D. Winter, and M. Hoffman. 2000. Field Sampling procedures for the New Jersey Algae Indicators project. PCER Procedure P-13-64. Patrick Center for Environmental Research, Academy of Natural Sciences, Philadelphia, PA. 18 pp.

Charles, D. F., C. Knowles, R. Davis (eds.). 2002. Protocols for the analysis of algal samples collected as part of the U.S. Geological Survey, National Water-Quality Assessment Program. Report No. 02-06. The Academy of Natural Sciences. Patrick Center for Environmental Research, Academy of Natural Sciences, Philadelphia, PA. 124 pp.

Cemagref. 1982. Etude des méthodes biologiques quantitatives d'appréciation de la qualité des eaux. Rapport Division Qualité des eaux Lyon- Agence financière de Bassin Rhône-Mediterranée- Corse, Pierre-Bénite, 28 pp.

Cotter, P. 1999-2000. Tabulator, version 3.7.0. Patrick Center for Environmental Research, ANSP, Philadelphia, PA.

Cotter, P. 2001. Tabulator Installation and Users Guide. Patrick Center for Environmental Research, ANSP, Philadelphia, PA. 24 pp.

Cuffney, T. F., Gurtz, M.. E., and M. R. Meador. 1993. Methods for collecting benthic invertebrate samples as part of the National Water-Quality Assessment Program. Open-File Report 93-406. U.S. Geological Survey, Raleigh, NC. 66 pp.

Dixit, S. S., Smol, J. P., Kingston, J. C., and D. F. Charles. 1992. Diatoms: Powerful indicators of environmental change. Environmental Science and Technology 26 (1): 23-33.

Dodds, W. K., V. H. Smith, and K. Lohman. 2002. Nitrogen and phosphorus relationships to benthic algal biomass in temperate streams. Canadian Journal of Fisheries and Aquatic Sciences 59: 865-874.

Dodds, W. K., and E. B. Welch. 2000. Establishing nutrient criteria in streams. Journal of the North American Benthological Society 19: 186-196.

Dytham, C. 1999. Choosing and Using Statistics. A Biologist's Guide. Blackwell. Oxford.

Ennis, G. L., and L. J. Albright. 1982. Distribution and abundance of periphyton and phytoplankton species in two Subarctic Canadian rivers. Canadian Journal of Botany 60:224-236.

Environmental and Energy Development Solutions (ENSR). 2001. The Relationships Between Nutrient Concentrations and Periphyton Levels in Rivers and Streams - A Review of Scientific Literature. New England Interstate Water Pollution Control Commission. Document Number 4933-001-400.

Fallu, M. A., N. Allaire, and R. Pienitz. 2000. Freshwater diatoms from northern Québec and Labrador (Canada). Bibliotheca Diatomologica, Band 45: 1-200.

Fitzpatrick, F. A., Waite, I. R., D'Arconte, P. J., Meador, M. R., Maupin, M. A., and Gurtz, M. E. 1998. Revised Methods for Characterizing Stream Habitat in the National Water-Quality Assessment Program: U.S. Geological Survey Water-Resources Investigations Report 98-4052. 67 pp.

Francoeur, S.N. 2001. Meta-analysis of lotic nutrient amendment experiments: Detecting and quantifying subtle responses. Journal of North American Benthological Society 20(3): 358-368.

Hall, R. L., and J. P. Smol. 1992. A weighted averaging regression and calibration model for inferring total phosphorous concentration from diatoms in British Columbia (Canada) lakes. Freshwater Biology 27: 417-434.

Horwitz, R. and C. Flinders. 2003. Bioassessment Integration Study: Systems Ecology Evaluation of US EPA Rapid Bioassessment Protocols in New Jersey (Macroinvertebrates, Periphyton, Fish and Habitat) Quality Control/Quality Assurance Project Plan.

Jongman, R. H. G., C. J. F. ter Braak, and O. F. R. van Tongeren.1995. Data Analysis in Community and Landscape Ecology. Cambridge University Press, Cambridge.

Juggins, S., and C. J. F. ter Braak. 1997. CALIBRATE- a computer program for species-environmental calibration by Weighted averaging Partial least square regression. 0.81. University of Newcastle. Newcastle, UK.

Juggins, S., and C. J. F. ter Braak. 2001. CALIBRATE Version 1.0. A C++ Program for analysing and visualising species environment relationships and for predicting environmental values from species assemblages. User Guide Version 1.0. Department of Geography. University of Newcastle. Newcastle, UK.

Kelly, M. G. 1998. Use of the trophic diatom index for monitoring eutrophication in rivers. Water Resources 32: 236-242.

Kelly, M. G., and B. A. Whitton, 1995. The trophic diatom index. A new index for monitoring eutrophication in rivers. Journal of Applied Phycology: 7: 433-444.

Kelly, M. G., and B. A. Whitton. 1998. Biological monitoring of eutrophication in rivers. Hydrobiologia 384: 55-67.

Kiry, P., D. Velinsky, and A.-M. Compton. 1999. Determination of dry weight and percent organic matter for sediments, tissues and benthic algae. PCER Procedure P-16-113. Patrick Center for Environmental Research, Academy of Natural Sciences, Philadelphia, PA. 3 pp.

Lecointe, C., M. Coste. and J. Prygiel. 1993. OMNIDIA: A software for taxonomy, calculation of diatom indices and inventories management. Hydrobiologia 269/270: 590-513.

Line, J. M., C. J. F. ter Braak, and H. J. B. Birks. 1994. WACALIB version 3.3- a computer program to reconstruct environmental variables from fossil assemblages by weighted averaging and to derive sample-specific errors of prediction. Journal of Paleolimnology 10: 147-152.

New Jersey Department of Environmental Protection (NJ DEP). 1999. Geologic Map of New Jersey. NJ DEP, Division of Science and Research, Geological Survey.

New Jersey State Department of Environmental Protection (NJ DEP). 1994. The establishment of ecoregion biological reference sites for New Jersey streams. Incorporating Habitat Quality and Benthic Macroinvertebrate Communities 1989 - 1993 Monitoring Data. State of New Jersey, Christine Todd Whitman Governor, Robert C. Shinn, Jr. Commissioner. Trenton, NJ.

New Jersey Department of Environmental Protection. (NJ DEP). 2000. Water Quality Monitoring Networks 2000. NJ DEP Division of Watershed Management. Trenton, NJ.

New Jersey Department of Environmental Protection (NJ DEP). 2001. Surface water quality standards. N.J.A.C. 7:9B. Trenton, NJ. 129 pp.

Moulton, S. R., J. G. Kennen, R. M. Goldstein, and J. A. Hambrook. 2002. Revised Protocols for Sampling Algal, Invertebrate and Fish Communities as Part of the National Water-Quality Assessment Program. Open-File Report 02-150. US Geological Survey, Reston, VA. 75 pp.

Omernik, J. M. 1987. Ecoregions of the conterminous United States. Annals of the Association of American Geographers. 77(1):118-125.

Office of the New Jersey State Climatologist (ONJSC). 1994-2002. Rutgers University.

Patrick Center for Environmental Research. 2000. Quality Assurance Project Plan, Project Name: Understanding the Relationship Between Natural Conditions And Loadings on Eutrophication: Algal Indicators of Eutrophication for New Jersey Streams, The Academy of Natural Sciences, Philadelphia, PA.

Patrick, R. 1951. A proposed biological measure of stream conditions. Verhandlungen. Internationale Vereinigung für Theoretische und Angewandte Limnologie 11: 299-307.

Ponader, K., and D. Winter. 2002. Procedure for semi-quantitative analysis of soft-algae and diatoms. Patrick Center for Environmental Research. Procedure No. P-13-65. Patrick Center for Environmental Research, The Academy of Natural Sciences, Philadelphia, PA. 6 pp.

Porter, S. D., T. F. Cuffney, M. E. Gurtz, and M. R. Meador. 1993. Methods for Collecting Algal Samples as Part of the National Water-Quality Assessment Program. Open-File Report 93-409. U.S. Geological Survey, Raleigh, NC. 39 pp.

Prygiel J. and M. Coste. 1999. Progress in the use of diatoms for monitoring rivers in France. In: J. Prygiel, B. A Whitton. and J. Bukowska (eds.), Use of Algae for Monitoring Rivers III. Agence de l'eau Artois-Picardie, Douai, (France). pp. 165-179.

Reed, T. J. , B. T. White, G. L. Centinaro, J. F. Dudek, V. Corcino, A. B Spehar, and A. R. Protz. 2002. Water Resources Data, New Jersey, Water Year 2001. Volume 1. Surface-Water Data. Water-Data Report, NJ-01-1. West Trenton, NJ. 297 pp.

Schwoerbel, J. 1999. Einführung in die Limnologie. Gustav Fischer Verlag. Stuttgart.

Smith, V. H., Tilman, G. D., and J. C. Nekola. 1999. Eutrophication: Effects of excess nutrient inputs on freshwater, marine, and terrestrial ecosystems. Environmental Pollution 100: 179-196.

Stevenson, R. J. and Y. Pan. 1999. Assessing environmental conditions in rivers and streams with diatoms. In: Stoermer E. F. and J. P. Smol (eds.). The Diatoms: Applications for the Environmental and Earth Sciences. Cambridge University Press, Cambridge, UK. Pp.11-40.

Tedrow, J. C. F. 1986. Soils of New Jersey. Robert E. Krieger Publishing Company, Inc., Malabar, FL.

ter Braak, C. J. F. and I. C. Prentice, 1998. A theory of gradient analysis. Advances in Ecological Research 18: 221-317.

ter Braak, C. J. F. and H. Van Dam. 1989. Inferring pH from diatoms: A comparison of old and new calibration methods. Hydrobiologia 178: 209-223.

U.S. EPA. 1992. Methods for the determination of chemical substances in marine and estuarine environmental samples. EPA/600/R-92/121, Office of Research and Development, U.S. EPA, Washington, D.C.

US. EPA. 2000a. Nutrient Criteria Technical Guidance Manual: Rivers and Streams, U.S. Environmental Protection Agency, Washington, DC. EPA-822-B-00-002.

US. EPA. 2000b. Ambient Water Quality Criteria Recommendations: Rivers and Streams in Nutrient Ecoregion IX. U.S. Environmental Protection Agency, Washington, DC. EPA-822-B-00-019.

Velinsky, D. 2000. Syringe water sampling and filtration for the collection of filtered nutrient samples and unfiltered nutrient samples. PCER Procedure P-16-119. Patrick Center for Environmental Research, Academy of Natural Sciences, Philadelphia, PA. 4 pp.

Velinsky, D., and J. DeAlteris. 2000. Benthic algae and sediment chlorophyll *a* preparation and analysis. PCER Procedure P-16-117. Patrick Center for Environmental Research, Academy of Natural Sciences, Philadelphia, PA. 4 pp.

Winter, J. G. and H. C. Duthie. 2000. Epilithic diatoms as indicators of stream total N and total P concentration. Journal of the North American Benthological Society 19(1): 32-49.

Wolfe, P. E. 1977. The Geology and Landscapes of New Jersey. Crane, Russak and Company, Inc., N.Y. 351 pp.

Appendix 1 Summary of site characteristics. All sites sampled in 2000 and 2001.

1a) Raw environmental data for variables measured at each site reach.

Note: All variables were measured by PCER staff in the field at time of algal sampling and/or in the laboratory. **General site characteristics**: Sr_no , site reach number; S_date, algae sampling date; B_size, basin size; %O, percent open canopy cover; R_len, reach length; R_wid, average river width at reach; Flow, flow estimate in categories (1=slow; 2=moderate; 3=fast); **Substrate**: %Bed, % bedrock; %Bo, %boulder; %Cob, % cobble; %Gra, % gravel; %S, % sand; %Si/Cl, % silt and clay. **Biomass**: AFDM, Ash free dry mass; Chl_a, chlorophyll a;

