

Violation and Elimination - Finetex, Inc., 418 Falmouth
Avenue, E. Paterson, N.J.
August 14, 1972 (J. Perrapato)

Mr. A. Fuenning of E. Paterson called the PVSC that Fleischer's Brook had a white substance in it. Mr. W. Fleming and Inspector J. Perrapato at 3:05 P.M. traced the white material to Finetex Inc. of E. Paterson.

The plant manager, Mr. T. Scaza, Jr., was made aware of the pollution at 3:30 P.M. A check in the plant revealed that a coil in one of their 550 gallon vats had a leak and a synthetic sperm oil was being carried to Fleischer's Brook through their floor drain. The coil was shut off and removed for repair. Follow-up-check revealed stream clear on Tuesday, August 15, 1972.

BAA000012

Appendix 8

Total
4 Pages



Kirk -
Your comments

Please -

BB

Mr. Neil Edsall
NJDEP Enforcement
327 E. Ridgewood Ave.
Paramus, New Jersey 07652

September 25, 2000

Dear Mr. Edsall

In response to your request we are pleased to provide you with our planned corrective action to hopefully eliminate reoccurrence of the incident experienced at our Elmwood Park facility September 14, 2000.

As best as we can determine we believe the incident was caused by operator error resulting in a build up of pressure in the respective unit. Under this circumstance and according to design, the pressure was released through an emergency release mechanism.

Planned corrective action includes re-enforcing our existing training and process controls. Additionally, we are investigating installation of automatic pressure sensor switches. In the event unacceptable pressure builds up, the pressure sensor switch will automatically shut down the unit. This new device will be in addition to the original design safety pressure release mechanism device.

A commitment to the automatic pressure switch will be made after engineering data is reviewed and we are confident the automatic system will work as intended. I will keep you informed.

FINETEX INC., P.O. BOX 216, ELMWOOD PARK, NEW JERSEY 07407 (201) 797-4686 FAX: (201) 797-6558

P.O. BOX 164, SPENCER, NORTH CAROLINA 28159 (704) 633-8028 FAX: (704) 633-3746

We have not had any such incident in decades and appreciate your very professional assistance in dealing with this incident.

Finetex looks forward to you and your staff's visit to our Elmwood Park plant. Danny Menchan, Plant Manager, will assist you in setting appropriate times for the tour.

Bob Scala,

A handwritten signature in black ink, appearing to read "Bob Scala", followed by a horizontal line.

Vice President
Finetex Inc.

FINETEX INC., P.O. BOX 216, ELMWOOD PARK, NEW JERSEY 07407 (201) 797-4686 FAX: (201)797-6558

P.O. BOX 164, SPENCER, NORTH CAROLINA 28159 (704) 633-8028 FAX: (704) 633-3746

BERGEN COUNTY PROSECUTOR'S OFFICE

REPORTING FORM

Date of Incident 09/14/2000

Time 0242 A.M. P.M.

1. Location (If business--name of company and type of business)

Finetex
418 Falmouth Ave. Elmwood Park

2. Type of call (spill, dump, etc.)

SPILL

3. Accidental ☒

4. Deliberate ☐

5. Undetermined ☐

6. Suspected substance involved Needle -> Final Product - Finsolv

7. Agencies Notified

Responded

Local Police Department ☒ Yes ☒ No ☐

Local Fire Department ☒ Yes ☒ No ☐

HAZMAT ☒ Yes ☒ No ☐

DEP ☒ Yes ☐ No ☐

BPA ☐ Yes ☐ No ☐

OSHA ☐ Yes ☐ No ☐

Local Health ☐ Yes ☐ No ☐

County Health ☐ Yes ☐ No ☐

Prosecutor's Office ☐ Yes ☐ No ☐

8. Other Agencies ☐ Yes ☐ No ☐

Name _____

Pt. Joseph Barone
Person Making Report

Elmwood Park Police 2000-10872
Agency & File #

B.C.P.O. USE ONLY

EP# 00-09-14-03-23-02 MISC# _____

REFERRAL# _____

Please make your check payable to:

H.M.H.T.T.C. Response Team, Inc.
PO Box 5215
Parsippany, NJ 07054
Tax ID#:22-3326-215

Voice: 973-335-6696
Fax: 973-335-7758

Invoice

Invoice Number:
R-5705

Invoice Date:
Sep 28, 2000

Page:
1

Sold To:
Finetex
PO Box 164
Spencer, NC 28159

Incident Date: Thurs., Sept. 14, 2000
Job Report: NJ 1285
Shift Rate: Shift1 & 2
Incident Ref: NJ-00-0190

Customer ID: Finetex07407

Customer PO	Payment Terms	Due Date	Sales Rep ID
	1% 10, Net 30 Days	10/28/00	

Quantity	Item	Description	Unit Price	Extension
		Location: Elmwood Park, NJ.		
		Respond to chemical reactor explosion with product on roof and ground. The roof was washed with hot water and soap. All waste was drummed for disposal.		
		Contact: Kirby Atwood		
		SHIFT 1		
5.50	EC	Emergency Response Haz-Mat Coordinator, (Supervisor)	60.00	330.00
5.50	ER	Emergency Response Technician	41.00	225.50
5.50	ER	Emergency Response Technician	41.00	225.50
5.50	ER	Emergency Response Technician	41.00	225.50
5.50	ER	Emergency Response Technician	41.00	225.50
		SHIFT 2		
1.50	ECOT	Emergency Response Haz-Mat Coordinator, (Supervisor) OT	90.00	135.00
1.50	EROT	Emergency Response Technician OT	61.50	92.25
1.50	ER	Emergency Response Technician	41.00	61.50
1.50	ER	Emergency Response Technician	41.00	61.50
7.00	100	Emergency Response, First Line	60.00	420.00

We will add finance charges on invoices more than 30 days old.

Transportation & Disposal invoice to follow - if applicable

Check No:
Port of Elizabeth

Subtotal	Continued
Sales Tax	Continued
Total Invoice Amount	Continued
Payment Received	
TOTAL	Continued

Please make your check payable to:

H.M.H.T.T.C. Response Team, Inc.
PO Box 5215
Parsippany, NJ 07054
Tax ID#:22-3326-215

Voice: 973-335-6696
 Fax: 973-335-7758

Invoice

Invoice Number:
 R-5705

Invoice Date:
 Sep 28, 2000

Page:
 2

Sold To:

Finetex
 PO Box 164
 Spencer, NC 28159

Incident Date: Thurs., Sept. 14, 2000
 Job Report: NJ 1285
 Shift Rate: Shift1 & 2
 Incident Ref: NJ-00-0190

Customer ID: Finetex07407

Customer PO	Payment Terms	Due Date	Sales Rep ID
	1% 10, Net 30 Days	10/28/00	

Quantity	Item	Description	Unit Price	Extension
7.00	500	Vehicle		
		Emergency Response Box Truck with Lift Gate	45.00	315.00
1.00	500A	Standard Box Truck with Lift Gate	200.00	200.00
1.00	E-09	Drum Vacuum System - Replacement	350.00	350.00
1.00	S-01	#100/#200 Sorbent Pads	83.00	83.00
15.00	S-09	Speedy Dry	8.50	127.50
7.00	S-10	Drum, 17H (Open Head) 55 Gallon	48.00	336.00
4.00	R-20	Chemical Resistant Overboots	14.00	56.00
		Friday, Sept. 15, 2000		
		Return to site to continue excavation of contaminated soil and to stock pile. Decon roof.		
7.00	EC	Emergency Response Haz-Mat Coordinator, (Supervisor)	60.00	420.00
7.00	GT	General Technician	38.00	266.00
7.00	GT	General Technician	38.00	266.00
1.00	300	Standard Pick-up Truck	100.00	100.00
1.00	500A	Standard Box Truck with Lift Gate	200.00	200.00
1.00	C-01	Mobile Phone Communications	30.00	30.00
1.00	2700	Mini-Excavator Crawler (Delivery Additional)	450.00	450.00
1.00	S-07	Polyethylene Film (6 Mil, 28 X 100)	105.00	105.00
		Monday, Sept. 18, 2000		
		Return and load roll off box and decon		

We will add finance charges on invoices more than 30 days old.

Transportation & Disposal invoice to follow - if applicable

Check No:

Port of Elizabeth

Subtotal
 Sales Tax
 Total Invoice Amount
 Payment Received
TOTAL

Continued
 Continued
 Continued
 Continued

Please make your check payable to:

H.M.H.T.T.C. Response Team, Inc.
PO Box 5215
Parsippany, NJ 07054
Tax ID#:22-3326-215

Voice: 973-335-6696
Fax: 973-335-7758

Invoice

Invoice Number:
R-5705

Invoice Date:
Sep 28, 2000

Page:
3

Sold To:
Finetex
PO Box 164
Spencer, NC 28159

Incident Date: Thurs., Sept. 14, 2000
Job Report: NJ 1285
Shift Rate: Shift1 & 2
Incident Ref: NJ-00-0190

Customer ID: Finetex07407

Customer PO	Payment Terms	Due Date	Sales Rep ID
	1% 10, Net 30 Days	10/28/00	

Quantity	Item	Description	Unit Price	Extension
		driveway.		
6.00	EC	Emergency Response Haz-Mat Coordinator, (Supervisor)	60.00	360.00
6.00	GT	General Technician	38.00	228.00
6.00	GT	General Technician	38.00	228.00
1.00	300	Standard Pick-up Truck	100.00	100.00
1.00	500A	Standard Box Truck with Lift Gate	200.00	200.00
1.00	2700	Mini-Excavator Crawler (Delivery Additional)	450.00	450.00
1.00	FS	* NOTE: Temporary Fuel Surcharge of 5% on Fuel Consuming Equipment	121.75	121.75

We will add finance charges on invoices more than 30 days old.

Transportation & Disposal invoice to follow - if applicable

Check No:

Port of Elizabeth

Subtotal	6,994.50
Sales Tax	411.57
Total Invoice Amount	7,406.07
Payment Received	
TOTAL	7,406.07



State of New Jersey

Christine Todd Whitman
Governor

Department of Environmental Protection
Northern Bureau of Water Compliance and Enforcement
1259 Route 46, Building 2
Parsippany, New Jersey 07054-4191
Telephone (973) 299-7592 Fax (973) 299-7719

Robert C. Shinn, Jr.
Commissioner

May 18, 1999

CERTIFIED MAIL
RETURN RECEIPT REQUESTED

Kirby Atwood, Compliance Coordinator
Finetex
PO Box 164
Spencer, North Carolina 28159

Re: Settlement Agreement
Finetex
NJPDES No. NJ0003573
Elmwood Park/Bergen County

Dear Mr. Atwood:

Enclosed is a Settlement Agreement ("Agreement") which was prepared by the New Jersey Department of Environmental Protection ("NJDEP" or "the Department") in order to resolve penalty liability accrued by Finetex for a permit effluent violation which occurred at your facility in Elmwood Park, New Jersey during October 1, 1998 through October 31, 1998 monitoring period.

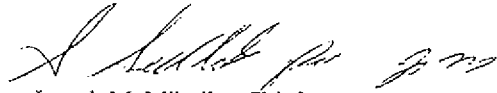
I have signed the attached copies. Both Settlement Agreement originals should be executed by a duly authorized representative of Finetex and one signed original returned to the Northern Bureau of Water Compliance and Enforcement within fourteen (14) calendar days of your receipt of this letter. Written certification that the person signing the Agreement is authorized to do so by Finetex must be submitted with the signed original. Your payment of \$1,000 in settlement of the violation shall be submitted within the time period specified in the Settlement Agreement along with the specified portion of the attached penalty invoice to the following address:

Division of Revenue
New Jersey Department of Treasury
P.O. Box 417
Trenton, New Jersey 08625-0417

Should Finetex not execute and return this Settlement Agreement within the time specified above, or make payment within the time specified in the Settlement Agreement, this offer shall be null and void. If

you have any questions regarding this Agreement, please contact Rebecca J. Manis of my staff at (973) 299-7592 or by letter through this Bureau.

Very truly yours,



Joseph M. Mikulka, Chief
Northern Bureau of Water
Compliance and Enforcement

E16
Enclosure

c: Administrator James K. Hamilton, Water Compliance and
Enforcement Element
Director Dennis Hart, Division of Water Quality
Bureau of Revenue



State of New Jersey

Christine Todd Whitman
Governor

Department of Environmental Protection
Northern Bureau of Water Compliance and Enforcement
1259 Route 46, Building 2
Parsippany, New Jersey 07054-4191
Telephone (973) 299-7592 Fax (973) 299-7719

Robert C. Shinn, Jr.
Commissioner

IN THE MATTER OF : SETTLEMENT AGREEMENT
FINETEX :

The following Settlement Agreement is entered into pursuant to the authority vested in the Commissioner of the New Jersey Department of Environmental Protection (hereinafter "NJDEP" or "Department") by N.J.S.A. 13:1D-1 et seq. and duly delegated to the Bureau Chief of the Northern Bureau of Water Compliance and Enforcement.

WHEREAS Finetex located at 418 Falmouth Avenue in Elmwood Park, Bergen County, New Jersey, Lot 803, Block 15, possesses a New Jersey Pollutant Discharge Elimination System (NJPDES) Permit No. NJ0003573; and,

WHEREAS Finetex has submitted monthly Discharge Monitoring Reports ("DMR's") to the Department in accordance with the NJPDES Permit; and,

WHEREAS a review of the DMR, shows that Finetex discharged pollutants to the waters of this State in excess of the discharge limitations contained in Part III - B/C of the permit, for the period of October 1, 1998 through October 31, 1998 as indicated below:

MONIT				MIN				
PERIOD		DSN		CONC	AVG			
END DATE	VIOL	NO.	PARAMETER	LOAD	MAX	LIMITS	UNITS	DATA
10/31/98	E94	001	PHC	CONC	AVG	10	mg/L	12.67

The following abbreviations were used above:

E94 - Monthly Average, Concentration



State of New Jersey

Christine Todd Whitman
Governor

Department of Environmental Protection
Northern Bureau of Water Compliance and Enforcement
1259 Route 46, Building 2
Parsippany, New Jersey 07054-4191
Telephone (973) 299-7592 Fax (973) 299-7719

Robert C. Shinn, Jr.
Commissioner

May 18, 1999

CERTIFIED MAIL
RETURN RECEIPT REQUESTED

Kirby Atwood, Compliance Coordinator
Finetex
PO Box 164
Spencer, North Carolina 28159

Re: Settlement Agreement
Finetex
NJDES No. NJ0003573
Elmwood Park/Bergen County

Dear Mr. Atwood:

Enclosed is a Settlement Agreement ("Agreement") which was prepared by the New Jersey Department of Environmental Protection ("NJDEP" or "the Department") in order to resolve penalty liability accrued by Finetex for a permit effluent violation which occurred at your facility in Elmwood Park, New Jersey during October 1, 1998 through October 31, 1998 monitoring period.

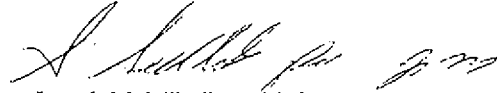
I have signed the attached copies. Both Settlement Agreement originals should be executed by a duly authorized representative of Finetex and one signed original returned to the Northern Bureau of Water Compliance and Enforcement within fourteen (14) calendar days of your receipt of this letter. Written certification that the person signing the Agreement is authorized to do so by Finetex must be submitted with the signed original. Your payment of \$1,000 in settlement of the violation shall be submitted within the time period specified in the Settlement Agreement along with the specified portion of the attached penalty invoice to the following address:

Division of Revenue
New Jersey Department of Treasury
P.O. Box 417
Trenton, New Jersey 08625-0417

Should Finetex not execute and return this Settlement Agreement within the time specified above, or make payment within the time specified in the Settlement Agreement, this offer shall be null and void. If

you have any questions regarding this Agreement, please contact Rebecca J. Manis of my staff at (973) 299-7592 or by letter through this Bureau.

Very truly yours,



Joseph M. Mikulka, Chief
Northern Bureau of Water
Compliance and Enforcement

E16
Enclosure

c: Administrator James K. Hamilton, Water Compliance and
Enforcement Element
Director Dennis Hart, Division of Water Quality
Bureau of Revenue



State of New Jersey

Christine Todd Whitman
Governor

Department of Environmental Protection
Northern Bureau of Water Compliance and Enforcement
1259 Route 46, Building 2
Parsippany, New Jersey 07054-4191
Telephone (973) 299-7592 Fax (973) 299-7719

Robert C. Shinn, Jr.
Commissioner

IN THE MATTER OF : SETTLEMENT AGREEMENT
FINETEX :

The following Settlement Agreement is entered into pursuant to the authority vested in the Commissioner of the New Jersey Department of Environmental Protection (hereinafter "NJDEP" or "Department") by N.J.S.A. 13:1D-1 et seq. and duly delegated to the Bureau Chief of the Northern Bureau of Water Compliance and Enforcement.