Site ID	Sr_no	S_date	% O	% Bed	% Bo	% Cob	% Gra	% San	% Si/Cl	R_wid (m)	R_len (m)	Flow	AFDM (g/m2)	Chl_a (mg/m^2)
AN0081	NJ_081_1	10:22 am	100	0	16.7	35	38.3	10	0	8	50	2	16.3	152.8
AN0081	NJ_081_2	10:22 am	62.4	20	15	35	31.7	11.7	0	8	50	2	8.9	83.3
AN0081	NJ_081_3	10:22 am	12.48	23.3	10	18.3	20	8.3	0	8	50	2	4.8	37.4
AN0111	NJ_111_1	10:22 am	16.64	0	5	20	45	30	0	7	15	1.5	17.1	73.4
AN0111	NJ_111_2	10:22 am	6.24	0	5	30	15	30	20	7	5	1	12.6	64.2
AN0115	NJ_115_1	10:22 am	23.92	0	0	5	15	80	0	4	30	2	78.9	98.4
AN0115	NJ_115_2	10:22 am	31.2	0	0	2	25	73	0	4	25	2	10.8	30.9
AN0115	NJ_115_3	10:22 am	26	0	0	0	15	85	0	4	25	2	32.1	32.8
AN0115	NJ_115_1	10:22 am	9.375	0	0	5	15	80	0	4	30	1	22.7	69.7
AN0115	NJ_115_2	10:22 am	11.46	0	0	2	25	73	0	4	25	1	24.8	85.1
AN0115	NJ_115_3	10:22 am	19.79	0	0	0	15	85	0	4	25	1	20.1	111.5
AN0118	NJ_118_1	10:22 am	82.16	0	50.8	20.8	25.8	4.2	0	20	50	3	153.1	1114.7
AN0118	NJ_118_2	10:22 am	86.32	0	21.7	39.2	38.3	0.8	0	20	50	3	11.8	64.5
AN0118	NJ_118_3	10:22 am	22.88	0	5	35	60	0	0	20	50	3	7.6	14.4
AN0192	NJ_192_1	10:22 am	14.58	0	40	30	20	10	0	2.75	20	1.5	8.8	41.4
AN0192	NJ_192_2	10:22 am	18.75	0	10	20	30	30	10	2.75	20	2	5.4	17.2
AN0192	NJ_192_3	10:22 am	53.13	0	5	5	20	70	0	2.75	20	1	8.1	69.9
AN0194	NJ_194_1	10:22 am	82.16	0	0	0	0	50	50	6	30	1.5	--	--
AN0194	NJ_194_2	10:22 am	100	0	0	0	0	50	50	6	30	2.5	--	--
AN0194	NJ_194_3	10:22 am	100	0	0	0	0	50	50	6	30	2	--	--
AN0195	NJ_195_1	10:22 am	21.84	0	0	35	55	10	0	11.5	40	1.5	23.2	128.8
AN0195	NJ_195_2	10:22 am	26	0	0	45	45	10	0	11.5	40	1	33.1	228.5
AN0195	NJ_195_3	10:22 am	76.96	0	0	35	60	5	0	11.5	40	1	22.8	152.3
AN0207	NJ_207_1	10:22 am	69.79	0	0	0	5	90	5	15	50	1	4.2	36.7
AN0207	NJ_207_2	10:22 am	55.21	0	15	25	40	20	0	15	50	2	9.5	40.4
AN0207	NJ_207_3	10:22 am	28.13	0	20	40	30	10	0	15	50	2	8.5	48.9
AN0209	NJ_209_1	10:22 am	1.042	0	0	30	30	40	0	3.5	30	2.5	16.1	86.0
AN0209	NJ_209_2	10:22 am	50	0	15	30	20	40	5	3.5	30	1.5	11.5	70.6
AN0209	NJ_209_3	10:22 am	7.292	0	10	50	10	30	0	3.5	30	2	12.5	62.7
AN0211	NJ_211_1	10:22 am	5.2	0	10	40	20	30	0	5	20	2	18.7	67.6
AN0211	NJ_211_2	10:22 am	17.16	0	10	20	10	60	0	5	20	1.5	21.9	75.0
AN0211	NJ_211_3	10:22 am	14.56	0	5	30	25	40	0	5	20	2	16.9	80.9
AN0211	NJ_211_1	10:22 am	9.375	0	10	40	20	30	0	5	20	1.5	17.5	52.3
AN0211	NJ_211_2	10:22 am	15.63	0	10	20	10	60	0	5	20	1.5	11.9	43.1
AN0211	NJ_211_3	10:22 am	16.67	0	5	30	25	40	0	5	20	1.5	14.4	75.8
AN0215	NJ_215_1	10:22 am	53.04	0	5	25	20	45	5	3	16.5	1.5	5.6	32.6
AN0215	NJ_215_2	10:22 am	3.12	0	5	30	20	40	5	3	16.5	2	3.8	2.2
AN0215	NJ_215_3	10:22 am	5.2	0	5	15	5	70	5	3	16.5	2	5.0	5.5
AN0227	NJ_227_1	10:22 am	75.92	0	0	0	0	50	50	7	25	1	--	--
AN0227	NJ_227_2	10:22 am	58.76	0	0	0	0	50	50	7	25	1	--	--
AN0227	NJ_227_3	10:22 am	78.52	0	0	0	0	50	50	7	25	1	--	--
AN0231	NJ_231_1	10:22 am	100	0	0	0	0	50	50	50	50	1	30.2	29.2
AN0234	NJ_234_1	10:22 am	11.44	0	5	45	45	5	0	9	40	2.5	17.9	179.3
AN0234	NJ_234_2	10:22 am	10.4	0	15	40	30	15	0	9	40	2.5	8.1	47.8

Site ID	Sr_no	S_date	% O	% Bed	% Bo	% Cob	% Gra	% San	% Si/Cl	R_wid (m)	R_len (m)	Flow	AFDM (g/m2)	Chl_a (mg/m^2)
AN0234	NJ_234_3	10:22 am	21.84	0	15	50	30	5	0	9	40	2.5	9.1	87.2
AN0234	NJ_234_1	10:22 am	89.58	0	5	45	45	5	0	9	40	2.5	12.9	82.5
AN0234	NJ_234_2	10:22 am	7.292	0	15	40	30	15	0	9	40	2	9.2	42.3
AN0234	NJ_234_3	10:22 am	100	0	15	50	30	5	0	9	40	2	9.8	52.5
AN0235	NJ_235_1	10:22 am	59.38	0	15	30	25	30	0	20	60	2	22.9	183.5
AN0235	NJ_235_2	10:22 am	48.96	0	0	25	35	40	0	20	60	2.5	13.2	97.9
AN0235	NJ_235_3	10:22 am	39.58	0	0	10	30	60	0	20	60	1.5	9.9	101.8
AN0237	NJ_237_1	10:22 am	2.083	0	30	30	20	20	0	5	20	1.5	5.2	43.0
AN0237	NJ_237_2	10:22 am	7.292	0	60	30	10	0	0	5	30	2.5	7.5	58.8
AN0237	NJ_237_3	10:22 am	30.21	0	70	20	10	0	0	5	30	2	10.0	122.1
AN0238	NJ_238_1	10:22 am	46.8	0	0	0	0	50	50	12	40	1.5	--	--
AN0238	NJ_238_2	10:22 am	6.24	0	0	0	0	50	50	12	40	1.5	--	--
AN0238	NJ_238_3	10:22 am	14.56	0	0	0	0	50	50	12	40	1.5	--	--
AN0267	NJ_267_1	10:22 am	80.08	0	23.3	44.2	17.5	15	0	34	125	2	11.4	82.4
AN0267	NJ_267_2	10:22 am	88.4	0	16.7	34.2	30	19.2	0	34	100	1.5	9.0	80.6
AN0267	NJ_267_3	10:22 am	82.16	0	5	25	46.7	23.3	0	34	120	2	6.3	44.6
AN0274	NJ_274_1	10:22 am	100	0	30	30	30	10	0	45	12.5	3	10.1	86.8
AN0274	NJ_274_1	10:22 am	64.48	0	33.3	33.3	33.3	0	0	45	12.5	2.5	--	--
AN0281	NJ_281_1	10:22 am	5.2	0	6.7	26.7	35	31.7	0	4.5	25	2.5	9.9	24.9
AN0281	NJ_281_2	10:22 am	13.52	0	10	16.7	35	38.3	0	4.5	32	2.5	4.5	29.4
AN0281	NJ_281_3	10:22 am	79.04	0	5	25	41.7	31.7	0	4.5	32	2.5	18.9	202.2
AN0291	NJ_291_1	10:22 am	64.89	0	20	40	35	5	0	16.5	60	2.5	73.5	592.8
AN0291	NJ_291_2	10:22 am	78	0	0	3.3	31.7	65	0	16.5	50	1.5	45.0	262.7
AN0291	NJ_291_3	10:22 am	74.36	0	1.7	15	30	53.3	0	16.5	50	2	44.9	522.6
AN0318	NJ_318_1	10:22 am	16.64	0	55	16.7	10	18.3	0	8	40	2	7.6	53.2
AN0318	NJ_318_2	10:22 am	19.76	0	40	30	13.3	16.7	0	8	40	2	10.0	63.2
AN0318	NJ_318_3	10:22 am	38.48	0	21.7	28.3	13.3	36.7	0	8	40	1.5	8.0	69.1
AN0321	NJ_321_1	10:22 am	40.56	0	13.3	38.3	18.3	30	0	8.5	40	2.5	12.4	93.8
AN0321	NJ_321_2	10:22 am	77.48	0	6.7	41.7	25	26.7	0	8.5	40	2	13.4	109.8
AN0321	NJ_321_3	10:22 am	16.02	0	2.5	35	25	37.5	0	8.5	40	2	6.9	36.9
AN0326	NJ_326_1	10:22 am	92.56	0	15	35	35	15	0	27	90	2	25.5	200.2
AN0326	NJ_326_2	10:22 am	87.36	0	15	35	35	15	0	27	90	2	20.7	178.4
AN0326	NJ_326_3	10:22 am	84.24	0	15	35	35	15	0	27	90	2	18.0	134.8
AN0333	NJ_333_1	10:22 am	65.63	0	0	20	65	10	5	11	50	1	26.4	280.6
AN0333	NJ_333_2	10:22 am	65.63	0	0	30	50	10	10	11	50	1	27.9	206.2
AN0333	NJ_333_3	10:22 am	92.71	0	0	30	50	10	10	11	50	1	12.7	95.7
AN0339	NJ_339_1	10:22 am	29.12	0	6.7	36.7	51.7	5	0	5	25	2	28.9	145.0
AN0339	NJ_339_2	10:22 am	11.44	0	5	20	53.3	21.7	0	5	25	1.5	12.0	51.3
AN0339	NJ_339_3	10:22 am	29.12	0	0	45	38.3	16.7	0	5	25	2	8.7	66.0
AN0341	NJ_341_1	10:22 am	100	0	1.7	58.3	31.7	6.7	1.7	25	77.5	2	9.4	60.4
AN0341	NJ_341_2	10:22 am	100	0	0	35	40	16.7	8.3	25	77.5	2	19.9	140.8
AN0341	NJ_341_3	10:22 am	100	0	8.3	26.7	30	30	5	25	75	2	15.2	112.0
AN0370	NJ_370_1	10:22 am	100	0	0	14.2	59.2	26.7	0	20	75	2.5	30.8	117.0
AN0370	NJ_370_2	10:22 am	100	0	0	44.2	30	25.8	0	20	65	2	71.9	288.1
AN0370	NJ_370_3	10:22 am	100	0	0	23.3	51.7	25	0	20	65	2	20.1	129.1
AN0374	NJ_374_1	10:22 am	100	0	16.7	16.7	36.7	25	5	35	100	2.5	7.0	153.3
AN0374	NJ_374_2	10:22 am	100	0	1.7	43.3	35	18.3	1.7	35	75	2.5	29.3	214.2
AN0374	NJ_374_3	10:22 am	100	0	0	38.3	36.7	25	0	35	75	3	48.1	324.8
AN0374	NJ_374_1	10:22 am	86.46	0	16.7	16.7	36.7	25	5	35	100	2	13.1	91.6
AN0374	NJ_374_2	10:22 am	75	0	1.7	43.3	35	18.3	1.7	35	75	2.5	3.8	21.9
AN0374	NJ_374_3	10:22 am	87.5	0	0	38.3	36.7	25	0	35	75	2	18.1	115.0
AN0382	NJ_382_1	10:22 am	87	0	0	0	0	0	100	20	25	1	--	--
AN0382	NJ_382_2	10:22 am	100	0	0	0	0	0	100	20	25	1	--	--

Site ID	Sr_no	S_date	% O	% Bed	% Bo	% Cob	% Gra	% San	% Si/Cl	R_wid (m)	R_len (m)	Flow	AFDM (g/m2)	Chl_a (mg/m^2)
AN0382	NJ_382_3	10:22 am	100	0	0	0	0	0	100	20	25	1	--	--
AN0396	NJ_396_1	10:22 am	23.92	0	15	25	30	30	0	5	35	1.5	55.2	103.8
AN0396	NJ_396_2	10:22 am	8.32	0	1.7	11.7	40	46.7	0	5	35	1	4.7	35.2
AN0396	NJ_396_3	10:22 am	9.88	0	6.7	30	36.7	23.3	3.3	5	42	1	34.0	62.5
AN0405	NJ_405_1	10:22 am	12.48	5	5	10	35	35	10	12	75	1	118.1	119.9
AN0405	NJ_405_2	10:22 am	35.36	5	5	15	50	20	5	12	65	1.5	65.2	94.1
AN0405	NJ_405_3	10:22 am	23.92	5	5	5	35	25	25	12	65	1.5	75.8	100.2
AN0405	NJ_405_1	10:22 am	12.5	5	5	10	35	35	10	3.5	75	1	7.3	42.0
AN0405	NJ_405_2	10:22 am	22.92	5	5	15	50	20	5	3.5	65	1	10.4	69.4
AN0405	NJ_405_3	10:22 am	20.83	5	5	5	35	25	25	3.5	65	1	28.1	87.3
AN0413	NJ_413_1	10:22 am	15.08	0	6.7	10	58.3	20	5	8	35	1.5	10.9	99.7
AN0413	NJ_413_2	10:22 am	22.88	0	10	16.7	53.3	20	0	8	36	1.5	22.9	143.5
AN0413	NJ_413_3	10:22 am	29.12	0	3.3	8.3	56.7	16.7	15	8	37	1.5	6.8	52.2
AN0414	NJ_414_1	10:22 am	57.5	0	0	0	0	0	100	35	1	1	--	--
AN0424	NJ_424_1	10:22 am	37.44	0	0	5	65	30	0	9	50	2	23.1	84.8
AN0424	NJ_424_2	10:22 am	27.56	0	3.3	5	20	71.7	0	9	50	1.5	16.8	46.3
AN0424	NJ_424_3	10:22 am	30.68	0	1.7	8.3	31.7	58.3	0	9	40	2	22.8	136.1
AN0429	NJ_429_1	10:22 am	55.12	0	5	15	50	30	0	6	36	2	8.5	81.2
AN0429	NJ_429_2	10:22 am	14.56	0	3.3	35	51.7	10	0	6	36	2	9.9	100.1
AN0429	NJ_429_3	10:22 am	16.64	0	0	13.3	70	16.7	0	6	36	2	5.8	56.6
AN0439	NJ_439_1	10:22 am	13.78	0	0	15	40	0	45	6	60	2	10.8	29.8
AN0439	NJ_439_2	10:22 am	18.2	0	0	5	75	20	0	6	40	2	8.0	9.8
AN0439	NJ_439_3	10:22 am	19.5	0	0	0	30	0	70	6	45	2.5	7.1	19.5
Min			1	0	0	0	0	0	0	2.75	1	1	3.8	2.1
Max			100	23.3	70	58.3	75	90	100	50	125	3	153.1	1114.8
Mean			46.9	0.6	8.6	22.4	29.4	28.7	10.2	12.7	44.1	1.8	20.5	109.8
Median			33.3	0	5	25	30	25	0	8.5	40	2	12.7	81.1
25th perct													8.7	48.1
75th perct													22.9	119.2

25th perct = 25th percentile

75th perct = 75th percentile (upper 25th percentile)

Appendix 1 Summary of site characteristics. All sites sampled in 2000 and 2001.

1b) Raw environmental data for variables measured at each site.