WHEREAS Finetex located at 418 Falmouth Avenue in Elmwood Park, Bergen County, New Jersey, Lot 803, Block 15, possesses a New Jersey Pollutant Discharge Elimination System (NJPDES) Permit No. NJ0003573; and,

WHEREAS Finetex has submitted monthly Discharge Monitoring Reports ("DMR's") to the Department in accordance with the NJPDES Permit; and,

WHEREAS a review of the DMR, shows that Finetex discharged pollutants to the waters of this State in excess of the discharge limitations contained in Part III - B/C of the permit, for the period of October 1, 1998 through October 31, 1998 as indicated below:

MONIT PERIOD END DATE	VIOL	DSN NO.	PARAMETER	CONC LOAD	MIN AVG MAX	LIMITS	UNITS	DATA
10/31/98	E94	001	PHC	CONC	AVG	10	mg/L	12.67

The following abbreviations were used above:

E94 - Monthly Average, Concentration

FINETEX INC.

418 Falmouth Ave
Elmwood Park, NJ

Appendix 12

Environmental Sampling Reports

BAA0000010

April 12, 2003



BRENNAN ENVIRONMENTAL, INC.
Environmental Consulting Services

T. 973.781.1801
F. 973.781.1804
www.bei-env.com

8 Great Meadow Lane / Unit D
East Hanover, NJ 07936-1703

Sent Via 1st Class Mail

January 13, 2003

Ms. Jeannette Cleary
NJDEP, DRPSR, Bureau of Field Operations
Initial Notice Section
401 E State Street
CN 435
Trenton, NJ 08625-0435

Re: Finetex, Inc.
410, 416 & 418 Falmouth Ave
Elmwood Park, Bergen County, NJ
BEI Job # 99704
ISRA Case # E20010242

Dear Ms. Cleary:

As per Brennan Environmental Inc.'s January 3, 2003 letter, please find attached a copy of the Site Investigation Report (SIR) prepared on behalf of Finetex, Inc. In the SIR, additional soil and ground water delineation has been recommended for the subject property.

If you have any questions concerning this matter, please call me at (973) 781-1801.

Sincerely,



John M. Brennan, President
Brennan Environmental, Inc.

Cc: Roger Porter
Bob Scala
Lawrence Jacobs, Esq.

G:\Brennan Env\Tcjobs\2001\99704, Finetex Inc\njdepnoticeltr.doc

TIERRA-B-010177



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

REPORT CERTIFICATION PAGE





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

Site Investigation Report

PER N.J.A.C. 7:26E-4.8

Finetex, Inc.
418 (includes 410 & 416) Falmouth Avenue
Block 803, Lots 14, 15
Elmwood Park, Bergen County, New Jersey
ISRA Case #E2001242
January 13, 2003
BEI Job #99704

Prepared for:

Finetex Inc.
418 Falmouth Avenue
Elmwood Park, NJ 07407

Prepared by:

Brennan Environmental Inc.
8D Great Meadow Lane
East Hanover, New Jersey 07936

TABLE OF CONTENTS

1.0) INTRODUCTION	1
2.0) HISTORICAL INFORMATION	1
3.0) PHYSICAL SETTING	2
3.1) SITE GEOLOGY	2
3.2) SITE HYDROGEOLOGY	2
3.3) SITE TOPOGRAPHY & SURROUNDING LAND USE	2
3.4) 1,000 FOOT RADIUS LAND USE	3
3.5) SURFACE WATER BODY ASSESSMENT	3
3.6) BASELINE ECOLOGICAL ASSESSMENT AND RECEPTOR ASSESSMENT	3
4.0) TECHNICAL OVERVIEW	4
4.1) RELIABILITY OF ANALYTICAL DATA	4
4.2) SITE CONTAMINATION SUMMARY	5
4.3) SIGNIFICANT EVENTS AND/OR SEASONAL VARIATIONS INFLUENCING THE INVESTIGATION	6
5.0) FINDINGS/RECOMMENDATIONS	6
5.1) SPECIFIC AREAS OF CONCERN DISCUSSION	6
5.1.A) <i>Impacted Soil</i>	6
5.1.B) <i>Ground Water Monitoring Well Installation and Sampling</i>	8
5.2) SIGNIFICANCE OF TENTATIVELY IDENTIFIED COMPOUNDS	11
6.0) RECOMMENDATIONS FOR AREAS OF CONCERN	11



TABLE OF CONTENTS

TABLES

Table 1– Summary Of May 23, 2002 Soil Analytical Results.....	7
Table 2– Summary Of November 2, 2002 Soil Analytical Results	8
Table 3– Summary Of November 18, 2002 Ground Water Analytical Results.....	10

FIGURES

SITE LOCATION MAP	1
U.S.G.S. 7.5 MINUTE TOPOGRAPHIC QUADRANGLE MAP.....	2
COPY OF NJDEP WETLAND INVENTORY MAP	3
SITE MAP	4
SAMPLE LOCATION MAP	5

APPENDICES

COPY OF NJDEP MARCH 7, 2002 CORRESPONDENCE.....	1
SOIL BORING LOGS	2
MONITORING WELL INSTALLATION INFORMATION.....	3
PURGE GUIDES	4
LABORATORY ANALYTICAL SOIL DATA REPORT (E02-3831)	5
LABORATORY ANALYTICAL SOIL DATA REPORT (E02-8630)	6
LABORATORY ANALYTICAL GROUND WATER DATA REPORT (E02-9109).....	7



1.0) INTRODUCTION

Brennan Environmental, Inc. (BEI) was retained by Finetex, Inc. (Finetex) to prepare this Site Investigation Report (SIR) for the facility located at 418 Falmouth Avenue, Block 803 Lots 14 and 15, Elmwood Park, Bergen County, New Jersey (subject property). The site location is indicated on Figure #1. The purpose of this report is to detail the actions implemented in response to NJDEP's March 7, 2002 letter.

According to NJDEP's March 7, 2002 letter, based on the information submitted in the Preliminary Assessment Report (PAR) dated November 15, 2001 and UST Closure/SIR dated November 8, 2001, one historic area of environmental concern required further investigation. BEI was required to address the location of the six (6) former USTs (AOC #4). These tanks were removed in 1988. The March 7, 2002 letter directed Finetex to collect one soil sample through the former UST excavation and have it analyzed for volatile organic compounds plus a forward library search (VO+10), base/neutral compounds plus a forward library search (BN+15) and total petroleum hydrocarbons (TPHC).

BEI is certified by the New Jersey Department of Environmental Protection (NJDEP) to perform UST closures and subsurface evaluations pursuant to N.J.S.A. 58:10A-24 (Certification #US00779).

This report was prepared in accordance with NJDEP's Technical Requirements for Site Remediation (TRSR) (N.J.A.C. 7:26E et. seq) and details additional soil and ground water investigation activities conducted at the property located at 418 Falmouth Avenue, Block 803 Lots 14 and 15, Elmwood Park, Bergen County, New Jersey.

2.0) HISTORICAL INFORMATION

A Preliminary Assessment Report was conducted for the Finetex property, pursuant to ISRA Case No. E2001242, dated November 15, 2001. The PAR identified two areas of environmental concern that warranted additional investigation. Specifically, an inactive 550-gallon #2 heating oil UST and an area of a former 1,000-gallon #2 heating oil UST were targeted (AOC 5 & 6).

BEI prepared a UST Closure/Site Investigation Report (SIR) to detail the tank closure process and post-excavation soil sampling activities, soil boring investigation, and BEI's evaluation of the post-excavation soil sampling data, which was compared to the then current applicable NJDEP soil cleanup criteria. The total aggregate volume of the two former USTs was less than 2,000 gallons. Therefore, the USTs were not subject to the underground Storage of Hazardous Substances Act (N.J.S.A. 58:10A-21), and a closure plan was not required. BEI proposed no further action (NFA) for the subject property. NJDEP approved no further action for the AOC 5 & 6 but required additional investigation of AOC 4 as detailed above.



3.0) PHYSICAL SETTING

3.1) Site Geology

The subject property is located in the Piedmont Physiographic Province of New Jersey. According to the USGS Bedrock Geologic Map of Northern New Jersey dated 1996, the bedrock in this section of the Piedmont Province of New Jersey is described as the Passaic Formation, which is Lower Jurassic and Upper Triassic in age and composed predominantly of conglomeratic sandstone with some shale. This sandstone is described as interbedded grayish-red to brownish-red, medium to fine grained and medium to thick bedded.

According to the Rutgers University Engineering Soil Survey of New Jersey (Report No. 4, Bergen and Hudson Counties) the overburden materials at the site are mapped as "GS12ig". The "GS" portion of the symbol signifies that the area is underlain by glacial stratified drift, composed of non-residual materials deposited during the Wisconsin glaciation. The general characteristic of the moraine is assorted, homogeneous materials, consisting predominantly of sand sizes, with varying amounts of silt, gravel, and occasionally clay. The "ig" indicates imperfect to good internal drainage conditions. Depth to the top of bedrock is expected to be greater than 10 feet. Bedrock was not encountered during soil or groundwater investigation activities.

The overburden soils encountered during monitoring well installation activities consisted predominantly of sands used to backfill the excavation. However, soils encountered in the former UST excavation area correlated well with the above description and were as follows:

- 0-3' Light brown SILT, little Clay, some coarse Gravel and organic material
- 3-8.5' Dark brown CLAY, little coarse Gravel
- 8.5-15.0' Gray brown CLAY, little Silt, trace fine Sand

3.2) Site Hydrogeology

According to the Rutgers University Engineering Soil Survey of New Jersey, the drainage conditions for areas mapped as "GS-12ig" are described as imperfect to good. During well installation activities, ground water stabilized at 6.24 feet bsg. According to site topography, groundwater is expected to flow in a southern direction. The recharge rate of the monitoring well is fair to poor.

3.3) Site Topography & Surrounding Land Use

The facility location is shown on an excerpt from the U.S.G.S. 7.5' Hackensack, Topographic Quadrangle Map (see Figure #2). According to the topographic quadrangle map, the site is located approximately 49 feet above mean sea level.



3.4) 1,000 Foot Radius Land Use

Based on visits to the site and the topographic map, land use within a 1,000 foot radius of the site appears to be a mixture of commercial, residential and heavy industrial.

The local land usage is zoned as heavy industrial and residential. The property is bounded by heavy industrial on three sides, city streets, a railroad line, and vacant land followed by a cemetery.

3.5) Surface Water Body Assessment

The closest surface body of water to the site is the Fleischer Brook, which is located approximately 755 feet west of the subject site. Pursuant to the New Jersey Surface Water Quality Standards (N.J.A.C. 7:9-4 et. seq.) the Fleischer Brook has not been assigned a designation. The brook is not a tributary to the Passaic River. Based on the distance of the brook from the subject property, the brook is not believed to be a receptor of concern.

3.6) Baseline Ecological Assessment and Receptor Assessment

Wellhead Assessment: According to our review of NJ Geological Survey's computer program PSW 2000, there are six public potable wells within a 2,000-foot radius of the subject property. The wells are owned by the Garfield Water Department. A copy of the printout from the PSW 2000 software can be found in Appendix 7 of the November 8, 2001 UST Closure/SI Report.

Surface Water Body Assessment: The closest body of water to the site is the Fleischer Brook, located approximately 755 feet west of the subject property. Fleischer Brook appears to run southeast from the subject property. Fleischer Brook is not classified in NJDEP's surface water quality standards, nor is it a formal tributary of the Passaic River. Due to the distance between the subject property and Fleischer Brook, the Fleischer Brook is not considered a receptor of concern.

Wetland Assessment: Review of the 1991 New Jersey Wetlands Inventory Map for the Hackensack southwest quarter quadrangle, which is compiled by the NJDEP, indicates no wetland / state open water areas on or adjacent to the subject property. The subject property and surrounding area are designated as "Upland." A copy of the Wetland Inventory Map is included as Figure #3 of this report.

Baseline Ecological Assessment: In line with N.J.A.C. 7:26E-3.11, BEI performed a baseline ecological assessment for the subject property. The results are as follows:

Contaminants of Potential Ecological Concern: Analytical results of the soil samples obtained from the subject tank area at the property, indicate benzene and chlorobenzene



detected above NJDEP's Impact to Groundwater Criteria. Ground water analytical results indicate several other compounds above NJDEP Ground Water Quality Standards.

Environmentally Sensitive Areas (ESA): After a site inspection and review of the NJDEP wetlands map and a surface water body assessment, no ESAs were found on or adjacent to the subject property. Therefore, ESAs are not considered to be a concern.

Potential Migration Pathways: Ground water was encountered during monitoring well installation activities at a depth of 6.25 feet bsg and is a potential migration pathway. There were no buried utilities in the areas of the former USTs that could have acted as a migration pathway.

Since the baseline ecological assessment indicates that the three criteria above do not co-occur at the subject property, no further ecological investigation is warranted.

4.0) TECHNICAL OVERVIEW

4.1) Reliability of Analytical Data

BEI's sampling activities were performed in accordance with the requirements outlined in the *NJDEP Field Sampling Procedures Manual* dated May 1992, as well as the *Alternative Ground Water Sampling Techniques Guide* dated July 1994. The soils encountered during investigative activities were classified in the field according to the Bermeister Soil Classification System (BSCS). The samples were placed in precleaned amber glass bottles with Teflon lined caps and placed in a cooler maintained at 4°C in order to ensure proper preservation. Proper chain of custody documentation was maintained until delivery of the samples to the laboratory.

All samples were analyzed by Integrated Analytical Laboratories, Inc. (IAL) of Randolph, New Jersey, a New Jersey certified laboratory (NJDEP Certification #14751). The data quality assurance deliverables conform to the Non-USEPA CLP Method as specified in the N.J.A.C. 7:26E - Technical Requirements for Site Remediation - Appendix A - Section IV. Please note that pursuant to NJDEP's letter dated March 7, 2002, the initial soil boring sample collected on May 23, 2002 was analyzed for VO+10, BN+15, and TPHC. The only contaminant found above standard during the May 23, 2002 sampling event was chlorobenzene. Subsequent soil samples were analyzed for VO+10 only.

All soil samples discussed herein were compared to the NJDEP's Soil Cleanup Criteria last revised on May 12, 1999. The Residential Direct Impact Soil Cleanup Criteria was used throughout the soil column. However, in cases where the Impact to Ground Water Criteria was lower, the strictest criteria were used throughout the soil column. The minimum detection limits (MDLs) for the May 23, 2002 sampling event did not exceed the applicable soil cleanup criteria. However, the MDL for chlorobenzene in MW-1A for the November 2, 2002 soil sampling event



exceeded applicable NJDEP Soil Cleanup Criteria. However, since the analytical results for chlorobenzene is over applicable Soil Cleanup Criteria, the elevated MDL is not of concern.

All ground water samples collected were analyzed for VO+10 and BN+15. All ground water sample results discussed herein, were compared to the current NJDEP Class IIa Ground Water Quality Standards (GWQS). The MDLs for vinyl chloride and benzene for the November 18, 2002 ground water sampling event exceeded applicable NJDEP GWQS. Since the analytical result for benzene is over the applicable GWQS, the elevated MDL is not of concern. The presence of vinyl chloride in the ground water samples is discussed further in section 5.1B.

A summary of analytical methods and quality assurance is included in the Analysis Conformance/Nonconformance Summary pages in the laboratory packages. All locations of the soil samples and the monitoring well are shown in the Figures section of this report.

The following "Key" provides the explanations necessary to understand the sampling result summary tables listed in the following report sections.

ABBREVIATION KEY

TVOCC - Indicates Total Volatile Organic Contaminant Concentration.
TOCC - Indicates Total Organic Contaminant Concentration.
MDL - Indicates Method Detection Limit / PQL - Indicates Practical Qualitative Limits.
C - Indicates common laboratory and/or bottle contaminant.
NA - Indicates compound was Not Analyzed for in the analysis.
ND - Indicates compound was analyzed for but not detected at the MDL.
B - Indicates compound was detected in the blank and the sample.
~ - Indicates no listed value.
J - Indicates compound was detected at a value below MDL and greater than 0.
<u>Bold & Underlined</u> - Indicate that compound was detected above NJDEP standard.
ppm - Indicates parts per million.
ppb - Indicates parts per billion.
All other compounds analyzed in the particular scan were not detected, therefore they are not listed in the summary table.

4.2) Site Contamination Summary

The information outlined in Section 5.1 summarizes the area of environmental concern (AOC) at the site that was investigated to comply with the TRSR, N.J.A.C. 7:26E. Based on NJDEP's March 7, 2002 letter, BEI installed one soil boring in AOC 4. As per NJDEP's letter, the soil sample was analyzed for VO+10, BN+15, and TPHC. Analytical results revealed chlorobenzene concentrations slightly above NJDEP's Impact to Groundwater Standard of 1 ppm. Therefore Finetex installed one (1) monitoring well within the former UST excavation to determine the potential impact on ground water quality. During well installation activities, soil samples were taken to vertically delineate the impacted soil.



4.3) Significant Events and/or Seasonal Variations Influencing the Investigation

There were no seasonal variations observed by BEI during the course of the site investigation that influenced the investigation, sampling procedures, or analytical results.

5.0) FINDINGS/RECOMMENDATIONS

5.1) Specific Areas of Concern Discussion

5.1.A) Impacted Soil

In order to satisfy the request in NJDEP's March 7, 2002 letter, BEI was onsite on May 23, 2002 to install one soil boring in AOC#4. BEI installed the boring via hand-held Portaprobe direct push technology. The Portaprobe is an electric pneumatic hammer that drives a narrow diameter 3.0-foot long macrocore. The macrocore is similar to the Geoprobe macrocore with dedicated acetate liners. New liners were used for each 3.0 feet of recovery.

Samples for laboratory analysis were field screened with a PID at 1-foot intervals. The PID used to field screen the borings was calibrated with isobutylene span gas and equipped with a 10.6 eV UV lamp. Soil boring logs are contained in Appendix #2.

The soil boring was advanced, taking three-foot macrocore samples, to a depth of 6 feet bsg. A soil sample was collected designated SB-01, that corresponds to a level 0-6" below the base of former USTs.

Sample SB-01 was stored in a cooler and transported to IAL for analysis. Pursuant to NJDEP's letter dated March 7, 2002, the soil sample was analyzed for VO+10, BN+15, and TPHC. All soil samples were preserved using the NJDEP methanol preservation method. A trip blank was generated as a measure of quality assurance/quality control.

The laboratory analytical results of SB-01 indicated the presence of chlorobenzene in concentrations slightly above NJDEP's Impact to Groundwater Standard. A summary of the soil sample results is provided in Table #1 and the complete laboratory analytical package is provided in Appendix #5.



Table 1-- Summary Of May 23, 2002 Soil Analytical Results

Sample ID:	SB-01	RDCC/ NRDCC/ IGWCC
Lab ID:	E02-3831	
Sample Date:	5/25/2002	
Sample Matrix:	Soil	
Sample Depth:	6.0-6.5'	
Parameter		
<i>Volatiles (ppb)</i>		
Benzene	0.385 J	3/13/1
Chlorobenzene	7.95	37/680/1
Ethylbenzene	1.08	1000/1000/100
Total Xylenes	5.42	410/1000/67
1,3-Dichlorobenzene	4.66	5100/10000/100
1,4-Dichlorobenzene	28.80	570/10000/100
1,2-Dichlorobenzene	8.01	5100/10000/50
Total Targeted VO's	49.096 J	
Total VO TIC's	20.24	
Total VO's	69.336 J	1000
<i>Semivolatiles (ppb)</i>		
1,3-Dichlorobenzene	0.603	5100/10000/100
1,4-Dichlorobenzene	4.48	570/10000/100
1,2-Dichlorobenzene	0.123	5100/10000/50
Naphthalene	3.27	230/4200/100
2-Methylnaphthalene	2.14	~
Phenanthrene	1.87 J	~
Fluoranthene	0.081 J	2300/10000/100
Bis(2-Ethylhexyl)phthalate	0.092 J	49/210/100
Total Targeted BN's	12.730 J	
Total BN TIC's	128.81	
Total BN's	141.54	
TPHC	892	10000

J= concentration was detected at a value below the MDL

Since analytical results revealed concentrations of chlorobenzene slightly above NJDEP's Impact to Groundwater Criteria, BEI, on behalf of Finetex, installed one (1) monitoring well on the subject property to evaluate ground water.

During the November 2, 2002 monitoring well installation activities, BEI collected 3 soil samples designated MW-1A, MW-1B and MW-1C. A Geoprobe 66DT with direct push technology was used to collect samples and install the well. Samples were collected with a five-foot stainless-steel macrocore with dedicated acetate liners. New liners were used for each 5.0 feet of recovery.

The boring was field-screened using the same methodology as described earlier in this section. BEI collected samples from 0-6" above ground water and at the soil horizon levels exhibiting the



highest recorded PID readings from both the May 23, 2002 and November 2, 2002 site activities. All soil samples were preserved using the NJDEP methanol preservation method.

The location of the samples and corresponding PID results can be found on the well construction logs included as Appendix #3. Analytical results for the samples are provided in Table #2 and the complete laboratory analytical package is provided in Appendix #6.

Table 2- Summary Of November 2, 2002 Soil Analytical Results

Sample ID:	MW-1A	MW-1B	MW-1C	RDCC/ NRDCC/ IGWCC
Lab ID:	8630-001	8630-002	8630-003	
Sample Date:	11/02/02	11/02/02	11/02/02	
Sample Matrix:	Soil	Soil	Soil	
Sample Depth:	3.0-3.5'	4.5-5.0'	5.5-6.0'	
Parameter				
<i>Volatiles (ppb)</i>				
Benzene	1.56	0.649 J	0.29 J	3/13/1
Chlorobenzene	51.2 D	40.5	3.82	37/680/1
Ethylbenzene	3.4	4.3	0.386 J	1000/1000/100
Total Xylenes	13	18.4	1.97	410/1000/67
1,3-Dichlorobenzene*	20.8	31.4	1.64	5100/10000/100
1,4-Dichlorobenzene*	43.2	23.5 D	10.9	570/10000/100
1,2-Dichlorobenzene*	3.77	6.35	0.347 J	5100/10000/50
Total Targeted VO's	136.93	125.099 J	19.353 J	
Total VO TIC's	12.4	142.67	14.02	
Total VO's	149.33	267.769 J	33.373 J	1000

D= the compound was reported from the diluted analysis

J= concentration was detected at a value below the MDL

*= analyte was measured in both BN and VO results; as per IAL, B/N analysis is a more accurate measure of these compounds.

5.1.B) Ground Water Monitoring Well Installation and Sampling

BEI, at the request of Finetex, was on-site on November 2, 2002, with a drill crew from S&S Subsurface Investigation, Inc. to install one (1) ground water monitoring well on site. A six-inch diameter auger attachment was used to install the well in the area of the former USTs. The well, (designated MW-1), was installed as an unconsolidated well.

The well was constructed according to NJDEP Standard Unconsolidated Well Specifications using two-inch diameter, Schedule 40 PVC. Machine slot PVC well screen (.010 inch slot size) was placed from 3-15 ft bsg; moist soils were encountered at approximately 4.5' and groundwater stabilized at approximately 6.0' bsg. A solid schedule 40, PVC riser completed the upper part of the well. The connection between screen and riser were flush-joint screw type (no adhesive required). The wells were filter packed with clean (<10% fines), #2 uniformity coefficient sand from the bottom of the well to two feet above the top of the screen level.



Following this, the well was grouted using a mixture of Portland Type 2 cement. The well installed at the site was of the "flushmount" lockable type with a locking cap inside. Specific stratigraphy and well construction logs were recorded in Well Certification Forms "A" and "B" for each of the wells (Appendix #3). The well location for the site is depicted in the Figures section of the report.

After a two week stabilization period, a representative from BEI was on site on November 18, 2002 to collect a ground water sample from MW-1. The ground water sample was collected in accordance with the NJDEP's May 1992 FSPM. The sample collected from the monitoring well was packed in a cooler and transported to IAL for analysis for VO+10 and BN+15. Analytical parameters for ground water were chosen based upon the soil parameters required by NJDEPs March 7, 2002 letter.

Static water level measurements were recorded with a Sinco Slope Indicator by BEI upon arrival at the site, following well purging, and after sample collection. The monitoring well purge guide is included as Appendix #4. Prior to, and following monitoring well purging, a ground water sample was obtained with a dedicated clear bailer to check for free-phase floating product and product sheen. BEI noted neither sheen nor free-phase floating product on the samples collected from MW-1 for both sampling events.

Additionally, dissolved oxygen, pH, water temperature, and specific conductivity were recorded prior to and following well purging and after sample collection. Trip blanks were generated as a measure of quality control/quality assurance for each sampling event.

The monitoring well sample results are found in the following table and the full analytical data package is contained as Appendix #7 of this report.



Table 3-- Summary Of November 18, 2002 Ground Water Analytical Results

Sample ID:	MW-1	NJDEP GWQS
Lab ID:	9109-001	
Sample Date:	11/18/2002	
Sample Matrix:	Aqueous	
Parameter		
<i>Volatiles (ppb)</i>		
Vinyl Chloride	46.8	5
Trans-1,2-Dichloroethene	37.8	100
Benzene	1030	1
Toluene	76.9	1000
Chlorobenzene	2320 D	50
Ethylbenzene	498	700
Total Xylenes	2320	1000
1,3-Dichlorobenzene	1130	600
1,4-Dichlorobenzene	9370 D	75
1,2-Dichlorobenzene	428	600
Total VO's	17257.5	~
Total TIC's	1364	100/500
Total VO's & TIC's	18621.5	100/500
<i>Semivolatiles (ppb)</i>		
1,3-Dichlorobenzene	190	600
1,4-Dichlorobenzene	1650	75
1,2-Dichlorobenzene	100	600
1,2,4-Trichlorobenzene	46.8	9
Naphthalene	136	300
2-Methylnaphthalene	12.8	ns
Total BN's	2135.6	~
Total TIC's	380	100/500
Total BN's & TIC's	2515.6	100/500

D= the compound was reported from the diluted analysis

ns= no standard

J= concentration was detected at a value below the MDL

*= analyte was measured in both BN and VO results; as per IAL, B/N analysis is a more accurate measure of these compounds.

Ground Water sample analytical results from the November 18, 2002 sampling event detected several compounds above NJDEP Ground Water Quality Standards. Most of the compounds are similar to those found in the soil investigation. However, vinyl chloride and cis-1,2-dichloroethene (in TICs) were found as well. These compounds are not related to contaminant compounds found in the soils, nor are they associated with the contents of any of the former USTs located at the subject property.



5.2) Significance of Tentatively Identified Compounds

Total VO and BN TICs for all soil samples were below the NJDEP Soil Cleanup Criteria of 1000 ppm. Total VO and BN TICs noted in the ground water analysis for MW-1 exceeded the NJDEP Class IIA Ground Water Quality Standards criteria of 100/500 ppb. Therefore, TICs in ground water are of concern.

6.0) RECOMMENDATIONS FOR AREAS OF CONCERN

Based on the analytical results, soil and ground water have been impacted with chlorobenzene and its derivatives at the subject site. Further soil and ground water delineation activities should be limited to these compounds (e.g. chlorobenzene, dichlorobenzene, benzene et al.).

BEI proposes delineation of the extent of impacted soils and ground water of chlorobenzene and like compounds. A proposed schedule for future delineation activities will follow this report.





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

FIGURES



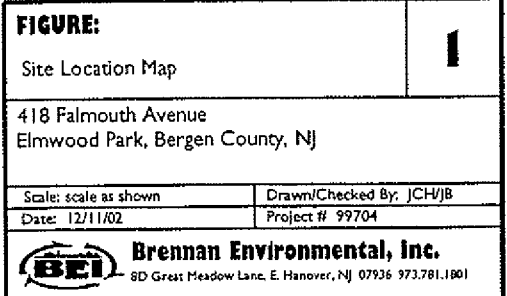




FIGURE:
USGS Topo Quad Map
(based on Hackensack, NJ Topo Quad)

2

418 Falmouth Avenue
Elmwood Park, Bergen County, NJ

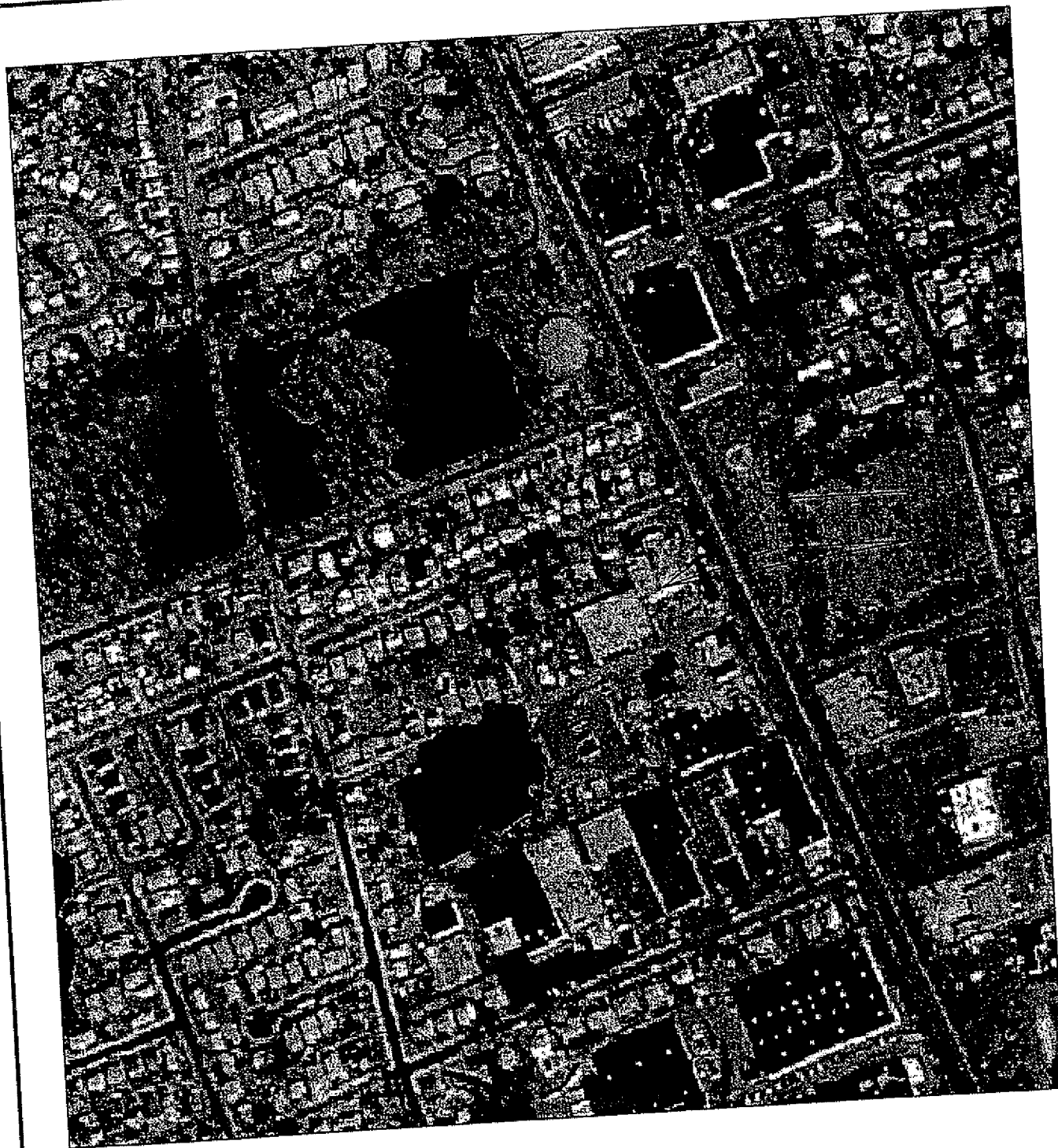
Scale: scale as shown
Date: 12/11/02

Drawn/Checked By: JCH/JB
Project # 99704



Brennan Environmental, Inc.