Note: S_date, algae sampling date[2]; **Landuse:** Urb %, percent urban[1] ; Agr %, percent agriculture[1] ; For %, percent forestry[1] ; **Waterchemistry:** Cond, specific conductivity[1]); NH_3-N[2] ; NO_3-N_2); TN, total nitrogen[1] (*TN calculated from combination of TKN[1]) and NO_3-N_2)); TKN, total Kjeldahl nitrogen[1]); PO_4, orthophospate[2]); TP, total phosphorus[2]); pH[1]); Alk, alkalinity[2]); Hard, hardness[2]);

[1] data provided by NJ DEP and/or data records collected by NJ DEP and USGS at surface water monitoring stations, measured within a maxiumum of 4 weeks from algal sampling.

[2] variable measured by PCER Geochemistry section.

Site ID	Lat N	Long W	S_date	Area (km^2)	Urb %	Agr %	For %	Cond (µS/cm)	NH_3-N (mg/L)	NO_3-N (mg/L)	TN (mg/L)	TKN (mg/L)	PO_4 (mg/L)	TP (mg/L)	pH	Alk (mg/L)	Hard (mg/L)
AN0081	40° 32'	75° 02'	36782	24.8	16	53	21	144	0.006	1.579	1.6	0.5	0.041	0.051	8	-	-
AN0111	40° 17'	74° 42'	36793	15.7	39	30	13	298	0.045	1.744	-	-	0.04	0.071	8	-	-
AN0115	40° 14'	74° 41'	36746	27.1	46	23	3	93	0.066	1.154	1.11	0.8	0.004	0.026	7	-	-
AN0115	40° 14'	74° 41'	37128	27.1	46	23	3	192	0.16	2.565	3.315*	0.75	0.001	0.012	7	20	55
AN0118	40° 13'	74° 45'	36801	233.2	43	20	10	292	0.126	2.492	3.7	0.7	0.268	0.322	8	-	-
AN0192	40° 46'	74° 16'	37124	10.1	76	0	20	983	0.086	0.397	0.937*	0.54	0.034	0.056	7	80	280
AN0194	40° 40'	74° 18'	36796	78.2	75	0	20	-	0.075	1.349	-	-	0.019	0.063	-	-	-
AN0195	40° 37'	74° 16'	36787	104.9	79	0	16	407	0.033	0.966	1.303	0.428	0.041	0.075	8	-	-
AN0207	40° 59'	74° 01'	37125	42.1	80	1	11	552	0.159	2.535	3.175*	0.64	0.232	0.337	8	90	140
AN0209	40° 58'	73° 58'	37125	22.3	86	0	10	453	0.177	1.529	1.829*	0.3	0.015	0.055	8	100	150
AN0211	40° 54'	74° 02'	36796	15.4	92	0	5	658	0.068	1.102	2.008	0.323	0.02	0.05	8	-	-
AN0211	40° 54'	74° 02'	37126	15.4	92	0	5	214	0.116	1.128	1.688*	0.56	0.042	0.082	7	48	65
AN0215	40° 46'	74° 32'	36783	1.4	6	0	93	-	0.008	0.265	-	-	0.008	0.022	-	-	-
AN0227	40° 38'	74° 31'	36800	146.6	37	7	26	-	0.023	7.553	-	-	1.135	1.297	-	-	-
AN0231	40° 49'	74° 20'	37124	339.1	48	4	23	670	0.097	5.032	5.742*	0.71	0.573	0.699	8	90	150
AN0234	40° 48'	74° 27'	36795	72.6	47	3	43	209	0.068	1.974	0.89	0.7	0.116	0.153	7		
AN0234	40° 48'	74° 27'	37123	72.6	47	3	43	327	0.061	1.366	1.856*	0.49	0.07	0.125	8	55	95
AN0235	40° 49'	74° 26'	37123	82.6	51	2	39	434	0.012	2.306	-	-	0.078	0.118	-	70	115
AN0237	40° 51'	74° 23'	37124	27.2	61	0	23	436	0.023	0.232	-	-	0.02	0.064	-	95	158
AN0238	40° 50'	74° 20'	36800	178.9	55	1	27	-	0.082	2.099	-	-	0.168	0.262	-	-	-
AN0267	41° 02'	74° 14'	36794	72.2	29	1	60	303	0.027	1.238	0.987	0.286	0.088	0.11	8	-	-
AN0274	40° 53'	74° 13'	36800	1612.2	37	1	43	389	0.032	3.106	2.5	0.8	0.447	0.506	7	-	-
AN0274	40° 53'	74° 13'	37126	1612.2	37	1	43	478	0.022	2.591	-	0.66	0.359	0.458	8	70	120
AN0281	41° 01'	74° 06'	36788	19.6	82	0	11	493	0.024	1.924	-	-	0.011	0.032	8	-	-
AN0291	40° 51'	74° 06'	36797	135.4	82	1	10	595	0.14	5.524	5.11	0.804	0.662	0.73	8	-	-
AN0318	40° 43'	74° 54'	36781	14.4	13	19	51	-	0.002	0.342	-	-	0.006	0.011	-	-	-
AN0321	40° 38'	74° 58'	36781	30	24	20	43	-	0.004	0.822	-	-	0.015	0.034	-	-	-
AN0326	40° 34'	74° 52	36782	386.9	25	21	41	269	0.008	1.308	1.766	0.326	0.022	0.042	8	-	-
AN0333	40° 28'	74° 49'	37122	65.5	24	41	22	275	0.012	1.091	1.381*	0.29	0.01	0.035	9	78	105
AN0339	40° 33'	74° 47'	36775	10.3	31	30	33	210	0.003	1.104	-	-	0.06	0.068	8	-	-
AN0341	40° 32'	74° 41	36780	685.6	25	28	33	269	0.026	1.275	1.9	0.5	0.079	0.103	8	-	-
AN0370	40° 38'	74° 41'	36775	256.3	24	23	43	255	0.005	1.046	1.5	0.4	0.064	0.082	7	-	-
AN0374	40° 34'	74° 40'	36779	484.2	28	21	42	251	0.006	0.516	1.421	0.457	0.012	0.032	8	-	-
AN0374	40° 34'	74° 40'	37122	484.2	28	21	42	298	0.021	0.652	0.982*	0.33	0.044	0.063	8	72	105
AN0382	40° 19'	74° 36'	36801	112.6	22	32	16	-	0.032	4.273	-	-	0.041	0.065	-	-	-
AN0396	40° 22'	74° 36'	36774	24.2	30	14	25	212	0.012	4.557	2.071	0.536	0.014	0.024	7	-	-
AN0405	40° 25'	74° 38'	36776	56.3	28	23	31	285	0.024	3.11	-	-	0.077	0.103	8	-	-
AN0405	40° 25'	74° 38'	37128	56.3	28	23	31	529	0.016	5.73	-	-	0.045	0.06	-	80	170
AN0413	40° 32'	74° 35'	36780	40.6	48	16	17	282	0.012	1.344	-	-	0.087	0.11	8	-	-
AN0414	40° 32'	74° 34'	36800	734.3	31	25	22	-	0.077	2.546	-	-	0.247	0.288	-	-	-

Site ID	Lat N	Long W	S_date	Area (km^2)	Urb %	Agr %	For %	Cond (µS/cm)	NH_3-N (mg/L)	NO_3-N (mg/L)	TN (mg/L)	TKN (mg/L)	PO_4 (mg/L)	TP (mg/L)	pH	Alk (mg/L)	Hard (mg/L)
AN0424	40˚ 34'	74˚ 29'	36786	60.1	79	0	3	460	0.15	0.969	1.25	0.6	0.067	0.1	8	-	-
AN0429	40˚ 30'	74˚ 28'	36786	14.8	91	0	5	461	0.024	1.307	-	-	0.076	0.093	8	-	-
AN0439	40˚ 17'	74˚ 23'	36773	51.9	23	24	19	-	0.057	0.538	-	-	0.012	0.062	-	-	-
Min	40˚ 13'	73˚ 58'	36746	1.4	6	0	3	93	0.002	0.232	0.89	0.286	0.001	0.011	7	20	55
Max	41˚ 02'	75˚ 02'	37128	1612.2	92	53	93	983	0.177	7.553	5.742	0.804	1.135	1.297	9	100	280
Mean	-	-	-	199	46	13	26	373	0.052	2.007	1.941	0.537	0.127	0.166	8	73	131
Median	-	-	-	60.1	39	7	23	300	0.027	1.349	1.6	0.536	0.044	0.071	8	78	120
25th perct	-	-	-	24.5	28	0	12	259	0.012	1.069	1.277	0.4	0.017	0.051	8	70	105
75th perct	-	-	-	162.8	68	23	41	461	0.067	2.541	2.04	0.7	0.088	0.122	8	90	150

25th perct = 25th percentile

75th perct = 75th percentile (upper 25th percentile)

Appendix 2: Diatom species list

Taxonomic index of 306 diatom species from samples collected in 2000 and 2001, used for development of models and metrics. Numbers correspond to numbers used in graphs in the text.

No.	Taxon name
1	*Achnanthidium minutissimum* (Kützing) Czarnecki
2	*Achnanthes exigua* Grunow
3	*Achnanthes lanceolata* (Brébisson in Kützing) Grunow
4	*Achnanthes lapidosa* Krasske
5	*Achnanthes linearis* (Smith) Grunow
6	*Achnanthes* sp.1 NEW JERSEY KCP
7	*Achnanthes peragalli* Brun et Héribaud
8	*Achnanthes pinnata* Hustedt
9	*Achnanthes lanceolata* var. *apiculata* Patrick
10	*Achnanthes delicatula* (Kützing) Grunow
11	*Achnanthes lanceolata subsp. rostrata* (Østrup) Lange-Bertalot
12	*Achnanthes chlidanos* Hohn et Hellermann
13	*Achnanthes lanceolata* var. *abbreviata* Reimer
14	*Achnanthes subhudsonis* var. *kraeuselii* Cholnoky
15	*Achnanthes minutissima* var. *saprophila* Kobayasi et Mayama
16	*Achnanthes harveyi* Reimer
17	*Achnanthes exigua var. constricta* Torka
18	*Achnanthes petersonii* Hustedt
19	*Achnanthes rupestoides* Hohn
20	*Achnanthes minutissima var. scotica* (Carter) Lange-Bertalot
21	*Achnanthes subatomus* Hustedt
22	*Achnanthes daui* Foged
23	*Achnanthes grana* Hohn & Hellermann
24	*Achnanthes lanceolata* subsp. *frequentissima* Lange-Bertalot
25	*Amphora ovalis* (Kützing) Kützing
26	*Amphora libyca* Ehrenberg
27	*Amphora montana* Krasske
28	*Amphora pediculus* (Kützing) Grun.
29	*Asterionella formosa* Hassal
30	*Aulacoseira pfaffiana* (Reinsch) Krammer
31	*Aulacoseira ambigua* (Grunow) Simonsen
32	*Aulacoseira distans* (Ehrenberg) Simonsen
33	*Aulacoseira subartica* (O. Müller) Haworth
34	*Aulacoseira granulata* (Ehrenberg) Simonsen
35	*Aulacoseira italica* (Ehrenberg) Simonsen
36	*Caloneis bacillum* (Grunow) Cleve
37	*Caloneis hyalina* Hustedt
38	*Caloneis silicula* (Ehrenberg) Cleve
39	*Capartogramma crucicula* (Grunow ex Cleve) Ross
40	*Cocconeis placentula* var. *lineata* (Ehrenberg) Van Heurck
41	*Cocconeis placentula* var. *euglypta* (Ehrenberg) Cleve

42 *Cocconeis fluviatilis* Wallace
43 *Cocconeis pediculus* Ehrenberg
44 *Cyclostephanos tholiformis* Stoermer, Håkansson et Theriot
45 *Cyclostephanos invisitatus* (Hohn et Hellermann) Theriot, Stoermer et Håkansson
46 *Cyclotella atomus* Hustedt
47 *Cyclotella meneghiniana* Kützing
48 *Cyclotella ocellata* Pantosek
49 *Cyclotella stelligera* (Cleve et Grunow) Van Heurck
50 *Cyclotella pseudostelligera* Hustedt
51 *Cyclotella operculata* (Agardh) Kützing
52 *Cyclotella* sp. 1 ANS NEW JERSEY KCP
53 *Craticula cuspidata* (Kützing) Mann
54 *Cymatopleura solea* (Brébisson) Smith
55 *Cymbella naviculiformis* Auerswald ex Héribaud
56 *Cymbella aspera* (Ehrenberg) Peragallo
57 *Cymbella tumida* (Brébisson ex Kützing) Van Heurck
58 *Cymbella proxima* Reimer
59 *Cymbella tumidula* Grunow ex Schmidt
60 *Denticula elegans* Kützing
61 *Diatoma mesodon* (Ehrenberg) Kützing
62 *Diatoma vulgaris* Bory
63 *Diploneis oblongella* (Naegeli ex Kützing) Ross
64 *Diploneis puella* (Schumann) Cleve
65 *Epithemia turgida* var. *westermannii* (Ehrenberg) Grunow
66 *Eunotia exigua* (Brébisson ex Kützing) Rabenhorst
67 *Eunotia formica* Ehrenberg
68 *Eunotia pectinalis* var. *minor* (Kützing) Rabenhorst
69 *Eunotia praerupta* Ehrenberg
70 *Eunotia praerupta* var. *bidens* (Ehrenberg) Grunow
71 *Eunotia minor* (Kützing) Grunow
72 *Eunotia bilunaris* (Ehrenberg) Mills
73 *Fragilaria capucina* Desmazières
74 *Fragilaria construens* (Ehrenberg) Grunow
75 *Fragilaria crotonensis* Kitton
76 *Fragilaria pinnata* Ehrenberg
77 *Fragilaria pinnata* var. *lancettula* (Schumann) Hustedt
78 *Fragilaria vaucheriae* (Kützing) Petersen
79 *Fragilaria brevistriata* var. *inflata* (Pantocsek) Hustedt
80 *Fragilaria fasciculata* (Agardh) Lange-Bertalot
81 *Fragilaria nanana* Lange-Bertalot
82 *Fragilaria capucina* var. *gracilis* (Oestrup) Hustedt
83 *Fragilaria capucina* var. *rumpens* (Kützing) Lange-Bertalot
84 *Fragilaria parasitica* var. *subconstricta* Grunow
85 *Fragilaria* sp. 1 ANS NEW JERSEY KCP
86 *Fragilaria* sp. 1 ?
87 *Frustulia rhomboides* (Ehrenberg) De Toni
88 *Frustulia rhomboides* var. *amphipleuroides* (Grun.) DeT.
89 *Frustulia vulgaris* (Thwaites) DeT.