8D Great Meadow Lane, E. Hanover, NJ 07936 973.781.1801



BERWET (LABEL)
AGRICULTURAL WETLANDS (MODIFIED)
CONIFEROUS WOODED WETLANDS
DECIDUOUS SCRUB/SHRUB WETLANDS
DECIDUOUS WOODED WETLANDS
DISTURBED WETLANDS (MODIFIED)
FRESHWATER/TIDAL MARSHES
HERBACEOUS WETLANDS
MANAGED WETLANDS (MODIFIED)
MIXED FORESTED WETLANDS (CONIFEROUS DOM)
MIXED FORESTED WETLANDS (DECIDUOUS DOM)
MIXED SCRUB/SHRUB WETLANDS (DECIDUOUS DOM)
SALINE MARSHES
WETLAND RIGHTS-OF-WAY (MODIFIED)
BERSTRA
BERLAKES

SOUTHWEST QUARTERQUAD
OF THE HACKENSACK
TOPOGRAPHIC MAP

FIGURE:

NJDEP GIS Information Map
(source: NJDEP GIS Website)

3

418 Falmouth Avenue
Elmwood Park, Bergen County, NJ

Scale: scale as shown

Date: 12/11/02

Drawn/Checked By: JCH/JB

Project # 99704



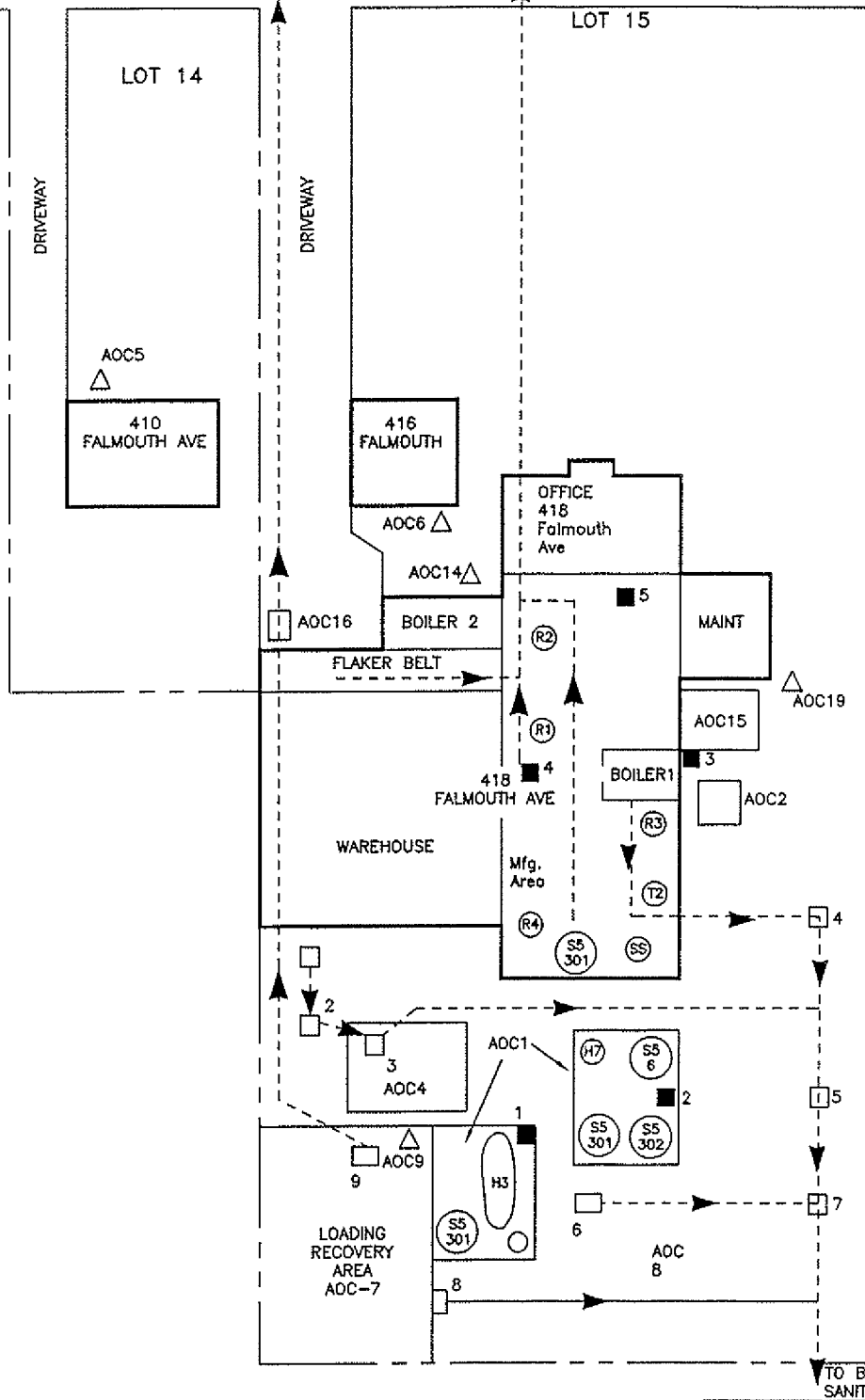
Brennan Environmental, Inc.

80 Great Meadow Lane, E. Hanover, NJ 07936 973.781.1801

TO STORM SEWER AND THEN TO
FLIESER'S BROOK

TO BOROUGH OF ELMWOOD PARK
SANITARY SEWER

FALMOUTH AVENUE



LELIARTS LANE

LEGEND:

- - SUMP
- - ABOVE GROUND STORAGE TANK
- - FLOOR DRAIN / CATCH BASIN
- △ - AOC

FIGURE #

4

SITE PLAN

FINETEX, INC.
418 FALMOUTH AVENUE
ELMWOOD PARK, NEW JERSEY

Scale: 1"=40'

Drawn By: AP

Date: 11/12/2001

Project # 99704



BRENNAN ENVIRONMENTAL, INC.
8 Great Meadow Lane E. Hanover, NJ 07936
Tel: (973)781-1801

*GW results in ppb, all others in ppm

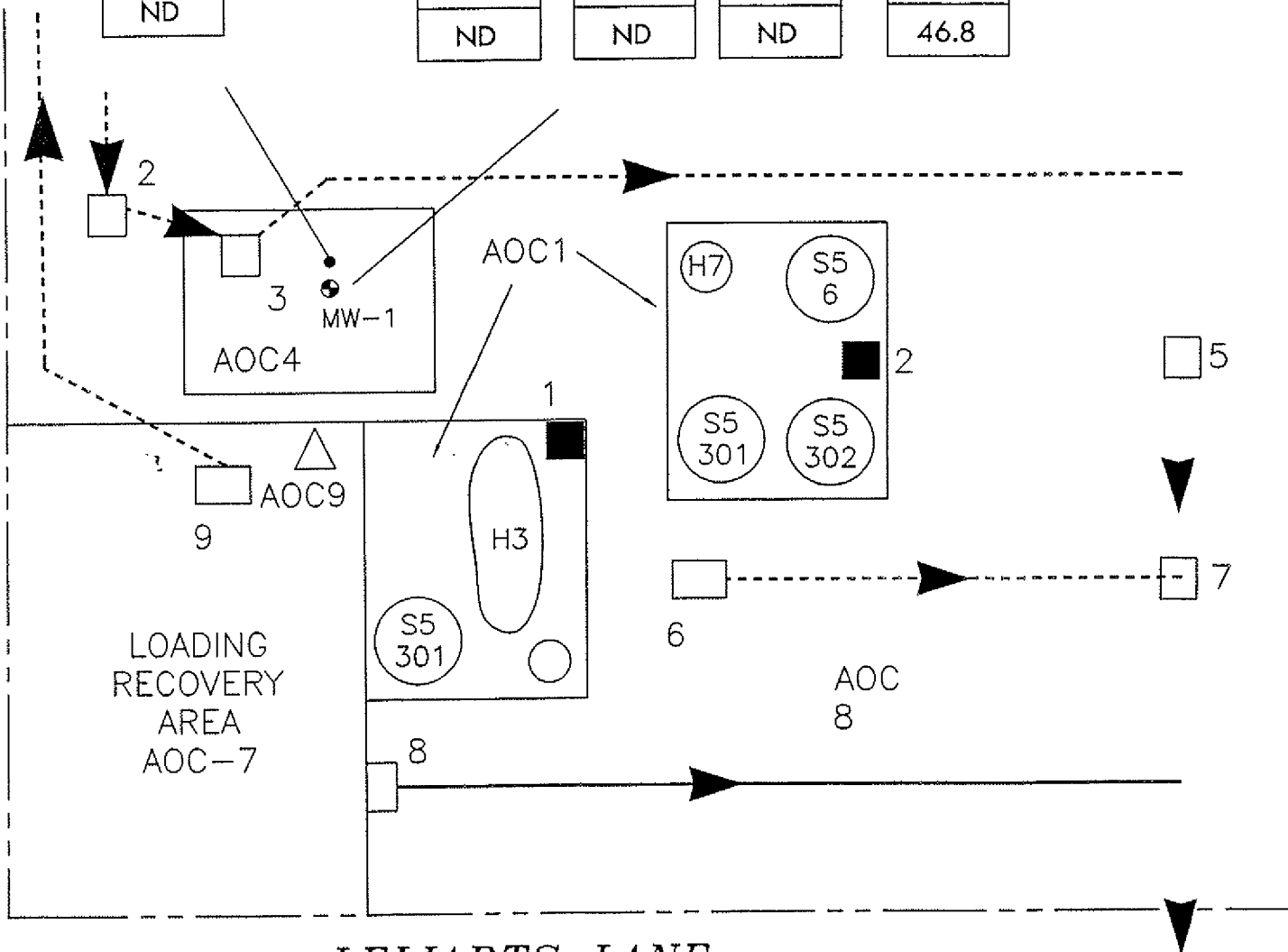
SB-01
7.95
0.385
5.42
4.66
28.80
ND

MW-1A
51.2
1.56
13
20.8
43.2
ND

MW-1B
40.5
0.649
18.4
31.4
23.5
ND

MW-1C
3.82
0.29
1.97
1.64
10.9
ND

GW
MW-1
2320
1030
2320
1130
9370
46.8



LELIARTS LANE

LEGEND:

- - SUMP
 ○ - ABOVE GROUND STORAGE TANK
 □ - FLOOR DRAIN / CATCH BASIN
 △ - AOC

SB-01	Sample Identification
1.0	Chlorobenzene
1.0	Benzene
1.0	Xylenes
1.0	1,3 Dichlorobenzene
1.0	1,4 Dichlorobenzene
1.0	Vinyl Chloride



FIGURE #

5

SITE PLAN

FINETEX, INC.
418 FALMOUTH AVENUE
ELMWOOD PARK, NEW JERSEY

Scale: not to scale

Drawn By: JCH

Date: 12/11/02

Project #	99704
-----------	-------



BRENNAN ENVIRONMENTAL, INC.

B Great Meadow Lane E. Hanover, NJ 07936
Tel: (973)781-1801



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDICES





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 1



NJDEP CORRESPONDENCE



James E. McGreevey
Governor

State of New Jersey
Department of Environmental Protection
Bureau of Field Operations
401 East State Street
PO Box 435
Trenton NJ 08625-0435

Bradley M. Campbell
Acting Commissioner

MAR 7 - 2002

John M. Brennan
Brennan Environmental, Inc.
8D Great Meadow Lane
East Hanover NJ 07936

Re: Finetex Incorporated
410, 416 & 418 Falmouth Avenue Elmwood Park Boro, Bergen County
ISRA Case # E20010242

This is to advise you, as authorized agent, of the status of the investigation initiated under the provisions of the Industrial Site Recovery Act (N.J.S.A 13:1K-6 et seq.) Based upon our review of the most recent information provided, the status of the submissions are:

General Information Notice
Preliminary Assessment & Site Investigation Report

Complete
Received

Based on the information provided in your Preliminary Assessment and Site Investigation Report, one historic area of environmental concern requires further investigation, specifically the former (6) underground storage tanks' (UST) excavation in which the tanks were removed in 1988. Please take one sample through the former UST excavation and have this sample analyzed for VOC+10, B/N+15 and TPHC and submit the results, when available to me, the case manager.

Kindly refer to the referenced case name and number in all future correspondence. Any questions you may have on this case should be directed to me the assigned case manager at (609) 633-1428.

Sincerely,


Jeannette Cleary
Bureau of Field Operations

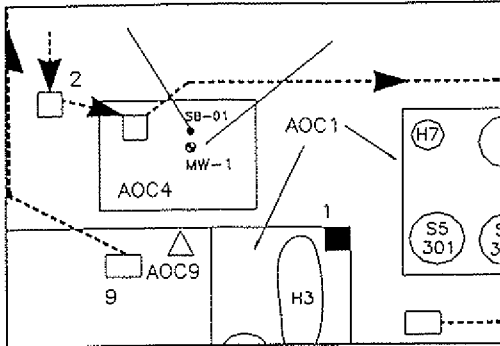


BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 2


SOIL BORING LOGS

Site Sketch for SB-01



Soil Boring Log for:

SB-01

BEI Job # 99704
 Site Name: Finetex
 Boring Install Date: May 24, 2002
 Type of Bit: na
 Sample: 3.0' macrocore sampler
 Geologist: A. Biskupski

Project: ISRA
 Site Location: 418 Falmouth Ave., Elmwood Park, NJ
 Boring Location: Former UST Area
 Drilling Company: BEI
 Drill Rig: Port-A-Probe
 Driller: Andrew Biskupski

Depth (ft.)	Sample ID and Depth	PID (MUJ)	Recovery (feet)	Soil/Geologic Description	Depth (ft.)	Boring Diagram and Details
1		0.0	1.5	0 - 3.0' Concrete to 0.5'. Brown CLAY w/ fine - medium Gravel. (dry)	1	
2		0.0			2	
3		3.0			3	
4		1.0			4	
5		75.0			5	
6	SB-01	5.0	2	3.0 - 6.0' Brown SAND w/ fine Gravel. (moist 4 - 5 feet)	6	
7				Boring advanced to 6 feet below grade. Sample collected at 5.5 - 6 feet below grade. No evident ground water encountered. Soils moist at 4 to 5 feet below grade.	7	
8					8	
9					9	
10					10	
11					11	
12					12	
13					13	
14					14	
15					15	
16					16	
17					17	
18					18	
19					19	
20					20	
21					21	
22					22	
23					23	
24					24	
25					25	



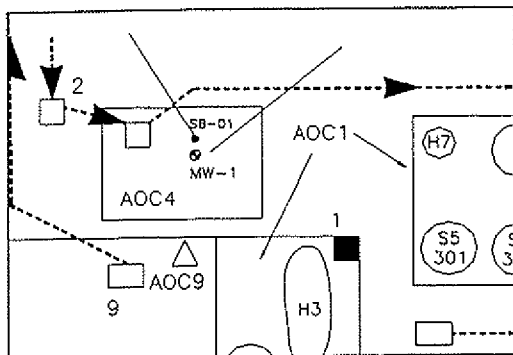
BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 3



MONITORING WELL INSTALLATION FORMS

Site Sketch for MW-1



Permanent Well Soil Log and Well Construction for: MW-1

BEI Job #: 99704

Site Name: Finetex, Inc

Well Permit #: 26-65907

Well Install Date: November 2, 2002

Well Removal Date: na

Type of Bit: na

Sample: 5.0' stainless steel macrocore

Auger Type: 6"

Project: monitoring well and boring installation

Site Location: former tank area (AOC #4)

Well Location: southwest portion of property

Well Driller: S&S Subsurface Investigations, Inc

Well Install Rig: Geoprobe 66DT

Driller/Helper: Art Salvatore/Pete Dudley

Geologist: John Hernandez

Depth (ft.)	Sample ID and Depth	PID (NU)	Recovery (Inches)	Soil/Geologic Description	Depth (ft.)	Well Diagram and Details
1			3.8	0.0-0.5 Concrete and Gravel	1	8.0" Diam.
2		0.0		0.5-2.0 Brown SAND, some Silt (dr)	2	Bentonite: NA
3		0.0		2.0-5.0 Brown-gray SILT, some Clay, some fine Sand (moist)	3	Cement Collar
4	MW1-a	148.3			4	Sand Pack: 2.0-15.0'
5	MW1-b	212.3			5	Solid Riser
6	MW1-c	176.8			6	Screen: PVC
7		220.4	4.5	5.0-7.0 Brown gray SILT, some Clay, little fine Sand (wet at ~6.0 ft)	7	Schedule 40, 0.01" slot PVC, 2.0" diam., 3.0' to 15.0'
8		70.6		7.0-10.0 Brown gray SILT, some Clay, trace fine Sand (wet)	8	Well - Depth: 15.0'
9		22.0			9	Diameter: 2.0"
10		20.0			10	G.W. Enc.: ~6.0
11		18.0			11	Depth to PVC Rim: 3"
12		16.0			12	
13		21.4			13	
14		22.0			14	
15		21.0			15	Sand pack
16		22.3			16	Screen
17		24.0			17	Threaded end cap
18		9.6	4.5	10.0-15.0 Gray brown CLAY and SILT, trace fine Sand	18	
19		6.3			19	
20		5.0			20	
21		6.0			21	
22		9.5			22	
23		28.1			23	
24		40.2			24	
25		33.0			25	
		15.6				
		18.0				
				Well completed at 15.0 feet bsg		
				Ground water encountered at ~6.0'		



MONITORING WELL CERTIFICATION - FORM A - AS BUILT CERTIFICATION

(One form must be completed for each well)

Name of Owner: Finetex, Inc.
Name of Facility: Finetex, Inc.
Location: 418 (includes 410 & 416) Falmouth Avenue
ISRA Case #: E2001242

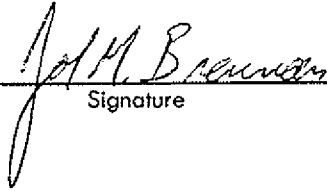
CERTIFICATION

Well Permit Number: 26-65907
Owner's Well Number (As shown on the application or plans): MW-1
Well Completion Date: 11/2/2002
Distance from Top of Casing (cap off) to Ground Surface (one-hundredth of a foot): 0.25
Total Depth of Well to the nearest 1/2 foot: 15.0
Depth to Top of Screen (or Top of Open Hole) From Top of Casing (one-hundredth of a foot): 3.00
Screen Length (or length of open hole) in feet: 12.0
Screen or Slot Size: 0.02
Screen or Slot Material: PVC
Casing Material: (PVC, Steel or Other-Specify): PVC
Casing Diameter (inches): 2.0
Static Water Level From Top of Casing at the Time of installation (one-hundredth of a foot): 6.44
Yield (gallons per minute): 0.5 g.p.m.
Development Technique (Specify): submersible pump
Length of Time Well is Developed/ Pumped or Bailed: 0 Hour(s) 30 Minute(s)
Lithologic Log: Pumped
Attached

Authentication

I certify under penalty of law that I have personally examined and am familiar with the information submitted in this document and all attachments and that, based on my inquiry of those individuals immediately responsible for obtaining the information, I believe the submitted information is true, accurate and complete. I am aware that there are significant penalties for submitting false information, including the possibility of fine and imprisonment.

John Brennan
Name (Type or Print)


Signature

NJDEP license # 0010045
Certification or License No.

Certification by Executive Officer or Duly Authorized Representative

Name (Type or Print)

Signature

Title

Date

DWR-138 M
8/00

New Jersey Department of Environmental Protection
Bureau of Water Allocation
MONITORING WELL RECORD

Well Permit No. 26 - 65907
Atlas Sheet Coordinates 26 - 03 - 181

OWNER IDENTIFICATION - Owner KINETIX INC
Address 418 FAIRMOUTH AVE
City ELMWOOD PARK State NJ Zip Code _____

WELL LOCATION - If not the same as owner please give address. Owner's Well No. _____
County BERGEN Municipality ELMWOOD PARK BO Lot No. 14 & 15 Block No. 803
Address 418 FAIRMOUTH AVE

TYPE OF WELL (as per Well Permit Categories) MONITORING DATE WELL STARTED 11 / 2 / 02
Regulatory Program Requiring Well _____ DATE WELL COMPLETED 11 / 2 / 02
Case I.D.# _____

CONSULTING FIRM/FIELD SUPERVISOR (if applicable) Brennan Environmental Tel. (973) 781-1801

WELL CONSTRUCTION

Total depth drilled 15.5 ft.
Well finished to 15.5 ft.

Borehole diameter:
Top 12 in.
Bottom 12 in.

Well was finished: ☐ above grade
☒ flush mounted

If finished above grade, casing height (stick up) above land surface _____ ft.

Was steel protective casing installed?
☐ Yes ☒ No

Static water level after drilling 6.5 ft.

Water level was measured using tape

Well was developed for .5 hours
at 2 gpm

Method of development 12V Pump

Was permanent pumping equipment installed? ☐ Yes ☒ No

Pump capacity _____ gpm

Pump type: _____

Drilling Fluid _____ Type of Rig Geoprobe 16

Health and Safety Plan submitted? ☒ Yes ☐ No

Level of Protection used on site (circle one) None (D) C B A

I certify that I have constructed the above referenced well in accordance with all well permit requirements and applicable State rules and regulations.

Drilling Company S & S SUBSURFACE INVEST.

Well Driller (Print) Arthur Salvatore

Driller's Signature Arthur Salvatore

Registration No. MD1264 Date 11 / 5 / 02

Note: Measure all depths from land surface	Depth to Top (ft.)	Depth to Bottom (ft.)	Diameter (inches)	Material	Wgt./Rating (lbs/sch no.)
Single/Inner Casing	<u>.5</u>	<u>3.5</u>	<u>2</u>	<u>PVC</u>	<u>sch 40</u>
Middle Casing (for triple cased wells only)					
Outer Casing (largest diameter)					
Open Hole or Screen (No. Used, OIO)	<u>3.5</u>	<u>15.5</u>	<u>2</u>	<u>PVC</u>	<u>sch 40</u>
Blank Casings (No. Used)					
Tail Piece					
Gravel Pack	<u>2.5</u>	<u>15.5</u>		<u>#1 Gravel</u>	
Grout	<u>.5</u>	<u>2.5</u>		<u>Neat Cement Bentonite</u>	<u>94</u> lbs. <u>5</u> lbs.

Grouting Method HSA
Drilling Method Pressure / TCMAC

GEOLOGIC LOG

Note each depth where water was encountered in consolidated formations.

0'-5' fill
5'-15.5' SP

**AS-BUILT WELL LOCATION
(NAD 83 HORIZONTAL DATUM)**

NJ STATE PLANE COORDINATE IN US SURVEY FEET

NORTHING: _____ EASTING: _____

LATITUDE: _____ OR _____ LONGITUDE: _____

COPIES: White - DEP Canary - Driller Pink - Owner Goldenrod - Health Dept.

DWR-133M
11/01

STATE OF NEW JERSEY
DEPARTMENT OF ENVIRONMENTAL PROTECTION
TRENTON, NJ

MONITORING WELL PERMIT

Permit No.

2665907

Mall To:
NJDEP
BUREAU OF WATER ALLOCATION
PO BOX 426
TRENTON, NJ 08625-0426

VALID ONLY AFTER APPROVAL BY THE D.E.P.

COORD #:

26.03.1.81

Owner Finetex, Inc.

Driller St. S. Subsurface Investigations, Inc.

Address 418 Falmouth Ave.
Elmwood Park, NJ

Address 936 Atlantic Ave.
Pt. Pleasant, NJ 08742

Name of Facility (same)

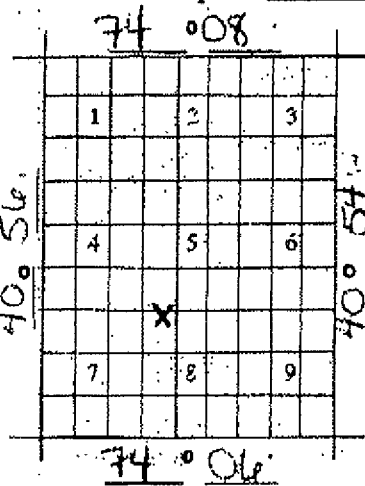
Address _____

Diameter of Well(s)	2	Feet	Proposed Depth of Well(s)	25	Feet
Type of Well	Monitoring		Will pumping equipment be utilized?	YES ☐ NO ☒	
Applied for (max. 10)	1		Is Yes, give pump capacity		capacities GPM

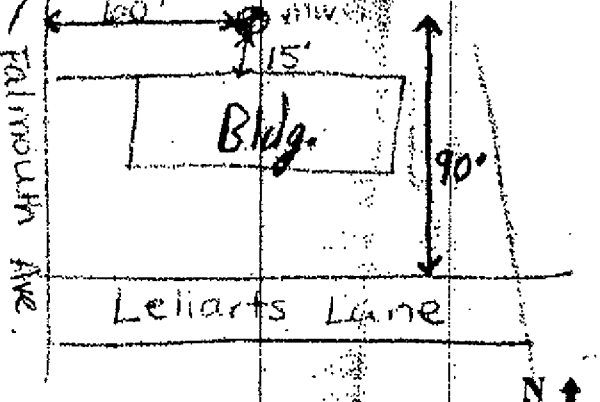
LOCATION OF WELL(S)

Lot # 14 + 15 Block # 803 Municipality Elmwood Park County Bergen

State Atlas Map No. 26



Draw sketch of well(s) nearest roads, buildings, etc. with marked distances in feet. Each well MUST be labeled with a name and/or number on the sketch.



PROPOSED WELL LOCATION (NAD 83 HORIZONTAL DATUM)
NJ STATE PLANE COORDINATE IN US SURVEY FEET

NORTHING: _____ EASTING: _____
OR
LATITUDE: _____ LONGITUDE: _____

FOR MONITORING WELLS, RECOVERY WELLS, OR PERZOMETERS, THE FOLLOWING MUST BE COMPLETED BY THE APPLICANT. PLEASE INDICATE WHY THE WELLS ARE BEING INSTALLED:

- ☐ RCRA Site
- ☐ Spill Site
- ☐ Underground Storage Tank Site
- ☒ ISRA Site
- ☐ Operational Ground Water Permit Site
- ☐ CERCLA (Superfund) Site
- ☐ Pretreatment and Residuals Site
- ☐ Water and Hazardous Waste Enforcement Case
- ☐ Water Supply Aquifer Test Observation Well
- ☐ Other (explain): _____

ISRA * ENVIRONMENTAL DIVISION

05-001-3 04-3-12

This Space for Approval Stamp

WELL NO. 1 / APPROVED
NJDEP

OCT 7 2002

BUREAU OF WATER ALLOCATION

FOR D.E.P. USE ☐ Issuance of this permit is subject to the conditions attached. (see next page)

SEE REVERSE SIDE FOR IMPORTANT PROVISIONS PERTAINING TO THIS PERMIT.
In compliance with N.J.S.A. 58:44-14, application is made for a permit to drill a well as described above.

Date 10/1/02

Signature of Driller [Signature]

Registration No. HD12104

Signature of Property Owner [Signature] For Bob Scala of Finetex, Inc.



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 4



PURGE GUIDE

Ground Water Sampling Field Measurement Guide

Site Name: Finetex, Inc.				Date/Time: 11/18/2002			
Address: 418 Falmouth Avenue, Elmwood Park, NJ				Weather: P. Cloudy, windy, 45			
BEI Case No.: 99704				Tech(s): JH			
Well ID#: MW-1							
Pre-Purge Data:							
PID /FID Reading (mu):	67.4						
Product Thickness (ft):	NA						
Well Diameter (Inches)	2						
Total Depth of Well (ft):	13.81						
Top of casing	49						
Depth to Water (ft):	5.06						
Volume of water in well (gal):	1.4						
pH:	8.9						
Dissolved oxygen (mg/L):	3.3						
Temperature (°C):	11.4						
Specific conductivity (uS):	2.6						
Post-Purge Data:							
Purge Start Time:	9:54						
Purge Stop Time:	10:07						
Purge Method:	pump						
Purge Rate (gpm):	<1						
Total Volume Purged (gal):	5.4						
Depth to Water (ft):	9.08						
pH:	7.69						
Dissolved oxygen (mg/L):	2.3						
Temperature (°C):	14.4						
Specific conductivity (uS):	2.22						
Before Sampling Data:							
Depth to Water (ft):	8.86						
Sampling Start Time:	10:10						
After Sampling Data:							
Sampling End Time:	10:16						
pH:	7.6						
Dissolved oxygen (mg/L):	2.2						
Temperature (°C):	14						
Specific conductivity (uS):	2.3						
Sample Method:	bailer						
Field Observations							
Recharge Rate (gpm)	slow						
Sheen	none						
Odor	trace						
Turbidity	minor						
Notes:							
Pump = The purge device used was a peristaltic pump with dedicated tubing.							
Bailer = The sampling device used was a disposable, dedicated bailer.							
mu = Meter units							





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 5



LABORATORY ANALYTICAL SOIL DATA REPORT (LAB CASE #E02-3831)



ANALYTICAL DATA REPORT

Brennan Environmental, Inc.
8 Great Meadow Lane
Unit D
East Hanover, NJ 07936

Project Name: **FINETEX - 99704**
IAL Case Number: **E02-3831**

These data have been reviewed and accepted by:


Michael H. Leftin, Ph.D.
Laboratory Director

273 Franklin Road
Randolph, NJ 07869
Phone: 973 361 4252
Fax: 973 889 5288



IAL is a NELAC New Jersey Certified Lab (14751) and maintains certification in Connecticut (PH-0698), New York (11402), Rhode Island (00126), Florida (E87670) and in the Department of Navy IR QA Program.

Sample Summary

Case No. **E02-3831**
Project Name FINETEX - 99704
Customer Brennan Environmental, Inc.
Received On 5/28/2002@11:15

<u>Lab ID</u>	<u>Client Sample ID</u>	<u>Depth Top / Bottom</u>	<u>Sampling Time</u>	<u>Matrix</u>	<u># of Cont.</u>
3831-001	SB-01	n/a	5/25/2002@10:21	Soil	2

INTEGRATED ANALYTICAL LABORATORIES, LLC.

TABLE OF CONTENTS

	<u>Page</u>
Qualifiers	1
Conformance / NonConformance Summary	2
Laboratory Deliverables Check List	3
GC/MS NonConformance Summary	4
PHC NonConformance Summary	5
Summary Report	6
Analytical Results	
PP VOA + 10	8
BN + 15	10
General Analytical	
TPHC	13
Methodology Summary *	
Quality Control	
PP VOA + 10	14
Tuning Results Summary	
Method Blank Results Summary	
Calibration Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Internal Standard Summary	
Chromatograms	
BN + 15	51
Tuning Results Summary	
Method Blank Results Summary	
Calibration Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Internal Standard Summary	
Chromatograms	
Petroleum Hydrocarbons	100
Blank Results Summary	
Spike Sample Results Summary	
Duplicate Sample Results Summary	
Calibration Curve	
Sample Tracking	
Chains of Custody	104
Laboratory Chronicle	107

* Methodology is included in the IAL Project Information Page

INTEGRATED ANALYTICAL LABORATORIES, LLC.

MATRIX QUALIFIERS

- A - Indicates the sample is an Aqueous matrix.
- O - Indicates the sample is an Oil matrix.
- S - Indicates the sample is a Soil, Sludge or Sediment matrix.
- X - Indicates the sample is an Other matrix as indicated by Client Chain of Custody.

DATA QUALIFIERS

- B - Indicates the analyte was found in the Blank and in the sample. It indicates possible sample contamination and warns the data user to use caution when applying the results of the analyte.
- C - Common Laboratory Contaminant.
- D - The compound was reported from the Diluted analysis.
- D.F. - Dilution Factor.
- E - Estimated concentration, reported results are outside the calibrated range of the instrument.
- J - Indicates an estimated value. The compound was detected at a value below the method detection limit but greater than zero. For GC/MS procedures, the mass spectral data meets the criteria required to identify the target compound.
- MDL - Method Detection Limit.
- MI - Indicates compound concentration could not be determined due to Matrix Interferences.
- NA - Not Applicable.
- ND - Indicates the compound was analyzed for but Not Detected at the MDL.

REPORT QUALIFIERS

All solid sample analyses are reported on a dry weight basis.

All solid sample values are corrected for original sample size and percent solids.

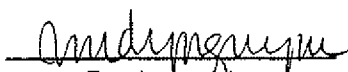
INTEGRATED ANALYTICAL LABORATORIES, LLC.

CONFORMANCE / NONCONFORMANCE SUMMARY

Integrated Analytical Laboratories, LLC. received one (1) soil sample(s) from Brennan Environmental, Inc. (Project: FINETEX - 99704) on May 28, 2002 for the analysis of:

- (1) PP VOA + 10
- (1) BN + 15
- (1) TPHC

A review of the QA/QC measures for the analysis of the sample(s) contained in this report has been performed by:


Reviewed by


Date

0002

INTEGRATED ANALYTICAL LABORATORIES, LLC.