90 *Frustulia weinholdii* Hustedt
91 *Frustulia crassinervia* (Brebisson) Lange-Bertalot et Krammer
92 *Gomphoneis herculeana* (Ehr.) Cl.
93 *Gomphoneis minuta* Kociolek & Stoermer
94 *Gomphonema affine* Kützing
95 *Gomphonema angustatum* (Kütz.) Rabh.
96 *Gomphonema gracile* Ehr. emend. V. H.
97 *Gomphonema parvulum* (Kütz.) Kütz.
98 *Gomphonema truncatum* Ehrenberg
99 *Gomphonema sphaerophorum* Ehrenberg
100 *Gomphonema turris* Ehrenberg
101 *Gomphonema olivaceoides* Hustedt
102 *Gomphonema manubrium* Fricke
103 *Gomphonema pumilum* (Grun.) Reich. & Lange-Bert.
104 *Gomphonema sarcophagus* Greg.
105 *Gomphonema micropus* Kützing
106 *Gomphonema minutum* (Ag.) Ag.
107 *Gomphonema lingulatiforme* Lange-Bertalot & Reichardt
108 *Gomphonema patrickii* Kociolek & Stoermer
109 *Gomphonema kobayasii* Kociolek & Kingston
110 *Gomphonema* aff. *parvulum* var. *saprophilum* ANS NEW JERSEY KCP
111 *Gomphonema* sp. 1 ANS NEW JERSEY KCP
112 *Gomphonema* sp. 2 ANS NEW JERSEY KCP
113 *Gyrosigma acuminatum* (Kütz.) Rabh.
114 *Gyrosigma obscurum* (W. Sm.) Griff. & Henfr.
115 *Hantzschia amphioxys* (Ehr.) Grun.
116 *Melosira varians* Ag.
117 *Meridion circulare* (Grev.) Ag.
118 *Navicula angusta* Grunow
119 *Navicula arvensis* Hustedt
120 *Navicula atomus* (Kütz.) Grun.
121 *Navicula biconica* Patr.
122 *Navicula cryptocephala* Kützing
123 *Navicula difficillima* Hustedt
124 *Navicula gregaria* Donk.
125 *Navicula kotschyi* Grunow
126 *Navicula minima* Grunow
127 *Navicula mutica* Kützing
128 *Navicula notha* Wallace
129 *Navicula paucivisitata* Patr.
130 *Navicula pupula* Kützing
131 *Navicula tripunctata* (O. F. Müll.) Bory
132 *Navicula rhynchocephala* Kützing
133 *Navicula capitata* Ehrenberg
134 *Navicula cryptocephala* var. *veneta* (Kütz.) Rabh.
135 *Navicula decussis* Østr.
136 *Navicula hustedtii* Krass.
137 *Navicula capitata* var. *hungarica* (Grun.) Ross

138 *Navicula peregrina* (Ehr.) Kütz.
139 *Navicula trivialis* Lange-Bert.
140 *Navicula canalis* Patr.
141 *Navicula capitata* var. *lueneburgensis* (Grun.) Patr.
142 *Navicula ingenua* Hustedt
143 *Navicula integra* (W. Sm.) Ralfs
144 *Navicula menisculus* Schum.
145 *Navicula placenta* Ehrenberg
146 *Navicula schroeteri* var. *escambia* Patr.
147 *Navicula secreta* var. *apiculata* Patr.
148 *Navicula salinarum* Grunow
149 *Navicula symmetrica* Patr.
150 *Navicula tenelloides* Hustedt
151 *Navicula tenera* Hustedt
152 *Navicula viridula* var. *rostellata* (Kütz.) Cl.
153 *Navicula agrestis* Hustedt
154 *Navicula protracta* (Grun.) Cl.
155 *Navicula minuscula* Grunow
156 *Navicula heufleri* var. *leptocephala* (Brèb ex Grun.) Perag.
157 *Navicula bacilloides* Hustedt
158 *Navicula absoluta* Hustedt
159 *Navicula veneta* Kützing
160 *Navicula longicephala* Hustedt
161 *Navicula molestiformis* Hustedt
162 *Navicula ignota* var. *acceptata* (Hust.) Lange-Bert.
163 *Navicula cryptotenella* L.B. in Kramm. & L.-B.
164 *Navicula perminuta* Grunow
165 *Navicula subminuscula* Mang.
167 *Navicula germainii* Wallace
168 *Navicula erifuga* Lange-Bert.
169 *Navicula recens* Lange-Bert.
170 *Navicula capitatoradiata* Germain
171 *Navicula atomus* var. *permitis* (Hust.) Lange-Bert.
172 *Navicula suchlandtii* Hustedt
173 *Navicula longicephala* var. *vilaplanii* Lange-Bertalot & Sabater
175 *Navicula lanceolata* (Ag.) Ehr.
176 *Navicula menisculus* var. *grunowii* Lange-Bertalot
177 *Navicula ruttnerii* var. *capitata* Hustedt
178 *Navicula parabilis* Hohn & Hellerman
179 *Navicula* sp. 1 ANS NEW JERSEY KCP
180 *Neidium affine* (Ehr.) Pfitz.
181 *Neidium alpinum* Hustedt
182 *Nitzschia acicularioides* Hustedt
183 *Nitzschia acicularis* (Kütz.) W. Sm.
184 *Nitzschia amphibia* Grunow
185 *Nitzschia capitellata* Hustedt
186 *Nitzschia dissipata* (Kütz.) Grun.
187 *Nitzschia fonticola* Grunow

188 *Nitzschia frustulum* (Kütz.) Grun.
189 *Nitzschia gracilis* Hantz. ex Rabh.
190 *Nitzschia heufleriana* Grunow
191 *Nitzschia linearis* (Ag. ex W. Sm.) W. Sm.
192 *Nitzschia microcephala* Grunow
193 *Nitzschia palea* (Kütz.) W. Sm.
194 *Nitzschia palea* var. *tenuirostris* Grunow
195 *Nitzschia recta* Hantz. ex Rabh.
196 *Nitzschia sigma* (Kütz.) W. Sm.
197 *Nitzschia dissipata* var. *media* (Hantz.) Grun.
198 *Nitzschia hungarica* Grunow
199 *Nitzschia inconspicua* Grunow
200 *Nitzschia perminuta* (Grun.) Peragallo
201 *Nitzschia clausii* Hantz.
202 *Nitzschia constricta* var. *subconstricta* Grun. in Cl. et Grun.
203 *Nitzschia filiformis* (W. Sm.) V. H.
204 *Nitzschia gracilis* var. *minor* Skabitschevsky in Proschkina-Lavrenko
205 *Nitzschia intermedia* Hantz. ex Cl. et Grun.
206 *Nitzschia liebethruthii* Rabenhorst
207 *Nitzschia littoralis* Grun. in Cl. et Grun.
208 *Nitzschia lorenziana* var. *subtilis* Grun. in Cl. et Grun.
209 *Nitzschia paleacea* Grun. in V. H.
210 *Nitzschia rosenstockii* Lange-Bertalot
211 *Nitzschia tryblionella* var. *debilis* (Arnott) Hust.
212 *Nitzschia vermicularis* (Kütz.) Hantz. in Rabh.
213 *Nitzschia brevissima* Grun. in V. H.
214 *Nitzschia commutata* Grunow
215 *Nitzschia sociabilis* Hustedt
216 *Nitzschia palea* var. *debilis* (Kütz.) Grun.
217 *Nitzschia angustatula* Lange-Bert.
218 *Nitzschia sinuata* var. *delognei* (Grun.) Lange-Bert.
219 *Nitzschia flexoides* Geitler
220 *Nitzschia acidoclinata* Lange-Bert.
221 *Nitzschia tubicola* Grun. in Cl. et Grun.
222 *Nitzschia terrestris* (Peterson) Hust.
223 *Nitzschia coarctata* Grunow
224 *Nitzschia levidensis* var. *salinarum* Grunow
225 *Nitzschia lorenziana* Grunow
226 *Nitzschia levidensis* var. *victoriae* Grunow
227 *Nitzschia tryblionella* var. *salinarum* Grun. in Cl. et Grun.
228 *Nitzschia* aff. *fonticola* ANS NEWJERSEY KCP
229 *Nitzschia archibaldii* Lange-Bertalot
230 *Nitzschia* sp. 1 ANS NEW JERSEY KCP
231 *Nitzschia subconstricta* Grunow
232 *Pinnularia divergens* W. Sm.
233 *Pinnularia maior* (Kütz.) Rabh.
234 *Pinnularia mesolepta* (Ehr.) W. Sm.
235 *Pinnularia microstauron* (Ehr.) Cl.

236 *Pinnularia obscura* Krass.
237 *Pinnularia rupestris* Hautz.
238 *Pinnularia subcapitata* Greg.
239 *Pinnularia lundii* Hustedt
240 *Pinnularia interrupta* W. Sm.
241 *Pinnularia parvulissima* Kram.
242 *Plagiotropis lepidoptera* var. *proboscidea* (Cl.) Reim.
243 *Pleurosigma angulatum* (Quek.) W. Sm.
244 *Reimeria sinuata* (Greg.) Kociolek & Stoermer
245 *Rhoicosphenia curvata* (Kütz.) Grun. ex Rabh.
246 *Stauroneis anceps* Ehrenberg
247 *Stauroneis smithii* Grunow
248 *Stauroneis phoenicenteron* (Nitz.) Ehr.
249 *Stauroneis agrestis* Peters.
250 *Stenopterobia intermedia* (Lewis) V. H.
251 *Stephanodiscus niagarae* Ehrenberg
252 *Stephanodiscus hantzschii* Grunow
253 *Stephanodiscus minutus* H. L. Sm.
254 *Surirella angusta* Kützing
255 *Surirella linearis* W. Sm.
256 *Surirella robusta* Ehrenberg
257 *Surirella stalagma* Hohn & Hellerm.
258 *Surirella minuta* Bréb.
259 *Surirella brebissonii* var. *kuetzingii* Kramm. & Lange-Bert.
260 *Surirella brebissonii* Kramm. & Lange-Bert.
261 *Surirella amphioxys* W. Sm.
262 *Surirella splendida* (Ehr.) Kütz.
263 *Synedra parasitica* (W. Sm.) Hust.
264 *Synedra rumpens* var. *familiaris* (Kütz.) Hust.
265 *Synedra ulna* (Nitz.) Ehr.
266 *Synedra ulna* var. *oxyrhynchus* fo. *mediocontracta* (Fonti) Hust.
267 *Synedra delicatissima* var. *angustissima* Grunow
268 *Synedra parasitica* var. *subconstricta* (Grun.) Hust.
269 *Tabellaria flocculosa* (Roth) Kütz.
270 *Thalassiosira weissflogii* (Grun.) Fryxell & Hasle
271 *Pseudostaurosira brevistriata* (Grun. in V.H.) Williams & Round
272 *Bacillaria paradoxa* Gmelin
273 *Nupela neglecta* Ponader, Lowe & Potapova
274 *Nupela wellneri* (Lange-Bert.) Lange-Bert.
275 *Navicula* sp. *2* ANS NEW JERSEY KCP
276 *Navicula* sp. 3 ANS NEW JERSEY KCP
277 *Encyonema minutum* (Hilse) Mann
278 *Encyonema silesiacum* (Bleisch) Mann
279 *Encyonema prostratum* (Berkeley) Kützing
280 *Fallacia pygmaea* (Kützing) Stickle et Mann
281 *Fallacia auriculata* (Hustedt) Mann
282 *Fallacia omissa* (Hustedt) Mann
283 *Fallacia tenera* (Hustedt) Mann

284 *Karayevia clevei* (Grunow) Round & Buktyiarova
285 *Karayevia laterostrata* (Hant.) Round and Bukt.
286 *Luticola goeppertiana* (Bleisch) Mann
287 *Sellaphora seminulum* (Grun.) Mann
288 *Staurosira construens var. binodis* (Ehrenberg) Hamilton
289 *Staurosira construens var. venter* (Ehr.) Hamilton
290 *Staurosirella leptostauron* (Ehrenberg) Williams et Round
291 *Staurosirella pinnata* (Her.) Williams & Round
292 *Tryblionella apiculata* Greg.
293 *Tryblionella levidensis* Wm. Sm.
294 *Tryblionella aerophila*
295 *Psammothidium bioretii* (Germ.) Bukht. et Round
296 *Psammothidium subatomoides* (Hustedt) Bukhtiyarova et Round
297 *Psammothidium ventralis* (Kras.) Bukht. et Round
298 *Eucocconeis laevis* (Oestrup) Lange-Bertalot
299 *Fragilariforma constricta* (Ehrenberg) Williams et Round
300 *Fragilariforma virescens* (Ralfs) Williams et Round
301 *Placoneis clementis* (Grun) Cox
302 *Placoneis elginensis* (Greg.) Cox
303 *Placoneis explanata* (Hust.) Cox
304 *Cavinula pseudoscutiformis* (Grunow ex Schmidt) Mann et Stickle
305 *Diadesmis confervacea* Kützing
306 *Diadesmis contenta* (Grunow ex Van Heurck) Mann

Appendix 3: Spearman's rank-order correlation for full dataset for both years (*n*=106)