LABORATORY DELIVERABLES CHECK LIST

Lab Case Number: E02-3831

	Check If Complete
1. Cover Page, Title Page listing Lab Certification #, facility name & address and date of report preparation.	<u>✓</u>
2. Table of Contents.	<u>✓</u>
3. Summary Sheets listing analytical results for all targeted and non-targeted compounds.	<u>✓</u>
4. Summary Table cross-referencing Field ID's vs. Lab ID's.	<u>✓</u>
5. Document bound, paginated and legible.	<u>✓</u>
6. Chain of Custody.	<u>✓</u>
7. Methodology Summary.	<u>✓</u>
8. Laboratory Chronicle and Holding Time Check.	<u>✓</u>
9. Results submitted on a dry weight basis (if applicable).	<u>✓</u>
10. Method Detection Limits.	<u>✓</u>
11. Lab certified by NJDEP for parameters or appropriate category of parameters or a member of the USEPA CLP.	<u>✓</u>
12. NonConformance Summary.	<u>✓</u>

Amel M. M. M.
QC Reviewed by

6/6/02
Date

INTEGRATED ANALYTICAL LABORATORIES
CONFORMANCE/NONCONFORMANCE SUMMARY
GC/MS VOLATILE ANALYSIS

Lab Case Number:

E02 -

3831

	<u>No</u>	<u>Yes</u>
1. Chromatograms Labeled/Compounds Identified (Field Samples and Method Blanks).	_____	_____/_____ ✓
2. GC/MS Tuning Specifications:		
a. BFB Passed	_____	_____/_____ ✓
3. GC/MS Tuning Frequency - Performed every 24 hours for 600 series, 12 hours for 8000 series and 8 hours for 500 series.	_____	_____/_____ ✓
4. GC/MS Calibration - Initial calibration performed within 30 days before sample analysis and continuing calibration performed within 24 hours before sample analysis for 600 series, 12 hours for 8000 series	_____	_____/_____ ✓
5. GC/MS Calibration Requirements:		
a. Calibration Check Compounds	_____	_____/_____ ✓
b. System Performance Check Compounds	_____	_____/_____ ✓
6. Blank Contamination - If yes, list compounds and concentrations in each blank:	_____/_____ ✓	_____

7. Surrogate Recoveries Meet Criteria (If not met, list those compounds and their recoveries which fall outside the acceptable range)	_____	_____/_____ ✓

If not met, were the calculations checked and the results qualified as "estimated"?	_____	_____/_____ N/A
8. Matrix Spike/Matrix Spike Duplicate meet criteria (if not, list those compounds and their recoveries/% differences which fall outside the acceptable range)	_____	_____/_____ ✓

9. Internal Standard Area/Retention Time Shift meet criteria	_____	_____/_____ ✓
10. Extraction Holding Time Met	_____	_____/_____ N/A
If not met, list number of days exceeded for each sample:	_____	

11. Analysis Holding Time Met	_____	_____/_____ ✓
If not met, list number of days exceeded for each sample:	_____	

12. Sample Dilution Performed	_____/_____ ✓	
High Target Compounds	High Nontarget Compounds	Matrix Interference
[]	[]	[]
		Other
		[]

13. Comments:

Organics Manager

5/30/02

Date

Lab Case Number: E02 - 3831

13. Comments:

Date _____

INTEGRATED ANALYTICAL LABORATORIES
CONFORMANCE/NONCONFORMANCE SUMMARY
PHC ANALYSIS

Lab Case Number: E02 - 3831

- | | <u>No</u> | <u>Yes</u> |
|--|-----------|------------|
| 1. Blank contamination
If yes, list the sample and the concentration in each blank:

_____ | <u>✓</u> | _____ |
| 2. Matrix Spike/Matrix Spike Duplicate Recoveries Meet Criteria (If not met, list the samples and corresponding recovery which falls outside the acceptable range)

_____ | _____ | <u>✓</u> |
| 3. IR spectra submitted for all standards, blanks & samples (as required).

_____ | _____ | <u>✓</u> |
| 4. Chromatograms submitted for all standards, blanks & samples if GC fingerprinting was conducted. | _____ | <u>NA</u> |
| 5. Extraction Holding Time Met
If not met, list number of days exceeded for each sample:

_____ | _____ | <u>✓</u> |
| 6. Analysis Holding Time Met
If not met, list number of days exceeded for each sample:

_____ | _____ | <u>✓</u> |

Additional Comments: _____

Paul Cornista
Dept. Supervisor

5.29.02
Date

SUMMARY REPORT

Client: Brennan Environmental, Inc.

Project: FINETEX - 99704

Lab Case No.: E02-3831

Lab ID:	3831-001		
Client ID:	SB-01		
Matrix:	Soil		
Sampled Date:	5/25/2002		
PARAMETER(Units)	Conc	Q	MDL
Volatiles (ppb)			
Benzene	385	J	793
Chlorobenzene	7950		793
Ethylbenzene	1080		793
Total Xylenes	5420		793
1,3-Dichlorobenzene	4660		793
1,4-Dichlorobenzene	28800		793
1,2-Dichlorobenzene	801		793
TOTAL VO's:			
	49096	J	
TOTAL TIC's:			
	20240		
TOTAL VO's & TIC's:			
	69336	J	
Semivolatiles - BN (ppb)			
1,3-Dichlorobenzene	603		107
1,4-Dichlorobenzene	4480		107
1,2-Dichlorobenzene	123		107
1,2,4-Trichlorobenzene	3270		107
Naphthalene	2140		107
2-Methylnaphthalene	1870		107
Phenanthrene	80.6	J	107
Fluoranthene	71.8	J	107
bis(2-Ethylhexyl)phthalate	92.1	J	107
TOTAL BN'S:			
	12730.5	J	
TOTAL TIC's:			
	128810		
TOTAL BN'S & TIC's:			
	141541	J	
General Analytical			
Total Petroleum Hydrocarbons (ppm)	892		25.0

ND = Analyzed for but Not Detected at the MDL

J = The concentration was detected at a value below the MDL

All qualifiers on individual Volatiles & Semivolatiles are carried down through summation.

0007



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

UST Closure / Site Investigation Report



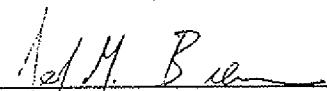
Finetex, Inc.
418 (includes 410 & 416 Falmouth Avenue)
Block 803, Lots 14, 15
Elmwood Park, Bergen County, New Jersey
ISRA Case #E2001242

November 8, 2001
BEI Job No. 99704

Prepared for:
Finetex Inc.
418 Falmouth Avenue
Elmwood Park, NJ 07407

Prepared by:
Brennan Environmental Inc.
8D Great Meadow Lane
East Hanover, NJ 07936

Prepared by:



John M. Brennan, President
NJDEP License Number 0010045

TABLE OF CONTENTS

1.) INTRODUCTION	1
1.1) HISTORICAL INFORMATION	1
2.0) PHYSICAL SETTING	1
2.1) SITE GEOLOGY	1
2.2) SITE HYDROGEOLOGY	2
2.3) SITE TOPOGRAPHY & SURROUNDING LAND USE	2
2.4) BASELINE ECOLOGICAL ASSESSMENT AND RECEPTOR ASSESSMENT	2
3.0) TECHNICAL OVERVIEW	3
3.1) RELIABILITY OF ANALYTICAL DATA	3
3.2) SITE CONTAMINATION SUMMARY	4
3.3) SIGNIFICANT EVENTS AND/OR SEASONAL VARIATION INFLUENCING THE INVESTIGATION	4
4.0) FINDINGS/RECOMMENDATIONS	4
4.1) AREAS OF CONCERN SITE INVESTIGATION RESULTS	4
4.2) SPECIFIC AREAS OF CONCERN DISCUSSION	5
4.2.1) <i>Detailed Description of Each Area of Concern</i>	5
4.2.2) <i>UST Closure and Site Assessment of 550-gallon UST (AOC-6)</i>	5
4.2.3) <i>Site Investigation of area of former 1,000-gallon UST (AOC-5)</i>	5
4.2.4) <i>Areas with Specific Changes in Sampling Protocol Due to Field Conditions</i>	6
4.2.5) <i>Significance of Tentatively Identified Compounds</i>	8
4.2.6) <i>Recommendations for Areas of Concern</i>	8
5.0) CERTIFICATIONS:	10

TABLE

SUMMARY OF SOIL SAMPLE RESULTS (AOC-6)	Table 1
SUMMARY OF SOIL SAMPLE RESULTS (AOC-5)	Table 2
SUMMARY OF SOIL SAMPLE RESULTS (AOC-5)	Table 3

FIGURES

SITE LOCATION MAP	1
U.S.G.S. 7.5 MINUTE TOPOGRAPHIC QUADRANGLE MAP	2
COPY OF NJDEP WETLAND INVENTORY MAP	3
SITE MAP	4
SAMPLE LOCATION MAP	5

APPENDICES

COPY OF LOCAL CONSTRUCTION PERMIT AND INSPECTION APPROVAL	1
TANK CONTENTS WASTE DISPOSAL DOCUMENTATION	2
CLEAN FILL CERTIFICATION DOCUMENTATION	3
UST DISPOSAL DOCUMENTATION	4
PHOTOGRAPHIC LOG	5
BEI'S STANDARD QA/QC PLAN	6
PSW 2000	7
LABORATORY ANALYTICAL DATA REPORT	8



1.) INTRODUCTION

Brennan Environmental, Inc. (BEI) was retained by Finetex, Inc. (Finetex) to prepare this Site Investigation Report (SIR) for the facility located at 418 Falmouth Avenue, Block 803 Lots 14 and 15, Elmwood Park, Bergen County, New Jersey. The site location is indicated on Figure #1. The purpose of this report is to detail the actions that were implemented at the above referenced site in order to close by removal, one 550-gallon heating oil UST and to evaluate the soils around a former UST on the adjacent property (416 Falmouth Ave). This report was prepared in accordance with New Jersey Department of Environmental Protection's (NJDEP's) Technical Requirements for Site Remediation (N.J.A.C. 7:26E et.-seq.). BEI is certified by the NJDEP to perform UST closures and subsurface evaluations pursuant to N.J.S.A. 58:10A-24, (Certification #US00779).

1.1) Historical Information

A Preliminary Assessment Report conducted for the Finetex site, pursuant to ISRA Case No. E2001242, identified two areas of environmental concern that warranted additional investigation. This site investigation was directed at the two areas of environmental concern. Specifically the focus of concern was an inactive 550- gallon underground #2 heating oil tank system (AOC 6), and an area where a former 1,000 underground #2 heating oil tank was located (AOC 5).

This report was prepared in accordance with New Jersey Department of Environmental Protection's (NJDEP's) Technical Requirements for Site Remediation. This UST Closure/Site Investigation Report (SIR), details the tank closure process and post-excavation soil sampling activities, soil boring investigation, and BEI's evaluation of the post-excavation soil sampling data, which was compared to the current applicable NJDEP soil cleanup criteria. The total aggregate volume of the two former USTs was less than 2,000 gallons. therefore, the USTs are not subject to the underground Storage of Hazardous Substances Act (N.J.S.A. 58:10A-21), and a closure plan was not required.

2.0) PHYSICAL SETTING

2.1) Site Geology

The subject site is located in the Piedmont Physiographic Province of New Jersey. According to the USGS Bedrock Geologic Map of Northern New Jersey dated 1996, the bedrock in this section of the Piedmont Province of New Jersey is described as the Passaic Formation, which is Lower Jurassic and Upper Triassic in age and composed predominantly of conglomeratic sandstone with some shale. This sandstone is described as interbedded grayish-red to brownish-red, medium to fine grained and medium to thick bedded.



According to the Rutgers University Engineering Soil Survey of New Jersey (Report No. 4, Bergen and Hudson Counties) the overburden materials at the site are mapped as "GS12ig". The "GS" portion of the symbol signifies that the area is underlain by glacial stratified drift, composed of non-residual materials deposited during the Wisconsin glaciation. The general characteristic of the moraine is assorted, homogeneous materials, consisting predominantly of sand sizes, with varying amounts of silt, gravel, and occasionally clay. The "ig" indicates imperfect to good internal drainage conditions. Depth to the top of bedrock is expected to be greater than 10 feet.

The overburden soils encountered during the UST closure activities correlated well with the above description. Soils encountered in UST excavation area were as follows:

- 0-3' Light brown SILT, little clay, some coarse gravel and organic material
- 3-8.5' Dark brown CLAY, little coarse gravel

2.2) Site Hydrogeology

According to the Rutgers University Engineering Soil Survey of New Jersey, the drainage conditions for areas mapped as "GS12ig" are described as imperfect to good. During UST removal activities, no ground water was encountered by BEI in the excavation, which extended to a depth of 8.5 feet below site grade. According to site topography, groundwater is expected to flow in a southern direction.

2.3) Site Topography & Surrounding Land Use

The facility location is shown on an excerpt from the U.S.G.S. 7.5' Hackensack, Topographic Quadrangle Map (see Figure #2). According to the topographic quadrangle map, the site is located approximately 49 feet above mean sea level. Land use is a mixture of commercial and residential.

2.4) Baseline Ecological Assessment and Receptor Assessment

Wellhead Assessment: According to our review of NJ Geological Survey's computer program PSW 2000, there are six public potable wells within a 2,000-foot radius of the subject site. The wells are owned by the Garfield Water Department. A copy of the printout from the PSW 2000 software is provided as an attachment in Appendix 7 of this report.

Surface Water Body Assessment: The closest body of water to the site is the Fleischer Brook, located approximately 755 feet west of the subject property. Fleischer Brook appears to run southeast from the subject property. Fleischer Brook is not classified in NJDEP's surface water quality standards. Since no contaminants were detected above applicable NJDEP cleanup criteria, The Fleischer Brook is not considered a receptor of concern.



Wetland Assessment: Review of the 1991 New Jersey Wetlands Inventory Map for the Hackensack southwest quarter quadrangle, which is compiled by the NJDEP, indicates no wetland / state open water areas on or adjacent to the subject property. The subject site and surrounding area are designated as "Upland." A copy of the Wetland Inventory Map is included as Figure #3 of this report.

Baseline Ecological Assessment: In line with N.J.A.C. 7:26E-3.11, BEI performed a baseline ecological assessment for the subject site. The results are as follows:

- *Contaminants of Potential Ecological Concern:* Analytical results of the soil samples obtained from the subject tank areas at the property, indicate all contaminants were detected well below applicable NJDEP Soil Cleanup Criteria. Therefore, there are no contaminants of potential environmental concern.
- *Environmentally Sensitive Areas (ESA):* After a site inspection and review of the NJDEP wetlands map and a surface water body assessment, no ESAs were found on or adjacent to the subject site. Therefore, ESAs are not considered to be a concern.
- *Potential Migration Pathways:* Ground water was not encountered beneath the subject UST. Therefore, groundwater is not considered a potential migration pathway.

Since the baseline ecological assessment indicates that the three criteria above do not co-occur at the subject site, no further ecological investigation is warranted.

3.0) TECHNICAL OVERVIEW

3.1) Reliability of Analytical Data

This report details environmental services performed or monitored by BEI at the subject property. These services at the property included the removal of the UST system at AOC 6, collection of post-excavation soil samples, and collection of soil samples from a former tank area at AOC 5.

BEI's sampling activities were performed in accordance with the requirements outlined in the *NJDEP Field Sampling Procedures Manual* dated May 1992 and BEI's QA/QC methodology contained in Appendix #6. The soils encountered during removal activities were classified in the field according to the Unified Soil Classification System (USCS). All soil samples were collected using dedicated polyethylene plastic hand trowels. All soil samples were placed in precleaned amber glass bottles with Teflon lined caps and placed in a cooler maintained at 4°C in order to ensure proper preservation. Proper chain of custody documentation was maintained until delivery of the samples to the New Jersey certified laboratory. All samples were analyzed by Integrated Analytical Laboratories, L.L.C. (IAL) of Randolph, NJ, a New Jersey certified laboratory (NJDEP Certification #14751). The data quality assurance deliverables conform with



the Non-USEPA CLP Method as specified in the N.J.A.C. 7:26E - Technical Requirements for Site Remediation - Appendix A - Section IV. Minimum detection limits (MDL) for all compounds analyzed in the soil samples were below the current most restrictive NJDEP Soil Cleanup Criteria.

All post-excavation soil samples were compared to the NJDEP's Soil Cleanup Criteria last revised on May 12, 1999. The Residential Direct Contact Criteria was used throughout the soil column. However, in cases where the Impact to Ground Water Criteria was lower, the strictest criteria was used throughout the soil column. All locations of the samples and the USTs are shown on Figure #5 of this report.

3.2) Site Contamination Summary

Based on the results of the soil sampling, all samples revealed either not detected (ND) or levels of contaminants below the NJDEP strictest Soil Cleanup Criteria. Therefore, no further action is proposed.

3.3) Significant Events and/or Seasonal Variation Influencing the Investigation

There were no significant events and/or seasonal variation observed by BEI during the course of the closure activities that would influence the investigation, sampling procedures or analytical results.

4.0) FINDINGS/RECOMMENDATIONS

4.1) Areas of Concern Site Investigation Results

A construction permit was obtained from the City of Elmwood Park Building Department and a utility markout was ordered for both UST closures. All soil sample results revealed concentrations of targeted contaminants below the NJDEP strictest soil cleanup criteria or they were not detected. All specific tank closure activities and sample results are discussed in the following report sections.



4.2) Specific Areas of Concern Discussion

4.2.1) Detailed Description of Each Area of Concern

On October 11, 2001, a BEI representative was on site to supervise the removal of one 550-gallon #2 heating oil UST (AOC-6) and to install soil borings around an area of a former 1,000 gallon #2 heating oil tank (AOC-5). A site map indicating the location of the UST is provided in Figure #4.

4.2.2) UST Closure and Site Investigation (550 gallon UST AOC-6)

The tank removal contractor, Joe Royce, Inc. of Paterson, New Jersey, excavated soil exposing the top of the tank. All piping associated with the tank was drained back into the tank; the tank was pumped out following the American Petroleum Institute's recommended Practice for the Abandonment or Removal of Used Underground Service Station Tanks, and the New Jersey Uniform Construction Code (N.J.A.C. 5:23).

The top of the tank was uncovered and the tank was opened. The tank contents were removed by All State O.R.C. of West Milford, New Jersey, a licensed waste removal service (DEP #S 50015/JA-389) and the residual contents were squeegeed clean. A copy of the tank waste removal receipt is included in Appendix #2 for documentation purposes.

Upon removal, the tank was thoroughly inspected by BEI. The UST appeared to be in good condition with no holes. The Elmwood Park Building Inspector inspected the tank and confirmed that there were no signs of a release or discharge from the UST and gave approval to backfill the excavation. A copy of the certification is attached in Appendix 1.

The following observations were noted during the closure activities:

UST	Dimensions	Capacity	Construction	Condition	Contents
1	4' dia. x 5'8" len.	550 gals.	Steel	Good	#2 Heating Oil

- No petroleum odor, elevated PID readings, or soil staining were noted in the soil samples obtained from below the tank.
- The UST was observed to be in good condition with no holes.
- Neither ground water nor bedrock was encountered during the closure activities.

After post-excavation sampling was completed, (which is described in the following section of this report), the excavation was filled to grade with NJDEP approved fill material from Braen Stone Industries, Inc. A copy of the clean fill receipt is contained in Appendix 3.



4.2.3) UST Closure and Site Investigation Former 1,000 Gallon UST (AOC-5)

The tank removal contractor, Joe Royce, Inc. of Paterson, New Jersey, excavated soil exposing the top of the tank on October 2, 2000. All piping associated with the tank was drained back into the tank; the tank was pumped out following the American Petroleum Institute's recommended Practice for the Abandonment or Removal of Used Underground Service Station Tanks, and the New Jersey Uniform Construction Code (N.J.A.C. 5:23).

The top of the tank was uncovered and the tank was opened. The tank contents were removed by All State O.R.C. of West Milford, New Jersey, a licensed waste removal service (DEP #S 50015/JA-389) and the residual contents were squeegeed clean. A copy of the tank waste removal receipt is included in Appendix #2 for documentation purposes.

Upon removal, the tank was inspected by the Elmwood Park Building Inspector who confirmed that there were no signs of a release or discharge from the UST and gave approval to backfill the excavation. A copy of the certification is attached in Appendix 1.

As described below, since post-excavation soil samples were not collected, BEI performed soil borings around the tank area to confirm subsurface conditions.

4.2.4) Areas with Specific Changes in Sampling Protocol Due to Field Conditions

There were no significant events or seasonal variations that required changes in sampling protocol. The exact location of the former UST was indicated to BEI by the Finetex plant manager. Therefore, post-excavation sampling activities for AOC-6 were as follows: Two soil samples labeled "6S-1" and "6S-2" were collected at a frequency of one sample for every five linear feet along the center line of the UST. The samples were collected from 0"-6" below the base of the UST which corresponded to 6.0'-6.5' below site grade. An additional sample (6P-1) was collected from the 0"-6" interval below the piping, which corresponded to 2-2.5' below site grade. Since there was less than 15 linear feet of piping, only one sample was collected.

Post-excavation sampling activities for AOC-5 were as follows: Soil borings were advanced with 1.5" outside diameter augers and samples were taken with a 7/8" diameter soil probe. Three soil borings labeled "5-S1" through "5-S3" were taken along the centerline of the former UST at a frequency of one sample for every five linear feet along the center line of the UST. The samples were collected from a depth of 6.0'-6.5' below site grade which corresponded to 0"-6" below the base of the former UST. A piping sample was taken at a depth of 2-2.5', which corresponded to 0-6" below the base of the former piping. Since there was less than 15 linear feet of piping, only one sample was collected.

When the UST was closed in October 2000 the backfill used to fill the UST void was not certified clean fill. Therefore, a soil sample was also collected from the backfill material and



designated sample 5-F1. The sample was collected in accordance with N.J.A.C. 7:26E-6.4(d) and analyzed for PP+40 to document the quality of the backfilled material.

All post-excavation soil samples were stored in a cooler and transported to LAL for analysis. The soil samples were analyzed for total petroleum hydrocarbons (TPHCs) via EPA method 418.1 modified for soils with conditional analysis for volatile organics (VO+10). All VO +10 soil samples were preserved using the NJDEP methanol preservation method.

Laboratory analytical results for AOC-6 indicated that a TPHC concentration was detected in sample "6-S1" at 168 ppm and was detected at a level of 3,820 ppm in "6-S2". The piping sample "6-P1" indicated a TPHC level of 170 ppm. Since the soil sample result from "6-S2" was above 1,000 ppm, conditional analysis for volatile organics (VO+10) was required on that sample. The analytical results indicated that no volatile organics were detected above applicable NJDEP Cleanup Criteria and the total volatile organic contaminant concentration (TVOCC) was 21.93, significantly below the soil cleanup criteria of 1,000 ppm for TVOCC.

Table 1: Summary of Soil Sample Results (AOC-6)					
Sample ID:	6-S1	6-S2	6-P1	TB	NJDEP Residential Soil Cleanup Criteria
Lab ID:	7040-004	7040-005	7040-006	7040-007	
Sample Date:	10/11/2001	10/11/2001	10/11/2001	10/11/2001	
Sample Matrix:	Soil	Soil	Soil	Soil	
Sample Depth:	6-6.5'	6-6.5'	2-2.5'	NA	
Total Xylenes	~	0.863	~	ND	410
Total VOs	~	0.863	~	ND	~
Total TICs	~	21.07	~	~	~
Total VOs and TICs	~	21.93	~	ND	1,000
TPHC (ppm):	168	3,820	170	~	1,000/ 10,000
Notes: ND - Not detected, ~ Not analyzed					

Laboratory analytical results for AOC-5 indicated that a TPHC concentration was detected at a level of 516 ppm in sample "5-S1". TPHC's were not detected in samples "5-S2", "5-S3", or "5-P1". The analytical results of the backfilled material also indicated that no contaminants were detected above applicable NJDEP cleanup criteria. Summarized analytical results are depicted in Table 2 and 3, and the complete laboratory analytical data package for the post-excavation sampling is contained Appendix #8.

Table 2: Summary of Soil Sample Results (AOC# 5)					
Sample ID:	5-S-1	5-S-2	5-S-3	5-P1	NJDEP Residential Soil Cleanup Criteria
Lab ID:	7040-001	7040-002	7040-003	7040-008	
Sample Date:	10/11/2001	10/11/2001	10/11/2001	10/11/2001	
Sample Matrix:	Soil	Soil	Soil	Soil	
Sample Depth:	6-6.5'	6-6.5'	6-6.5'	2-2.5'	
TPHC (ppm):	516	ND	ND	ND	1,000/ 10,000



Table 3: Summary of Soil Sample Results (AOC-5) cont.		
Sample ID:	5-F1	NJDEP Residential Soil Cleanup Criteria
Lab ID:	7041-001	
Sample Date:	10/11/2001	
Sample Matrix:	Soil	
Sample Depth:	2-2.5'	
Parameters:		
Volatiles (ppm)		
Total VO's	ND	1,000
Total TICs	4.43	~
Total VO's & TICs	4.43	~
Semivolatiles (ppm)		
Total BNAs	ND	~
Total TICs	ND	~
Total BNA's & TICs	ND	~
PCBs	ND	~
Pesticides (ppm)	ND	0.49
Metals (ppm)		
Antimony	ND	14
Arsenic	1.70	20
Beryllium	ND	2
Cadmium	ND	39
Chromium	9.00	120,000
Copper	13.1	600
Lead	14.4	50
Mercury	0.02	14
Nickel	8.48	250
Selenium	ND	63
Silver	ND	110
Thallium	ND	2
Zinc	26.1	1500
Notes: ND -- Not detected		
-- Not analyzed		

4.2.5) Significance of Tentatively Identified Compounds

Based on laboratory data, the tentatively identified compounds which totaled 21.07 ppm are below NJDEP standards and therefore not of concern. The TICs consisted of two unknown alkanes, a substituted benzene, undecane, unknown cyclic hydrocarbon, and dodecane.

4.2.6) Recommendations for Areas of Concern

Former tank area AOC-5: The former 1,000-gallon UST was removed on October 2, 2000. Since the UST was not subject to the Underground Storage of Hazardous Substances Act, no post-excavation soil samples were collected at the time of closure. As detailed in this report, the UST was closed by Joe Royce Excavating. Documentation confirming the proper closure of the



UST is contained in this report. Since there was no evidence of a discharge when the UST was closed, as confirmed by the Elmwood Park municipal official and since all site investigation soil sample results were below the NJDEP's Cleanup Criteria of 10,000 ppm for #2 fuel oil, BEI recommends no further action for this area of concern.

Former UST AOC-6: As detailed in this report, the 550 gallon UST was closed by Joe Royce Excavating. Documentation confirming the proper closure of the UST is contained in this report. Since there was no evidence of a discharge when the UST was closed, as confirmed by BEI and the Elmwood Park municipal official, and since all post-excavation soil sample results were below the NJDEP's most restrictive soil cleanup criteria, BEI recommends no further action for this area of concern.



5.0 CERTIFICATIONS:

A. The following certification shall be signed by the highest ranking individual at the site with overall responsibility for that site or activity. Where there is no individual at the site with overall responsibility for that site or activity, this certification shall be signed by the individual having responsibility for the overall operation of the site or activity.

I certify under penalty of law that the information provided in this document is true, accurate and complete. I am aware that there are significant civil penalties for knowingly submitting false, inaccurate or incomplete information, and that I am committing a crime of the fourth degree if I make a written false statement which I do not believe to be true. I am also aware that if I knowingly direct or authorize the violation of any statute, I am personally liable for the penalties.

Typed/Printed Name Robert H. Seala

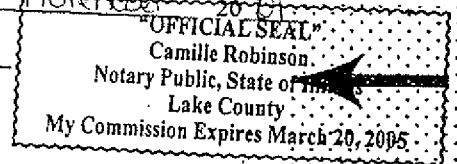
Title V.P.

Signature [Signature]

Date 11/17/01

Sworn to and Subscribed Before Me on this 17th Date of November 2001

[Signature]
Notary



**SIGN
HERE**

B. The following certification shall be signed as follows:

1. For a corporation, by a principal executive officer of at least the level of vice president;
2. For a partnership or sole proprietorship, by a general partner or the proprietor, respectively; or
3. For a municipality, State, Federal or other public agency, by either a principal executive officer or ranking elected official; or
4. For persons other than 1-3 above, by the person with the legal responsibility for the site.

Sign as Finetex

I certify under penalty of law that I have personally examined and am familiar with the information submitted herein and all attached documents, and that, based on my inquiry of those individuals immediately responsible for obtaining the information, I believe that the submitted information is true, accurate and complete. I am aware that there are significant civil penalties for knowingly submitting false, inaccurate or incomplete information, and that I am committing a crime of the fourth degree if I make a written false statement which I do not believe to be true. I am also aware that if I knowingly direct or authorize the violation of any statute I am personally liable for the penalties.

Typed/Printed Name Robert H. Seala

Title V.P.

Signature [Signature], V.P. Finetex

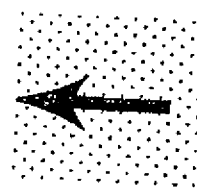
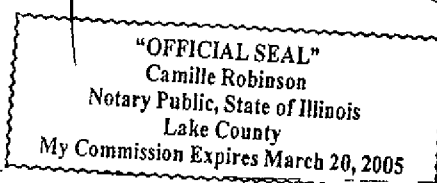
Date 11/17/01

Sworn to and Subscribed Before Me on this _____ Date of _____ 20 01

[Signature], V.P. Finetex, Inc.

Notary

[Signature]



**SIGN
HERE**

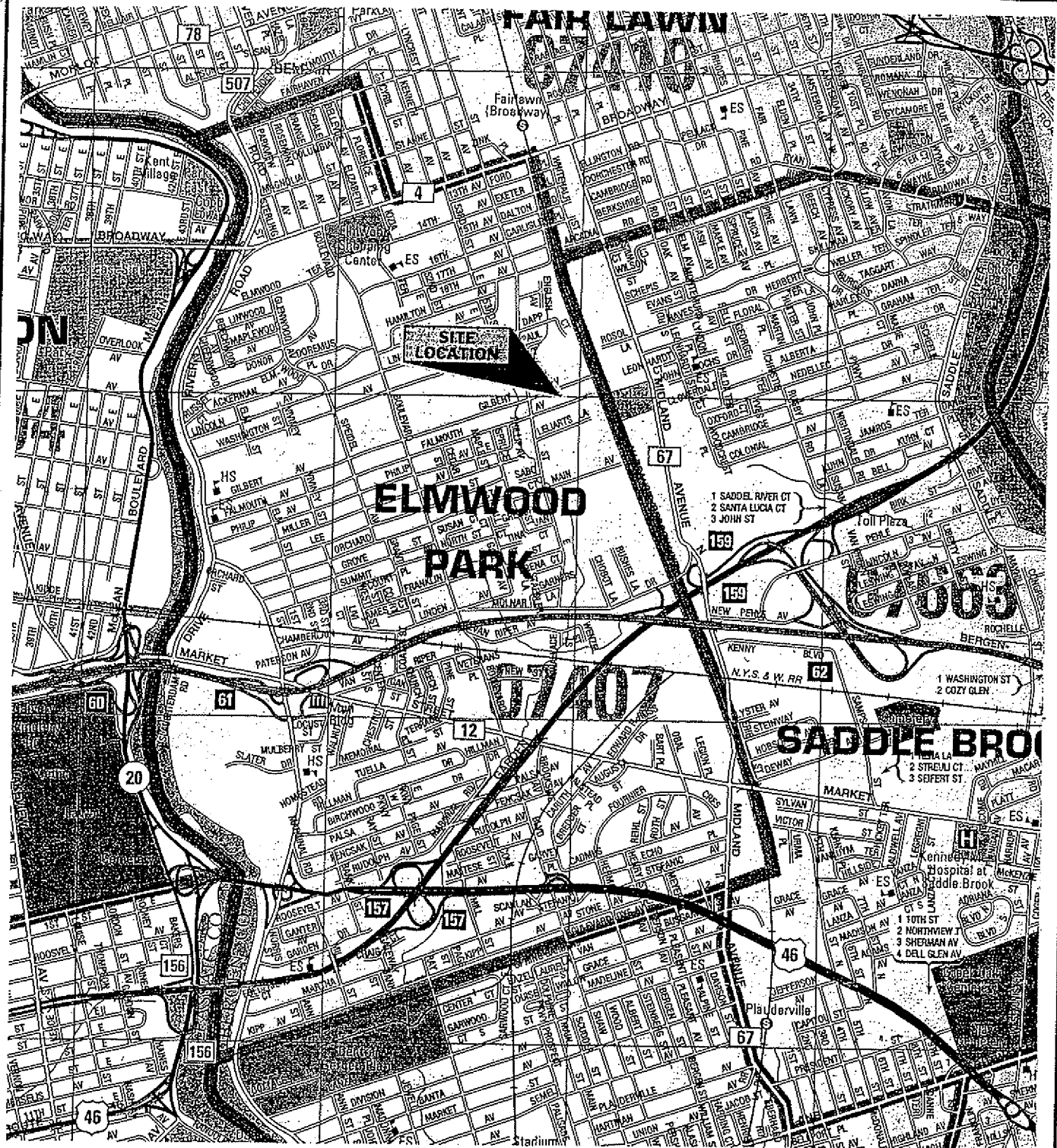
Sign as Finetex



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

FIGURES





SOURCE MAP

Bergen County, NJ Hagstrom Street Map



FIGURE:

Site Location Map

1

418 Falmouth Avenue
Elwood Park, Bergen County, NJ

Scale: As Shown

Drawn/Checked By: JPP/JMB

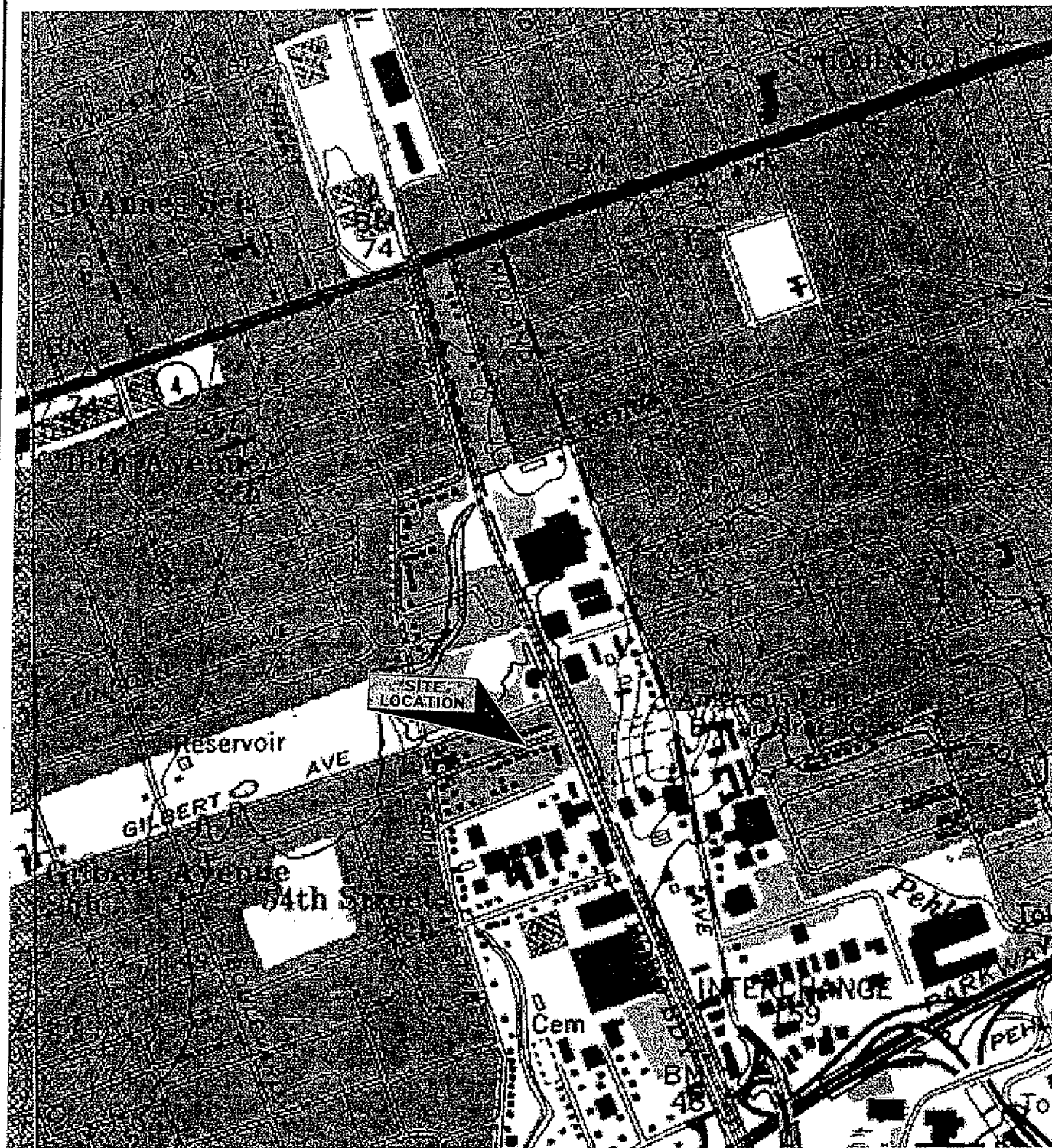
Date: 11/02/01

Project #: 99704



Brennan Environmental, Inc.