Note: Waterchemistry: Cond, specific conductivity[1]); NH_3-N[2]) ; NO_3-N[2]); TN, total nitrogen[1]) (*TN calculated from combination of TKN[1]) and NO_3-N_2)); TKN, total Kjeldahl nitrogen[1]); PO_4, orthophospate[2]); TP, total phosphorus[2]); Alk, alkalinity[2]); Hard, hardness[2]); pH[1]); DO, dissoved oxygen[1]); **General site characteristics**: B_size, basin size; %O, percent open canopy cover; R_len, reach length; R_wid, average river width at reach; Flow, flow estimate in categories (1=slow; 2=moderate; 3=fast); **Substrate**: %Bed, percent bedrock; %Bo, percent boulder; %Cob, percent cobble; %Gra, percent gravel; %S, percent sand; %Si/Cl, percent silt and clay, BRBDCOG, percent bedrock, boulder, cobble and gravel combined; SDSTCL, percent sand, silt and clay combined; BRBD, percent bedrock and boulder combined; CBGR, percent cobble and gravel combined; **Biomass**: AFDM, Ash free dry mass; Chl_a, chlorophyll a; [1)] data provided by NJ DEP and/or data records collected by NJ DEP and USGS at surface water monitoring stations, measured within a maxiumum of 4 weeks from algal sampling; [2)] variable measured by PCER Geochemistry section. **Algal cover/biomass estimate:** *Rapid biomass assessment (RBA), visual estimate of percent cover (%estimated cover):* %DGCov, %Dark Green (%estimated cover); %LsSCov, %Long spiny Spirogira (%estimated cover); %MdbCov, %Moss-like (Dark brown) (%estimated cover); %BgslCov, blue-green (slimey) (%estimated cover), BlgrCov, %blue-green (%estimated cover); %BCCov, %Brown Coating (%estimated cover); %ChldCov, %Chladophora (%estimated cover); %DfilCov, %Diatoms (filamentous) (%estimated cover); %DTBCov, %Diatoms (thin layer)+ Bare (%estimated cover); %GrFthCov, %Green (Feathery) (%estimated cover); %GrF1Cov, %Green Filamentous #1 (%estimated cover); %GrF2Cov, %Green Filamentous #2 (%estimated cover); %GFBCov, %Green Filamentous (Bushy) (%estimated cover); %GrFCov, %green filamentous (moss-like) (%estimated cover); %GrFsCov, %Green Fliamentous (Segemented branches) (%estimated cover); %HBCov, %Honey-Brown (%estimated cover); %LlGCov, %Long light green (%estimated cover); %OrgCov, %Orange (%estimated cover); %TgLCov, %thin green layer (%estimated cover); %TsCdCov, %Thin short Cladophera (%estimated cover); %GrBGCov, %Green (blue-green) (%estimated cover). *Semi-quantitative count method (SQCM): percent estimate of numbers of cells (% # cells)*: %DiatC, Diatoms (% # cell); %ChladC, Cladophora sp.(% # cell); %OegodC, Oedogonium sp.(% # cells); %RhizC, Rhizoclonium sp.(% # cells); %ScenC, Scenedesmus sp.(% # cells); %SpirogC, Spirogyra sp.(% # cells); %MerismopC, Merismopedia sp.(% # cells); %OscillC, Oscillatoria sp.(% # cells); %AudouC, Audouinella sp.(% # cells);

p- value: ** correlation significant at the 0.01 level (2-tailed test)
* correlation significant at the 0.05 level (2-tailed test)

	NH_3-N	PO_4	TP	Chl_a	AFDM	R_wid	R_len	Flow	
NO_3-N	0.32**	0.454**	0.383**	0.115	0.261**	0.141	0.222*	-0.176	*r*
	0	0	0	0.242	0.00703	0.149	0.0226	0.0718	*p*-value
NH_3-N		0.233*	0.366**	-0.164	0.113	-0.161	-0.232*	-0.0547	*r*
		0.0165	0	0.0939	0.247	0.0987	0.0169	0.577	*p*-value
PO_4			0.95**	0.167	0.0637	0.374**	0.207*	0.236*	*r*
			0	0.0873	0.516	0	0.0338	0.0151	*p*-value
TP				0.127	0.0679	0.361**	0.138	0.235*	*r*
				0.195	0.488	0	0.159	0.0156	*p*-value
Chl_a					0.716**	0.408**	0.341**	0.0771	*r*
					0	0	0	0.431	*p*-value
AFDM						0.251**	0.222*	-0.141	*r*
						0.00968	0.0224	0.15	*p*-value
R_wid							0.757**	0.28**	*r*
							0	0.00373	*p*-value
R_len								0.143	*r*
								0.144	*p*-value

	B_size	DO	pH	Cond	% O	% DiatC	% ChladC	% OegodC	
NO_3-N	0.0397	-0.0901	-0.289**	0.215*	0.0125	0.152	-0.0996	0.0443	*r*
	0.703	0.358	0.00273	0.0267	0.899	0.12	0.309	0.652	*p*-value
NH_3-N	-0.105	-0.648**	-0.243*	0.431**	-0.199*	-0.0245	-0.128	-0.0386	*r*
	0.315	0	0.0121	0	0.0405	0.803	0.192	0.694	*p*-value
PO_4	0.177	0.047	-0.00448	0.492**	0.195*	0.174	-0.115	0.107	*r*
	0.0881	0.632	0.964	0	0.0449	0.0739	0.24	0.274	*p*-value
TP	0.164	-0.0382	-0.0374	0.497**	0.158	0.158	-0.0953	0.112	*r*
	0.115	0.697	0.703	0	0.106	0.106	0.331	0.252	*p*-value
chl_a	0.0261	0.177	0.107	0.0663	0.42**	0.216*	0.246*	0.18	*r*
	0.803	0.0688	0.274	0.499	0	0.0264	0.0112	0.0644	*p*-value
AFDM	-0.0869	-0.115	-0.199*	0.0155	0.241*	0.152	0.127	0.219*	*r*
	0.404	0.24	0.0413	0.875	0.013	0.12	0.194	0.0244	*p*-value
R_wid	0.14	0.191*	0.227*	0.084	0.698**	0.269**	0.222*	0.0588	*r*
	0.179	0.0497	0.0193	0.391	0	0.00535	0.0222	0.549	*p*-value
R_len	0.0535	0.368**	0.159	0.0377	0.606**	0.268**	0.229*	0.0265	*r*
	0.608	0	0.104	0.701	0	0.00551	0.0184	0.787	*p*-value
Flow	0.145	0.0652	0.13	-0.0581	0.2*	0.0487	0.0589	-0.178	*r*
	0.163	0.506	0.183	0.554	0.0401	0.619	0.548	0.0672	*p*-value
B_size		0.0984	0.0783	0.215*	0.222*	-0.0196	0.0191	-0.0744	*r*
		0.345	0.452	0.0371	0.0314	0.851	0.854	0.475	*p*-value
DO			0.556**	-0.0358	0.209*	0.0211	0.252**	0.0944	*r*
			0	0.715	0.0315	0.83	0.00918	0.335	*p*-value
pH				0.31**	0.209*	-0.0355	0.328**	-0.014	*r*
				0.00128	0.0316	0.717	0	0.886	*p*-value
Cond					0.0671	0.158	-0.0353	0.0227	*r*
					0.494	0.105	0.719	0.817	*p*-value
% O						0.21*	0.288**	-0.0382	*r*
						0.0307	0.00282	0.697	*p*-value
% DiatC							-0.0925	0.0338	*r*
							0.345	0.731	*p*-value
% ChladC								-0.0608	*r*
								0.535	*p*-value

	% RhizC	% ScenC	% SpirogC	% MerismoC	% OscillC	% AudouC	% DGCov	% LsSCov	
NO_3-N	-0.063	-0.0877	-0.136	0.0638	-0.11	0.136	-0.0363	-0.193*	*r*
	0.52	0.37	0.165	0.515	0.261	0.164	0.712	0.0479	*p*-value
NH_3-N	0.11	0.0303	0.0942	0.0383	0	0.235*	0.114	0.134	*r*
	0.26	0.757	0.336	0.696	1	0.0157	0.242	0.171	*p*-value
PO_4	0.123	-0.00957	-0.0303	-0.0223	0.0502	-0.111	-0.271**	-0.0431	*r*
	0.209	0.922	0.757	0.82	0.609	0.258	0.00513	0.661	*p*-value
TP	0.116	0.0239	-0.0399	0.016	0.0502	-0.0976	-0.22*	-0.0567	*r*
	0.236	0.807	0.684	0.871	0.609	0.319	0.0233	0.563	*p*-value
Chl_a	0.00552	0.0909	-0.107	-0.0144	0.0971	-0.0612	-0.122	-0.186	*r*
	0.955	0.353	0.275	0.884	0.322	0.533	0.213	0.0561	*p*-value
AFDM	-0.0399	0.0941	-0.0813	0.0367	0.0712	0.0507	0.148	-0.159	*r*
	0.684	0.337	0.407	0.708	0.468	0.605	0.13	0.103	*p*-value
R_wid	-0.06	0.0528	-0.165	-0.032	-0.0162	0.0272	-0.213*	-0.234*	*r*
	0.541	0.59	0.0915	0.744	0.869	0.781	0.0289	0.0159	*p*-value
R_len	-0.0562	-0.0112	-0.135	-0.162	-0.0561	-0.0352	-0.171	-0.191*	*r*
	0.567	0.909	0.168	0.0972	0.567	0.72	0.0796	0.0495	*p*-value
Flow	0.116	-0.0802	-0.147	-0.0802	-0.0331	0.0866	0.0526	-0.161	*r*
	0.236	0.413	0.132	0.413	0.736	0.376	0.592	0.099	*p*-value

	% RhizC	% ScenC	% SpirogC	% MerismoC	% OscillC	% AudouC	% DGCov	% LsSCov	
B_size	-0.0936	0.0861	0.0574	-0.119	-0.00389	-0.101	-0.1	0.132	*r*
	0.369	0.409	0.582	0.254	0.97	0.334	0.335	0.203	*p*-value
DO	-0.0254	-0.0942	-0.117	-0.0942	-0.00081	-0.289**	-0.246*	-0.166	*r*
	0.796	0.336	0.234	0.336	0.993	0.00271	0.0113	0.0897	*p*-value
pH	0.0359	-0.0208	-0.0978	-0.0577	0.0195	-0.192*	-0.247*	-0.139	*r*
	0.714	0.832	0.318	0.556	0.842	0.0489	0.0109	0.155	*p*-value
Cond	0.145	0.0526	0.164	0.0207	0.136	-0.0409	-0.287**	0.233*	*r*
	0.137	0.591	0.0922	0.833	0.164	0.677	0.0029	0.0161	*p*-value
% O	0.0708	-0.0383	0.0431	-0.0718	0.0324	-0.0688	-0.0109	-0.0326	*r*
	0.47	0.696	0.66	0.464	0.741	0.483	0.912	0.739	*p*-value
% DiatC	0.0275	0.0193	0.0193	0.0193	0.0392	-0.105	0.0338	0.0275	*r*
	0.78	0.844	0.844	0.844	0.689	0.283	0.731	0.78	*p*-value
% ChladC	-0.0494	-0.0348	-0.0348	-0.0348	-0.0706	-0.0286	-0.0608	-0.0494	*r*
	0.614	0.723	0.723	0.723	0.471	0.771	0.535	0.614	*p*-value
% OegodC	-0.0237	-0.0167	-0.0167	-0.0167	-0.0338	-0.058	-0.0291	-0.0237	*r*
	0.809	0.865	0.865	0.865	0.73	0.554	0.767	0.809	*p*-value
% RhizC		-0.0135	-0.0135	-0.0135	-0.0275	-0.0471	-0.0237	-0.0192	*r*
		0.89	0.89	0.89	0.78	0.631	0.809	0.845	*p*-value
% ScenC			-0.00952	-0.00952	-0.0193	-0.0332	-0.0167	-0.0135	*r*
			0.923	0.923	0.844	0.735	0.865	0.89	*p*-value
% SpirogC				-0.00952	-0.0193	-0.0332	-0.0167	0.697**	*r*
				0.923	0.844	0.735	0.865	0	*p*-value
% Merismop					-0.0193	-0.0332	-0.0167	-0.0135	*r*
					0.844	0.735	0.865	0.89	*p*-value
% OscillC						-0.0673	-0.0338	-0.0275	*r*
						0.493	0.73	0.78	*p*-value
% AudouC							0.113	-0.0471	*r*
							0.25	0.631	*p*-value
% DGCov								-0.0237	*r*
								0.81	*p*-value

	% MdbCov	% BGslCov	% BlgrCov	% BCCov	% ChldCov	% DFilCov	% DTBCov	% GrFthCov	
NO_3-N	-0.0363	0.0307	-0.106	-0.126	0.0231	0.245*	-0.239*	0.0307	*r*
	0.712	0.754	0.279	0.198	0.813	0.0114	0.0138	0.754	*p*-value
NH_3-N	0.114	-0.17	-0.31**	-0.121	-0.00124	0.149	-0.187*	-0.17	*r*
	0.242	0.0809	0.00129	0.215	0.99	0.128	0.0556	0.0809	*p*-value
PO_4	-0.271**	-0.0698	0.16	-0.0973	0.164	0.137	-0.183	-0.0698	*r*
	0.00513	0.477	0.1	0.32	0.0922	0.162	0.0602	0.477	*p*-value
TP	-0.22*	-0.137	0.0527	-0.112	0.137	0.144	-0.16	-0.137	*r*
	0.0233	0.162	0.591	0.254	0.161	0.141	0.102	0.162	*p*-value
chl_a	-0.12	0.205*	0.247*	0.12	0.403**	-0.0177	-0.186	0.205*	*r*
	0.221	0.0347	0.0107	0.222	0	0.857	0.0561	0.0347	*p*-value
AFDM	0.152	0.125	0.0843	-0.123	0.124	0.0469	-0.193*	0.125	*r*
	0.12	0.2	0.39	0.209	0.206	0.632	0.0476	0.2	*p*-value
R_wid	-0.213*	0.227*	0.197*	0.154	0.173	-0.136	0.0784	0.227*	*r*
	0.0289	0.0197	0.0426	0.116	0.076	0.163	0.424	0.0197	*p*-value
R_len	-0.171	0.261**	0.15	0.159	0.197*	-0.188	0.123	0.261**	*r*
	0.0801	0.00706	0.124	0.104	0.0429	0.0543	0.207	0.00706	*p*-value
Flow	0.0526	0.0526	0.192*	0.132	0.175	-0.441**	0.389**	0.0526	*r*
	0.592	0.592	0.0486	0.177	0.0736	0	0	0.592	*p*-value
B_size	-0.1	0.211*	-0.133	-0.0287	0.00971	0.108	-0.117	0.211*	*r*
	0.335	0.0413	0.201	0.783	0.926	0.298	0.261	0.0413	*p*-value
DO	-0.246*	0.262**	0.248*	0.0686	0.103	-0.0579	0.132	0.262**	*r*
	0.0113	0.00669	0.0105	0.484	0.291	0.555	0.176	0.00669	*p*-value

	%MdbCov	%BGslCov	%BlgrCov	%BCCov	%ChldCov	%DFilCov	% DTBCov	%GrFthCov	
pH	-0.247*	0.23*	0.0623	0.0321	0.213*	-0.00503	-0.019	0.23*	*r*
	0.0109	0.0179	0.525	0.744	0.0282	0.959	0.847	0.0179	*p*-value
Cond	-0.287**	-0.0614	-0.0221	-0.059	0.174*	0.276**	-0.3**	-0.0614	*r*
	0.0029	0.531	0.822	0.547	0.074	0.00434	0.00184	0.531	*p*-value
%O	-0.0116	0.194*	0.187	0.148	0.289**	-0.203*	-0.00514	0.194*	*r*
	0.906	0.0458	0.0546	0.129	0.00273	0.0373	0.958	0.0458	*p*-value
%DiatC	0.0338	0.0338	0.189	0.0193	0.0155	0.113	-0.0363	0.0338	*r*
	0.731	0.73	0.0525	0.844	0.875	0.247	0.711	0.73	*p*-value
%ChladC	-0.0608	-0.0608	0.0092	-0.0348	0.316**	-0.13	-0.0553	-0.0608	*r*
	0.535	0.535	0.925	0.723	0.00101	0.184	0.573	0.535	*p*-value
%OegodC	-0.0291	-0.0291	0.0995	-0.0167	-0.132	0.0519	0.0268	-0.0291	*r*
	0.767	0.767	0.31	0.865	0.178	0.597	0.785	0.767	*p*-value
%RhizC	-0.0237	-0.0237	0.105	-0.0135	0.188	-0.0543	-0.0603	-0.0237	*r*
	0.809	0.809	0.283	0.89	0.0542	0.58	0.539	0.809	*p*-value
%ScenC	-0.0167	-0.0167	0.0474	-0.00952	0.0116	-0.0207	0.0399	-0.0167	*r*
	0.865	0.865	0.629	0.923	0.906	0.833	0.684	0.865	*p*-value
%SpirogC	-0.0167	-0.0167	-0.0931	-0.00952	-0.104	0.0798	0.0224	-0.0167	*r*
	0.865	0.865	0.342	0.923	0.288	0.416	0.82	0.865	*p*-value
%Merismop	-0.0167	-0.0167	0.0474	-0.00952	-0.104	0.0016	0.0815	-0.0167	*r*
	0.865	0.865	0.629	0.923	0.288	0.987	0.406	0.865	*p*-value
%OscillC	-0.0338	-0.0338	0.254**	-0.0193	0.0302	0.0138	-0.214*	-0.0338	*r*
	0.73	0.73	0.00867	0.844	0.759	0.888	0.0278	0.73	*p*-value
%AudouC	0.116	-0.058	0.0853	-0.0332	0.041	-0.1	0.0907	-0.058	*r*
	0.236	0.554	0.384	0.735	0.676	0.307	0.354	0.554	*p*-value
%DGCov	1**	-0.0291	-0.163	-0.0167	-0.182	-0.287**	0.26**	-0.0291	*r*
	0	0.767	0.0953	0.865	0.062	0.00291	0.00734	0.767	*p*-value
%LsSCov	-0.0237	-0.0237	-0.132	-0.0135	-0.148	0.164	-0.0897	-0.0237	*r*
	0.809	0.809	0.176	0.89	0.13	0.0936	0.36	0.809	*p*-value
%MdbCov		-0.0291	-0.163	-0.0167	-0.182	-0.287**	0.26**	-0.0291	*r*
		0.767	0.0953	0.865	0.062	0.00291	0.00726	0.767	*p*-value
%BGslCov			-0.163	-0.0167	-0.182	0.114	0.00559	1**	*r*
			0.0952	0.865	0.062	0.243	0.955	0	*p*-value
%BlgrCov				0.0897	0.358**	-0.405**	0.131	-0.163	*r*
				0.36	0	0	0.18	0.0952	*p*-value
%BCCov					0.134	-0.142	0.104	-0.0167	*r*
					0.171	0.146	0.289	0.865	*p*-value
%ChldCov						-0.285**	-0.129	-0.182	*r*
						0.0032	0.187	0.062	*p*-value
%DFilCov							-0.766**	0.114	*r*
							0	0.243	*p*-value
% DTBCov								0.00559	*r*
								0.955	*p*-value

	%GrF1Cov	%GrF2Cov	%GFBCov	%GrFCov	%GrFsCov	%HBCov	%LlGCov	%OrgCov	
NO_3-N	0.155	0.22*	0.176	-0.0377	0.344**	-0.0703	0.287**	0.0814	*r*
	0.113	0.0237	0.0716	0.7	0	0.473	0.00291	0.406	*p*-value
NH_3-N	0.126	0.179	0.204*	0.265**	-0.0983	0.12	-0.0865	0.075	*r*
	0.197	0.0661	0.0363	0.00612	0.316	0.219	0.377	0.444	*p*-value
PO_4	0.164	0.234*	0.259**	-0.117	-0.014	-0.0771	0.0419	0.129	*r*
	0.0922	0.0161	0.00736	0.234	0.886	0.432	0.67	0.186	*p*-value
TP	0.164	0.234*	0.243*	-0.0638	-0.0816	-0.0975	-0.053	0.129	*r*
	0.0922	0.0161	0.0123	0.515	0.405	0.32	0.589	0.186	*p*-value

	%GrF1Cov	%GrF2Cov	%GFBCov	%GrFCov	%GrFsCov	%HBCov	%LlGCov	%OrgCov	
chl_a	0.164	0.22*	-0.0141	0.144	0.0274	-0.00906	-0.0574	0.0271	***r***
	0.0924	0.0238	0.886	0.14	0.78	0.926	0.558	0.782	***p*-value**
AFDM	0.155	0.195*	0.0263	0.194*	0.246*	0.0748	-0.0234	-0.0718	***r***
	0.113	0.0454	0.789	0.0465	0.0111	0.445	0.811	0.464	***p*-value**
R_wid	0.0816	0.116	0.176	-0.0618	-0.0251	-0.111	-0.246*	0.024	***r***
	0.405	0.236	0.0709	0.528	0.798	0.255	0.0111	0.807	***p*-value**
R_len	0.085	0.0706	0.0869	-0.206*	0.0912	-0.191*	0.192*	-0.0112	***r***
	0.386	0.471	0.375	0.0342	0.352	0.0495	0.0484	0.909	***p*-value**
Flow	0.132	-0.0364	0.295**	-0.188	-0.285**	-0.0356	-0.257**	0.132	***r***
	0.177	0.711	0.0022	0.0535	0.00311	0.716	0.00791	0.177	***p*-value**
B_size	0.136	0.193	0.191	-0.322**	-0.162	-0.19	--	-0.109	***r***
	0.191	0.0623	0.0654	0.00161	0.119	0.0661	--	0.295	***p*-value**
DO	-0.126	-0.179	-0.0726	-0.424**	0.0411	-0.0907	0.162	-0.164	***r***
	0.197	0.0661	0.459	0	0.675	0.354	0.0972	0.092	***p*-value**
pH	-0.0721	-0.103	-0.126	-0.203*	-0.189	0.144	-0.0364	-0.141	***r***
	0.462	0.295	0.197	0.0365	0.0527	0.142	0.71	0.149	***p*-value**
Cond	0.142	0.202*	0.0139	-0.0291	-0.115	0.215*	0.215*	-0.107	***r***
	0.146	0.0382	0.887	0.766	0.239	0.0267	0.0272	0.275	***p*-value**
%O	0.0591	0.111	0.0991	-0.203*	-0.166	-0.11	-0.101	-0.0383	***r***
	0.547	0.256	0.312	0.0372	0.0893	0.261	0.304	0.696	***p*-value**
%DiatC	0.0193	0.0275	0.0338	0.0831	0.0485	0.0275	0.0338	0.0193	***r***
	0.844	0.78	0.731	0.396	0.621	0.78	0.73	0.844	***p*-value**
%ChladC	-0.0348	-0.0494	-0.0608	-0.0113	-0.0873	-0.0494	-0.0608	-0.0348	***r***
	0.723	0.614	0.535	0.908	0.373	0.614	0.535	0.723	***p*-value**
%OegodC	-0.0167	-0.0237	-0.0291	0.0982	0.202*	-0.0237	-0.0291	-0.0167	***r***
	0.865	0.809	0.767	0.316	0.0382	0.809	0.767	0.865	***p*-value**
%RhizC	-0.0135	-0.0192	-0.0237	0.113	-0.0339	-0.0192	-0.0237	-0.0135	***r***
	0.89	0.845	0.809	0.248	0.729	0.845	0.809	0.89	***p*-value**
%ScenC	-0.00952	-0.0135	-0.0167	0.264**	-0.0239	-0.0135	-0.0167	-0.00952	***r***
	0.923	0.89	0.865	0.00642	0.808	0.89	0.865	0.923	***p*-value**
%SpirogC	-0.00952	-0.0135	-0.0167	-0.041	-0.0239	-0.0135	-0.0167	-0.00952	***r***
	0.923	0.89	0.865	0.676	0.808	0.89	0.865	0.923	***p*-value**
%Merismop	-0.00952	-0.0135	-0.0167	0.23*	-0.0239	-0.0135	-0.0167	-0.00952	***r***
	0.923	0.89	0.865	0.0176	0.808	0.89	0.865	0.923	***p*-value**
%OscillC	-0.0193	-0.0275	-0.0338	0.074	-0.0485	0.336**	-0.0338	-0.0193	***r***
	0.844	0.78	0.73	0.45	0.621	0	0.73	0.844	***p*-value**
%AudouC	-0.0332	-0.0471	0.135	0.0959	-0.0832	0.199*	-0.058	0.265**	***r***
	0.735	0.631	0.166	0.328	0.396	0.0408	0.554	0.00612	***p*-value**
%DGCov	-0.0167	-0.0237	-0.0291	-0.0716	-0.0418	-0.0237	-0.0291	-0.0167	***r***
	0.865	0.809	0.767	0.465	0.67	0.809	0.767	0.865	***p*-value**
%LsSCov	-0.0135	-0.0192	-0.0237	-0.0582	-0.0339	-0.0192	-0.0237	-0.0135	***r***
	0.89	0.845	0.809	0.553	0.729	0.845	0.809	0.89	***p*-value**
%MdbCov	-0.0167	-0.0237	-0.0291	-0.0716	-0.0418	-0.0237	-0.0291	-0.0167	***r***
	0.865	0.809	0.767	0.465	0.67	0.809	0.767	0.865	***p*-value**
%BGslCov	-0.0167	-0.0237	-0.0291	-0.0716	-0.0418	-0.0237	-0.0291	-0.0167	***r***
	0.865	0.809	0.767	0.465	0.67	0.809	0.767	0.865	***p*-value**
%BlgrCov	0.0152	0.0131	-0.0352	0.124	0.0415	0.17	-0.163	0.0474	***r***
	0.877	0.894	0.72	0.206	0.672	0.0821	0.0953	0.629	***p*-value**
%BCCov	-0.00952	-0.0135	-0.0167	-0.041	-0.0239	-0.0135	-0.0167	-0.00952	***r***
	0.923	0.89	0.865	0.676	0.808	0.89	0.865	0.923	***p*-value**
%ChldCov	0.0908	0.209	-0.182	0.04	-0.222	0.0774	-0.0103	0.0908	***r***
	0.354	0.0316	0.062	0.683	0.0224	0.429	0.916	0.354	***p*-value**

	%GrF1Cov	%GrF2Cov	%GFBCov	%GrFCov	%GrFsCov	%HBCov	%LlGCov	%OrgCov	
%DFilCov	-0.0925	-0.005	-0.135	0.0641	0.0977	0.0544	0.169	0.0207	***r***
	0.345	0.959	0.166	0.513	0.318	0.579	0.084	0.833	***p*-value**
% DTBCov	0.0607	-0.209*	0.213*	-0.142	-0.0151	-0.209*	-0.121	-0.0527	***r***
	0.536	0.0318	0.0283	0.145	0.878	0.0318	0.218	0.591	***p*-value**
%GrFthCov	-0.0167	-0.0237	-0.0291	-0.0716	-0.0418	-0.0237	-0.0291	-0.0167	***r***
	0.865	0.809	0.767	0.465	0.67	0.809	0.767	0.865	***p*-value**
%GrF1Cov		-0.0135	-0.0167	-0.041	-0.0239	-0.0135	-0.0167	-0.00952	***r***
		0.89	0.865	0.676	0.808	0.89	0.865	0.923	***p*-value**
%GrF2Cov			-0.0237	-0.0582	-0.0339	-0.0192	-0.0237	-0.0135	***r***
			0.809	0.553	0.729	0.845	0.809	0.89	***p*-value**
%GFBCov				-0.0716	-0.0418	-0.0237	-0.0291	-0.0167	***r***
				0.465	0.67	0.809	0.767	0.865	***p*-value**
%GrFCov					-0.103	0.135	-0.0716	0.192	***r***
					0.294	0.169	0.465	0.0487	***p*-value**
%GrFsCov						-0.0339	-0.0418	-0.0239	***r***
						0.729	0.67	0.808	***p*-value**
%HBCov							-0.0237	-0.0135	***r***
							0.809	0.89	***p*-value**
%LlGCov								-0.0167	***r***
								0.865	***p*-value**

	%TgLCov	%TsCdCov	%GrBGCov	%Bed	%Bo	%Cob	%Gra	%Si/Cl	
NO_3-N	0.287**	-0.187	0.0398	0.363**	0.0207	-0.122	0.112	0.064	***r***
	0.00291	0.0551	0.685	0	0.833	0.212	0.254	0.514	***p*-value**
NH_3-N	-0.0865	-0.0698	0.104	-0.156	-0.0984	-0.153	-0.206*	0.202*	***r***
	0.377	0.477	0.29	0.111	0.315	0.117	0.0341	0.0382	***p*-value**
PO_4	0.0418	0.0251	0.0974	0.106	0.0829	0.0909	0.316**	-0.197*	***r***
	0.67	0.798	0.32	0.279	0.398	0.353	0.00102	0.0428	***p*-value**
TP	-0.053	-0.0195	0.131	0.00418	0.0142	0.0738	0.285**	-0.23*	***r***
	0.589	0.842	0.182	0.966	0.885	0.452	0.00317	0.0179	***p*-value**
chl_a	-0.0594	-0.0206	-0.0315	-0.0101	-0.0832	0.151	0.33**	-0.141	***r***
	0.545	0.833	0.748	0.918	0.396	0.122	0	0.15	***p*-value**
AFDM	-0.026	-0.0634	0.00935	0.0644	-0.257**	-0.0434	0.158	0.0904	***r***
	0.791	0.518	0.924	0.511	0.00802	0.658	0.106	0.356	***p*-value**
R_wid	-0.246*	0.268**	0.00733	-0.0955	-0.00311	0.277**	0.283**	-0.338**	***r***
	0.0111	0.00551	0.94	0.33	0.975	0.00418	0.00342	0	***p*-value**
R_len	0.193*	0.239*	-0.254**	0.275**	-0.0621	0.0957	0.344**	-0.29**	***r***
	0.0478	0.0139	0.00885	0.00437	0.527	0.329	0	0.00261	***p*-value**
Flow	-0.257**	0.111	0.00761	-0.245*	0.231*	0.382**	0.063	-0.33**	***r***
	0.00791	0.255	0.938	0.0114	0.0174	0	0.521	0	***p*-value**
B_size	--	--	-0.00764	-0.273**	0.168	0.341**	0.0748**	-0.17	***r***
	--	--	0.942	0.00788	0.106	0	0.473	0.101	***p*-value**
DO	0.162	-0.112	-0.174	0.237*	0.125	0.0723	0.198	-0.17	***r***
	0.0972	0.254	0.0748	0.0144	0.203	0.461	0.0415	0.0814	***p*-value**
pH	-0.0364	0.255**	-0.0696	-0.061	0.279**	0.308**	-0.036	-0.176	***r***
	0.71	0.00844	0.478	0.534	0.00391	0.00139	0.713	0.0705	***p*-value**
Cond	0.215*	0.0363	-0.0268	0.0257	0.0915	-0.025	0.0638	0.0412	***r***
	0.0272	0.712	0.785	0.793	0.35	0.798	0.515	0.675	***p*-value**
%O	-0.102	0.172	-0.0248	-0.123	-0.0575	0.16	0.231*	-0.192*	***r***
	0.296	0.0774	0.8	0.207	0.557	0.101	0.0175	0.0485	***p*-value**
%DiatC	0.0338	0.0338	-0.254**	0.0565	-0.0699	-0.0484	0.229*	-0.197*	***r***
	0.731	0.73	0.00887	0.564	0.476	0.622	0.0181	0.0431	***p*-value**

	%TgLCov	%TsCdCov	%GrBGCov	%Bed	%Bo	%Cob	%Gra	%S	
%ChladC	-0.0608	0.118	-0.0608	-0.102	-0.0802	0.203*	0.0744	-0.109	***r***
	0.535	0.227	0.535	0.298	0.413	0.037	0.448	0.264	***p*-value**
%OegodC	-0.0291	-0.0291	-0.0291	0.166*	-0.0403	-0.0283	0.149	-0.0677	***r***
	0.767	0.767	0.767	0.0882	0.681	0.773	0.126	0.49	***p*-value**
%RhizC	-0.0237	-0.0237	-0.0237	-0.0396	-0.0515	-0.0902	0.0631	0.137	***r***
	0.809	0.809	0.809	0.686	0.6	0.357	0.52	0.161	***p*-value**
%ScenC	-0.0167	-0.0167	-0.0167	-0.0279	-0.123	0.0752	0.139	-0.102	***r***
	0.865	0.865	0.865	0.776	0.208	0.443	0.154	0.296	***p*-value**
%SpirogC	-0.0167	-0.0167	-0.0167	-0.0279	-0.00811	-0.131	-0.096	0.141	***r***
	0.865	0.865	0.865	0.776	0.934	0.18	0.327	0.15	***p*-value**
%Merismop	-0.0167	-0.0167	-0.0167	-0.0279	-0.00811	-0.04	0.096	0.0479	***r***
	0.865	0.865	0.865	0.776	0.934	0.684	0.327	0.625	***p*-value**
%OscillC	-0.0338	-0.0338	-0.0338	-0.0565	-0.0642	-0.026	0.0795	0.0454	***r***
	0.73	0.73	0.73	0.564	0.513	0.791	0.417	0.644	***p*-value**
%AudouC	-0.058	-0.058	0.14	-0.097	0.156	0.126	-0.21	0.0149	***r***
	0.554	0.554	0.151	0.322	0.11	0.196	0.031	0.879	***p*-value**
%DGCov	-0.0291	-0.0291	-0.0291	-0.0487	-0.215*	-0.258**	-0.187	0.274**	***r***
	0.767	0.767	0.767	0.619	0.0267	0.00769	0.0553	0.00464	***p*-value**
%LsSCov	-0.0237	-0.0237	-0.0237	-0.0396	0.106	-0.0705	-0.136	0.0256	***r***
	0.809	0.809	0.809	0.686	0.279	0.472	0.163	0.794	***p*-value**
%MdbCov	-0.0291	-0.0291	-0.0291	-0.0487	-0.215*	-0.258**	-0.188	0.274**	***r***
	0.767	0.767	0.767	0.619	0.0267	0.00771	0.0536	0.00461	***p*-value**
%BGslCov	-0.0291	-0.0291	-0.0291	-0.0487	0.17	0.131	0.0476	-0.129	***r***
	0.767	0.767	0.767	0.619	0.0812	0.179	0.628	0.189	***p*-value**
%BlgrCov	-0.163	-0.163	-0.163	0.0515	0.00911	0.172	0.209*	-0.0666	***r***
	0.0953	0.0953	0.0953	0.6	0.926	0.0781	0.0319	0.497	***p*-value**
%BCCov	-0.0167	-0.0167	-0.0167	-0.0279	0.123	-0.0624	0.056	0.00799	***r***
	0.865	0.865	0.865	0.776	0.208	0.525	0.568	0.935	***p*-value**
%ChldCov	-0.0108	-0.126	-0.00223	-0.064	-0.0631	0.19	0.199*	0.0191	***r***
	0.913	0.199	0.982	0.514	0.52	0.0506	0.0413	0.846	***p*-value**
%DfilCov	0.168	-0.143	0.205*	0.175	0.0903	0.00391	-0.209*	0.0564	***r***
	0.0853	0.143	0.0355	0.072	0.356	0.968	0.032	0.565	***p*-value**
% DTBCov	-0.12	0.0617	-0.257**	-0.113	0.056	-0.0635	0.15	-0.163	***r***
	0.222	0.529	0.00796	0.248	0.568	0.517	0.125	0.0942	***p*-value**
%GrFthCov	-0.0291	-0.0291	-0.0291	-0.0487	0.17	0.131	0.0476	-0.129	***r***
	0.767	0.767	0.767	0.619	0.0812	0.179	0.628	0.189	***p*-value**
%GrF1Cov	-0.0167	-0.0167	-0.0167	-0.0279	0.133	0.118	0.0272	-0.136	***r***
	0.865	0.865	0.865	0.776	0.174	0.227	0.782	0.165	***p*-value**
%GrF2Cov	-0.0237	-0.0237	-0.0237	-0.0396	-0.134	-0.161	-0.0214	0.182	***r***
	0.809	0.809	0.809	0.686	0.17	0.0999	0.827	0.0623	***p*-value**
%GFBCov	-0.0291	-0.0291	-0.0291	-0.0487	0.172	0.0893	0.0947	-0.269**	***r***
	0.767	0.767	0.767	0.619	0.0779	0.362	0.334	0.00541	***p*-value**
%GrFCov	-0.0716	-0.0716	-0.0716	-0.12	-0.202	-0.034	-0.0224	0.0908	***r***
	0.465	0.465	0.465	0.221	0.0379	0.729	0.819	0.354	***p*-value**
%GrFsCov	-0.0418	-0.0418	-0.0418	0.372**	0.0195	-0.162	0.114	0.0896	***r***
	0.67	0.67	0.67	0	0.843	0.0974	0.244	0.361	***p*-value**
%HBCov	-0.0237	-0.0237	-0.0237	-0.0396	0.0369	-0.00682	-0.156	0.162	***r***
	0.809	0.809	0.809	0.686	0.707	0.945	0.111	0.0963	***p*-value**
%LlGCov	1**	-0.0291	-0.0291	0.585**	-0.0142	-0.185	0.0984	0.0345	***r***
	0	0.767	0.767	0	0.885	0.0583	0.315	0.725	***p*-value**
%OrgCov	-0.0167	-0.0167	-0.0167	-0.0279	0.0973	0.162	-0.0304	-0.136	***r***
	0.865	0.865	0.865	0.776	0.321	0.098	0.757	0.165	***p*-value**

	%TgLCov	%TsCdCov	%GrBGCov	%Bed	%Bo	%Cob	%Gra	%S	
%TgLCov		-0.0291	-0.0291	0.585**	-0.0142	-0.185	0.097	0.0365	***r***
		0.767	0.767	0	0.885	0.0579	0.322	0.71	***p*-value**
%TsCdCov			-0.0291	-0.0487	-0.0354	0.0987	0.0813	-0.0156	***r***
			0.767	0.619	0.718	0.314	0.407	0.874	***p*-value**
%GrBGCov				-0.0487	0.118	0.103	-0.113	0.0244	***r***
				0.619	0.227	0.291	0.248	0.803	***p*-value**
%Bed					0.0432	-0.219*	0.0831	-0.0356	***r***
					0.66	0.0242	0.396	0.717	***p*-value**
%Bo						0.328**	-0.313**	-0.317**	***r***
						0	0.00116	0	***p*-value**
%Cob							0.0507	-0.53**	***r***
							0.605	0	***p*-value**
%Gra								-0.417**	***r***
								0	***p*-value**

	%Si/Cl	BRBDCOG	SDSTCL	BRBD	CBGR	
NO_3-N	0.0177	-0.0571	0.0493	0.0797	-0.00966	***r***
	0.857	0.561	0.615	0.416	0.922	***p*-value**
NH_3-N	-0.105	-0.22*	0.224*	-0.124	-0.196*	***r***
	0.281	0.0234	0.0212	0.206	0.0446	***p*-value**
PO_4	-0.0434	0.233*	-0.235*	0.0938	0.259**	***r***
	0.658	0.0162	0.0156	0.338	0.00756	***p*-value**
TP	-0.049	0.223*	-0.223*	0.0114	0.252**	***r***
	0.618	0.0215	0.0218	0.908	0.0094	***p*-value**
chl_a	-0.113	0.226*	-0.211*	-0.0845	0.294**	***r***
	0.248	0.02	0.0298	0.389	0.00229	***p*-value**
AFDM	-0.0469	-0.0693	0.0785	-0.245*	0.115	***r***
	0.632	0.479	0.423	0.0115	0.241	***p*-value**
R_wid	0.00415	0.358**	-0.356**	-0.00729	0.413**	***r***
	0.966	0	0	0.941	0	***p*-value**
R_len	0.216	0.23*	-0.226*	0.000828	0.333**	***r***
	0.026	0.0181	0.0197	0.993	0	***p*-value**
Flow	-0.311**	0.37**	-0.373**	0.188	0.261**	***r***
	0.00123	0	0	0.0539	0.00711	***p*-value**
B_size	-0.235*	0.274**	-0.271**	0.117	0.248*	***r***
	0.023	0.00757	0.00849	0.262	0.0162	***p*-value**
DO	0.27**	0.159	-0.149	0.16	0.163	***r***
	0.00516	0.104	0.128	0.101	0.0945	***p*-value**
pH	0.117	0.267**	-0.255**	0.264**	0.154	***r***
	0.233	0.00576	0.00844	0.00634	0.114	***p*-value**
COND	-0.0148	0.0226	-0.0109	0.0908	-0.00373	***r***
	0.88	0.818	0.912	0.354	0.97	***p*-value**
%O	0.0396	0.249*	-0.217*	-0.0702	0.262**	***r***
	0.687	0.0101	0.0255	0.474	0.00688	***p*-value**
%DiatC	-0.182	0.164	-0.178	-0.0517	0.141	***r***
	0.0614	0.0924	0.0683	0.598	0.15	***p*-value**
%ChladC	0.103	0.135	-0.126	-0.0899	0.217*	***r***
	0.294	0.168	0.198	0.359	0.0254	***p*-value**
%OegodC	0.0502	0.032	-0.0302	-0.0132	0.13	***r***
	0.608	0.744	0.758	0.893	0.185	***p*-value**
%RhizC	-0.0838	-0.0855	0.115	-0.0583	-0.104	***r***
	0.393	0.383	0.239	0.552	0.288	***p*-value**

	% Si/Cl	BRBDCOG	SDSTCL	BRBD	CBGR	
% ScenC	-0.059	0.113	-0.113	-0.123	0.155	***r***
	0.548	0.247	0.247	0.209	0.113	***p*-value**
% SpirogC	-0.059	-0.134	0.134	-0.0178	-0.144	***r***
	0.548	0.17	0.17	0.856	0.142	***p*-value**
% Merismop	-0.059	-0.0287	0.024	-0.0178	-0.112	***r***
	0.548	0.77	0.807	0.856	0.254	***p*-value**
% OscillC	-0.12	-0.0154	0.013	-0.0755	0.0769	***r***
	0.221	0.875	0.895	0.441	0.432	***p*-value**
% AudouC	-0.205*	0.0384	-0.0411	0.135	-0.0396	***r***
	0.0349	0.696	0.675	0.168	0.686	***p*-value**
% DGCov	-0.103	-0.266**	0.266**	-0.215*	-0.259**	***r***
	0.292	0.00598	0.00595	0.0269	0.0074	***p*-value**
% LsSCov	-0.0838	-0.0131	0.0131	0.0989	-0.143	***r***
	0.393	0.894	0.894	0.313	0.142	***p*-value**
% MdbCov	-0.103	-0.266**	0.266**	-0.215*	-0.26**	***r***
	0.292	0.00592	0.0059	0.0269	0.00729	***p*-value**
% BGslCov	-0.103	0.159	-0.154	0.161	0.109	***r***
	0.292	0.103	0.116	0.0984	0.266	***p*-value**
% BlgrCov	-0.159	0.151	-0.15	0.0118	0.256**	***r***
	0.102	0.121	0.124	0.904	0.00814	***p*-value**
% BCCov	0.138	-0.0128	0.024	0.117	-0.0415	***r***
	0.157	0.896	0.807	0.234	0.672	***p*-value**
% ChldCov	-0.134	0.0704	-0.0583	-0.077	0.226*	***r***
	0.171	0.473	0.552	0.432	0.0201	***p*-value**
% DFilCov	-0.0338	-0.0318	0.0108	0.123	-0.128	***r***
	0.731	0.746	0.912	0.209	0.191	***p*-value**
% DTBCov	0.0718	0.0917	-0.0893	0.0344	0.0454	***r***
	0.464	0.349	0.362	0.726	0.643	***p*-value**
% GrFthCov	-0.103	0.159	-0.154	0.161	0.109	***r***
	0.292	0.103	0.116	0.0984	0.266	***p*-value**
% GrF1Cov	-0.059	0.137	-0.142	0.126	0.101	***r***
	0.548	0.16	0.146	0.197	0.304	***p*-value**
% GrF2Cov	-0.0838	-0.173	0.173	-0.134	-0.137	***r***
	0.393	0.077	0.0769	0.171	0.16	***p*-value**
% GFBCov	-0.103	0.273**	-0.278**	0.162	0.118	***r***
	0.292	0.00481	0.00398	0.0963	0.226	***p*-value**
% GrFCov	-0.179	-0.0688	0.0576	-0.214*	-0.0178	***r***
	0.0657	0.483	0.557	0.0275	0.856	***p*-value**
% GrFsCov	0.207*	-0.123	0.121	0.0779	-0.0232	***r***
	0.033	0.21	0.215	0.427	0.813	***p*-value**
% HBCov	-0.0838	-0.141	0.138	0.0218	-0.11	***r***
	0.392	0.15	0.157	0.824	0.261	***p*-value**
% LlGCov	0.3**	-0.101	0.102	0.0849	-0.088	***r***
	0.00186	0.303	0.298	0.386	0.369	***p*-value**
% OrgCov	-0.059	0.137	-0.142	0.0923	0.128	***r***
	0.548	0.16	0.146	0.346	0.192	***p*-value**
% TgLCov	0.3**	-0.103	0.104	0.0849	-0.0895	***r***
	0.00183	0.295	0.29	0.386	0.361	***p*-value**
% TsCdCov	0.108	0.0339	-0.0236	-0.0392	0.0998	***r***
	0.27	0.73	0.81	0.689	0.308	***p*-value**
% GrBGCov	-0.103	0.00715	-0.0118	0.102	-0.0233	***r***
	0.292	0.942	0.905	0.298	0.813	***p*-value**
% Bed	0.324**	-0.0638	0.0371	0.22*	-0.0885	***r***
	0	0.515	0.705	0.0238	0.367	***p*-value**

	% Si/Cl	BRBDCOG	SDSTCL	BRBD	CBGR	
% Bo	-0.19	0.42**	-0.422**	0.983**	-0.0829	***r***
	0.0512	0	0	0	0.398	***p*-value**
% Cob	-0.269**	0.677**	-0.669**	0.278**	0.641**	***r***
	0.00534	0	0	0.00396	0	***p*-value**
% Gra	0.0801	0.407**	-0.384**	-0.3**	0.66**	***r***
	0.414	0	0	0.00184	0	***p*-value**
% S	-0.00147	-0.878**	0.892**	-0.328**	-0.603**	***r***
	0.988	0	0	0	0	***p*-value**
% Si/Cl		-0.286**	0.299**	-0.137	-0.0783	***r***
		0.00307	0.0019	0.161	0.424	***p*-value**
BRBDCOG			-0.986**	0.407**	0.674**	***r***
			0	0	0	***p*-value**
SDSTCL				-0.417**	-0.656**	***r***
				0	0	***p*-value**
BRBD					-0.102	***r***
					0.295	***p*-value**
CBGR						***r***

Appendix 4: Results of Forward stepwise regression for dependent variables Chl *a* and AFDM. Abbreviations used for variables are the same as listed under Notes in Appendix 3.

a) Forward Stepwise Regression: Dependent Variable: chl_a

F-to-Enter: 4.000 P = 0.048
F-to-Remove: 3.900 P = 0.051

Step 0:
Standard Error of Estimate = 0.380

Analysis of Variance:

Group	DF	SS	MS	F	P
Residual	105	15.130	0.144		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	1.879		0.0369		

Variables not in Model

Group	F-to-Enter	P
NH_3-N	0.275	0.601
PO4	7.278	0.008
TP	5.272	0.024
NO_3-N	9.949	0.002
DO	0.00381	0.951
pH	1.534	0.218
COND	3.718	0.057
B_size	31.019	<0.001
% O	30.218	<0.001
%Bed	0.0771	0.782
%Bo	0.0157	0.901
%Cob	3.969	0.049
%Gra	6.569	0.012
%S	1.716	0.193
%Si/Cl	4.349	0.039
BBDCOGR	7.793	0.006
SDSTCL	5.617	0.020
BRBD	0.00291	0.957
CBGR	6.045	0.016
R_wid	19.025	<0.001
R_len	13.716	<0.001
Flow	0.298	0.587

Step 1: BASIN Entered

R = 0.479 Rsqr = 0.230 Adj Rsqr = 0.222
Standard Error of Estimate = 0.335

Analysis of Variance:

Group	DF	SS	MS	F	P
Regression	1	3.476	3.476	31.019	<0.001
Residual	104	11.654	0.112		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	1.259		0.116		
B_size	0.421	0.479	0.0756	31.019	<0.001

Variables not in Model

Group	F-to-Enter	P
NH_3-N	0.113	0.738
PO4	1.418	0.236
TP	0.941	0.334
NO_3-N	9.104	0.003
DO	0.951	0.332
pH	0.351	0.555
COND	3.643	0.059
%O	0.245	0.622
%Bed	0.00813	0.928
%Bo	0.292	0.590
%Cob	3.126	0.080
%Gra	3.266	0.074
%S	0.517	0.474
%Si/Cl	5.433	0.022
BRBDCOGR	6.164	0.015
SDSTCL	3.127	0.080
BRBD	0.275	0.601
CBGR	4.119	0.045
R_wid	1.014	0.316
R_len	0.900	0.345
Flow	0.0631	0.802

Step 2: NO_3-N Entered

R = 0.541 Rsqr = 0.292 Adj Rsqr = 0.279

Standard Error of Estimate = 0.322

Analysis of Variance:

Group	DF	SS	MS	F	P
Regression	2	4.423	2.211	21.270	<0.001
Residual	103	10.708	0.104		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	0.407		0.304		
NO_3-N	0.285	0.251	0.0943	9.104	0.003
B_size	0.400	0.455	0.0732	29.833	<0.001

Variables not in Model

Group	F-to-Enter	P
NH_3-N	0.445	0.506
PO4	0.0309	0.861
TP	0.115	0.735
DO	0.344	0.559
pH	3.190	0.077
COND	1.712	0.194

%O	0.765	0.384
%Bed	0.636	0.427
%Bo	1.348	0.248
%Cob	6.115	0.015
%Gra	2.012	0.159
%S	1.563	0.214
%Si/Cl	6.686	0.011
BRBDCOGR	10.180	0.002
SDSTCL	5.350	0.023
BRBD	0.942	0.334
CBGR	4.708	0.032
R_wid	0.539	0.465
R_len	0.197	0.658
Flow	0.135	0.714

Step 3: BRBDCOGR Entered

R = 0.597 Rsqr = 0.357 Adj Rsqr = 0.338

Standard Error of Estimate = 0.309

Analysis of Variance:

Group	**DF**	**SS**	**MS**	**F**	**P**
Regression	3	5.394	1.798	18.837	<0.001
Residual	102	9.736	0.0955		

Variables in Model

Group	**Coef.**	**Std. Coeff.**	**Std. Error**	**F-to-Remove**	**P**
Constant	-0.156		0.340		
NO_3-N	0.333	0.294	0.0917	13.201	<0.001
B_size	0.369	0.420	0.0708	27.187	<0.001
BRBDCOGR	0.0566	0.259	0.0177	10.180	0.002

Variables not in Model

Group	**F-to-Enter**	**P**
NH_3-N	0.0289	0.865
PO4	1.144	0.287
TP	0.720	0.398
DO	1.460	0.230
pH	0.672	0.414
COND	1.251	0.266
%O	0.455	0.501
%Bed	1.365	0.245
%Bo	0.0409	0.840
%Cob	0.00237	0.961
%Gra	0.274	0.602
%S	2.654	0.106
%Si/Cl	2.690	0.104
SDSTCL	1.172	0.282
BRBD	0.185	0.668
CBGR	0.852	0.358
R_wid	0.0434	0.835
R_len	0.0343	0.854
Flow	0.682	0.411

Summary Table

Step #	Vars. Entered	Vars. Removed	R	RSqr	Delta RSqr	Vars in Model
1	B_size		0.479	0.230	0.230	1
2	NO_3-N		0.541	0.292	0.0626	2
3	BRBDCOGR		0.597	0.357	0.0642	3

The dependent variable chl_a can be predicted from a linear combination of the independent variables:

	P
NO_3-N	**<0.001**
B_size	**<0.001**
BRBDCOGR	**0.002**

The following variables did not significantly add to the ability of the equation to predict chl_a and were not included in the final equation:

NH_3-N PO4 TP DO pH COND %O %Bed %Bo %Cob %Gra %S %Si/Cl SDSTCL BRBD CBGR R_wid R_len Flow

Normality Test: Passed (P = 0.657)
Constant Variance Test: Passed (P = 1.000)
Power of performed test with alpha = 0.050: 1.000

Appendix 4

b) Forward Stepwise Regression: Dependent Variable: AFDM
F-to-Enter: 4.000 P = 0.048
F-to-Remove: 3.900 P = 0.051

Step 0:
Standard Error of Estimate = 0.332

Analysis of Variance:

Group	DF	SS	MS	F	P
Residual	105	11.561	0.110		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	1.165		0.0322		

Variables not in Model

Group	F-to-Enter	P
NH_3-N	1.382	0.242
PO4	1.970	0.163
TP	2.534	0.114
NO_3-N	18.620	<0.001
DO	1.155	0.285
pH	3.669	0.058
COND	0.00651	0.936
B_size	6.616	0.012
%O	5.824	0.018
%Bed	0.263	0.609
%Bo	3.708	0.057
%Cob	0.824	0.366
%Gra	1.132	0.290
%S	0.804	0.372
%Si/Cl	0.00152	0.969
BRBDCOGR	0.540	0.464
SDSTCL	0.591	0.444
BRBD	3.357	0.070
CBGR	0.0408	0.840
R_wid	5.040	0.027
R_leng	4.552	0.035
Flow	0.734	0.394

Step 1: NO_3-N Entered
R = 0.390 Rsqr = 0.152 Adj Rsqr = 0.144
Standard Error of Estimate = 0.307

Analysis of Variance:

Group	DF	SS	MS	F	P
Regression	1	1.755	1.755	18.620	<0.001
Residual	104	9.805	0.0943		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	-0.0332		0.279		
NO_3-N	0.386	0.390	0.0894	18.620	<0.001

Variables not in Model

Group	F-to-Enter	P
NH_3-N	0.00181	0.966
PO4	0.312	0.578
TP	0.0512	0.821
DO	0.408	0.524
pH	0.453	0.502
COND	0.874	0.352
B_size	5.519	0.021
%O	5.404	0.022
%Bed	0.432	0.513
%Bo	1.526	0.220
%Cob	0.0545	0.816
%Gra	0.184	0.669
%S	0.122	0.727
%Si/Cl	0.0253	0.874
BRBDCOGR	0.0198	0.888
SDSTCL	0.0923	0.762
BRBD	1.834	0.179
CBGR	0.0577	0.811
R_wid	3.259	0.074
R_len	2.035	0.157
Flow	0.0139	0.906

Step 2: BASIN Entered

R = 0.442 Rsqr = 0.195 Adj Rsqr = 0.179

Standard Error of Estimate = 0.301

Analysis of Variance:

Group	DF	SS	MS	F	P
Regression	2	2.254	1.127	12.474	<0.001
Residual	103	9.307	0.0904		

Variables in Model

Group	Coef.	Std. Coeff.	Std. Error	F-to-Remove	P
Constant	-0.207		0.283		
NO_3-N	0.366	0.369	0.0879	17.295	<0.001
B_size	0.160	0.209	0.0682	5.519	0.021

Variables not in Model

Group	F-to-Enter	P
NH_3-N	0.203	0.653
PO4	2.024	0.158
TP	0.851	0.358
DO	1.315	0.254
pH	1.336	0.250
COND	1.045	0.309

%O	0.0430	0.836
%Bed	0.184	0.669
%Bo	1.290	0.259
%Co	0.240	0.626
%Gra	0.000224	0.988
%S	0.529	0.469
%Si/Cl	0.0176	0.895
BRBDCOGR	0.217	0.642
SDSTCL	0.541	0.464
BRBD	1.486	0.226
CBGR	0.00478	0.945
R_wid	0.0850	0.771
R_len	0.0201	0.887
Flow	0.299	0.585

Summary Table

Step #	**Vars. Entered**	**Vars. Removed**	**R**	**RSqr**	**Delta RSqr**	**Vars in Model**
1	NO_3-N		0.390	0.152	0.152	1
2	B_size		0.442	0.195	0.0431	2

The dependent variable AFDM can be predicted from a linear combination of the independent variables:

	P
NO_3-N	**<0.001**
B_size	**0.021**

The following variables did not significantly add to the ability of the equation to predict AFDM and were not included in the final equation:

NH_3-N PO4 TP DO pH COND %O %Bed %Bo %Cob %Gra %S %Si/Cl BRBDCOGR SDSTCL BRBD CBGR R_wid R_len Flow

Normality Test: Passed (P = 0.453)
Constant Variance Test: Failed (P = <0.001)
Power of performed test with alpha = 0.050: 0.998