80 Great Meadow Lane, E. Hanover, NJ 07936 973.781.1001



SOURCE MAP

USGS TOPOGRAPHIC QUADRANGLE
HACKENSACK, NJ
7.5'SERIES
CONTOUR INTERVAL: 10'

SITE COORDINATES
40° 54' 44.28" N, 74° 06' 46.02" W

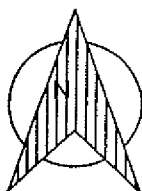


FIGURE:

Topographic Map

2

418 Falmouth Avenue
Elwood Park, Bergen County, NJ

Scale: As Shown

Drawn/Checked By: JPP/MB

Date: 11/02/01

Project #: 99704



Brennan Environmental, Inc.

80 Great Meadow Lane, E. Hanover, NJ 07936 973.781.1801



SOURCE MAP

NJDEP WETLANDS MAP BASED ON
HACKENSACK, NJ SW QUARTERQUADRANGLE



FIGURE:

Wetlands Inventory Map

3

418 Falmouth Avenue
Elwood Park, Bergen County, NJ

Scale: As Shown

Drawn/Checked By: JPP/JMB

Date: 11/02/01

Project #: 99704



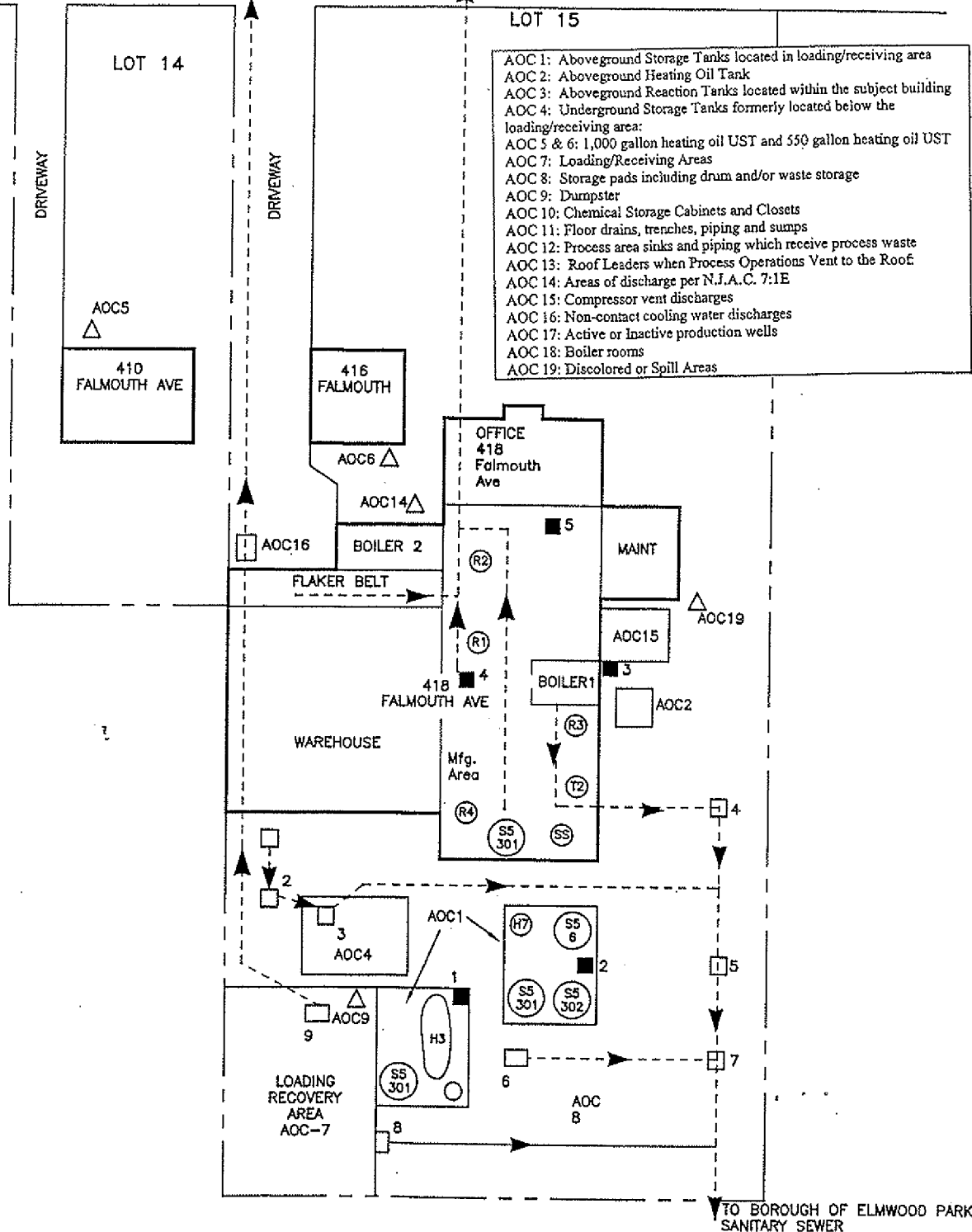
Brennan Environmental, Inc.

80 Great Meadow Lane, E. Hanover, NJ 07936 973.781.1801

TO STORM SEWER AND THEN TO
FLIESER'S BROOK

TO BOROUGH OF ELMWOOD PARK
SANITARY SEWER

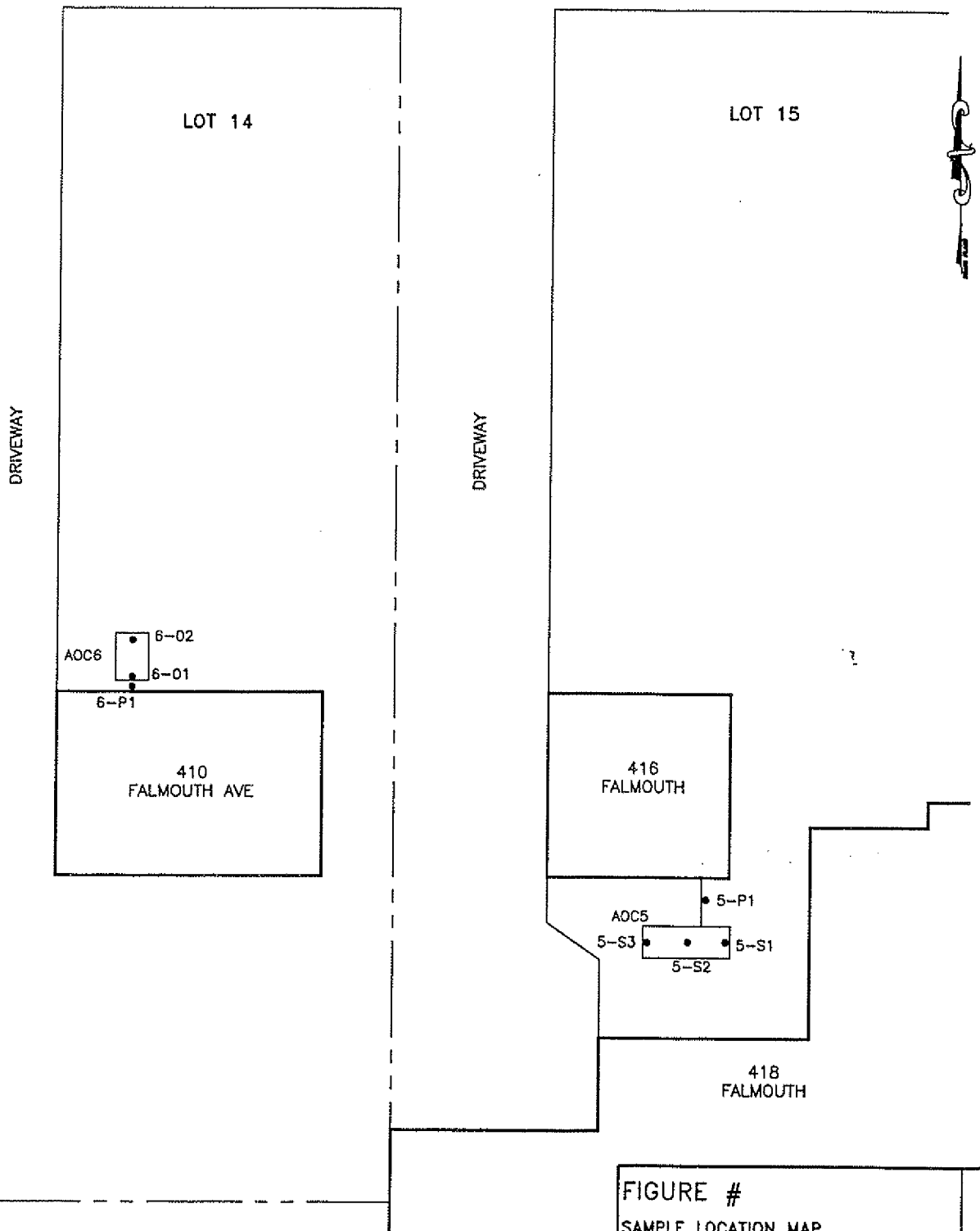
FALMOUTH AVENUE



LELIARTS LANE

FIGURE #		4
SITE PLAN		
FINETEX, INC. 418 FALMOUTH AVENUE ELMWOOD PARK, NEW JERSEY		
Scale: 1"=40'	Drawn By: AP	
Date: 11/12/2001	Project # 99704	
 BRENNAN ENVIRONMENTAL, INC. 8 Great Meadow Lane E. Hanover, NJ 07936 Tel: (973)781-1801		

FALMOUTH AVENUE



LEGEND:

● - SAMPLE LOCATION

FIGURE #

5

SAMPLE LOCATION MAP

FNETEX, INC.
418 FALMAOUTH AVENUE
ELMWOOD PARK, NEW JERSEY

Scale: 1"=20'

Drawn By: AP

Date: 11/12/2001

Project # 99704



BRENNAN ENVIRONMENTAL, INC.

8 Great Meadow Lane E. Hanover, NJ 07936
Tel: (973)781-1801



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDICES





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 1

Copy of Local Construction Permits and Inspection Approval Tickets





BUILDING SUBCODE TECHNICAL SECTION

A. IDENTIFICATION—APPLICANT: COMPLETE ALL APPLICABLE INFORMATION. WHEN CHANGING CONTRACTORS, NOTIFY THIS OFFICE. CALL UTILITY DIG NO: 1-800-272-1000.

Block

Work Site Location

Lot

Owner In Fee

Address

City

State

Zip

Contractor

Address

City

State

Zip

Telephone

Fax

License No. or Skills Reg. No.

Federal Emp. No.

JOB SUMMARY (Office Use Only)

PLAN REVIEW

☐ No Plans Required

☐ All

☐ Footing

☐ Foundation

☐ Frame

☐ Other

Joint Plan Review Required:

☐ Elec. ☐ Plumb. ☐ Fire ☐ Elevator

SUBCODE APPROVAL

☐ CO ☐ OCO ☐ CA

Date:

Approved by:

INSPECTIONS

Type:

Footing

Foundation

Slab

Frame

Barrier-Free

Insulation

Finishes

Energy

Mechanical

TCO

Other

Final

Dates (Month/Day)

Failure

Approval

Initial

B. BUILDING CHARACTERISTICS

Use Group Present Proposed

Constr. Class Present Proposed

No. of Stories

Height of Structure

Area — Largest Floor

New Bldg. Area/All Floors

Volume of New Structure

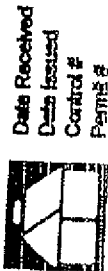
Total Land Area Disturbed

Est. Cost of Bldg. Work:

1. New Bldg. \$

2. Alteration \$

3. Total (1 + 2) \$



Date Received
Data Inspected
Control #
Permit #

C. CERTIFICATION IN LIEU OF OATH

I hereby certify that I am the (agent of) owner of record and am authorized to make this application.

Signature

D. TECHNICAL SITE DATA

DESCRIPTION OF WORK

Demolition of 1,000 Sq. Ft. of existing building.

TYPE OF WORK:

☐ New Building

☐ Addition

☐ Alteration

☐ Roofing

☐ Siding

☐ Fence

☐ Sign

☐ Pool

☐ Asbestos Abatement Subchapter 8

☐ Lead Haz. Abatement NJAC 5:17

☐ Other

☒ Demolition

FEE (Office Use Only)

\$

Administrative Surcharge

Minimum Fee

OCA Training Fee

TOTAL FEE

U.C.C. F110
(rev. 3/06)

1 White = Inspector Copy

2 Canary = Office Copy

3 Pink = Office Copy

4 Gold = Applicant Copy



CONSTRUCTION PERMIT

Date Issued 1-1-13
Control # 01-632
Permit # 803

IDENTIFICATION Block 803
Work Site Location 110 E. 11th St. New York, NY 10003
Owner in Fee 110 E. 11th St. LLC
Address 110 E. 11th St. New York, NY 10003
Tel. (212) 512-1111
Lic. No. or Bldgs. Reg. No. 10000000000000000000

Is hereby granted permission to perform the following work:

- ☒ BUILDING ☐ PLUMBING ☐ LEAD HAZARD ABATEMENT
- ☐ ELECTRICAL ☐ FIRE PROTECTION ☐ DEMOLITION
- ☐ ELEVATOR DEVICES ☐ ASBESTOS ABATEMENT ☐ OTHER

DESCRIPTION OF WORK:
Removal of lead paint from interior walls and ceiling of 110 E. 11th St. New York, NY 10003.

NOTE: If construction does not commence within one (1) year of date of issuance, or if construction ceases for a period of six (6) months, this permit is void.

Estimated Cost of Work \$ 10,000.00

Construction Official

U.C.C. F170 (rev. 5/28)

1 WHITE—INSPECTOR

2 CANARY—OFFICE

3 PINK—TAX ASSESSOR

4 GOLD—APPLICANT

(see reverse side)

PAYMENTS (Office Use Only)	
Building	
Electrical	
Plumbing	
Fire Protection	
Elevator Devices	
Other	
DCA Training Fee	
Cert. of Occupancy	
Other	
Total	
Check No.	
Cash	
Collected by	



BUILDING SUBCODE TECHNICAL SECTION

A. IDENTIFICATION—APPLICANT: COMPLETE ALL APPLICABLE INFORMATION. WHEN CHANGING CONTRACTORS, NOTIFY THIS OFFICE. CALL UTILITY DIG NO: 1-800-273-4000.

Block 1000 Lot 1000

Work Site Location 415 E. 1st St.

Owner in Fee 1000

Address 415 E. 1st St.

Telephone 202-337-1166

Contractor Joe R. Rouse

Address 213 E. 1st St.

Telephone 202-337-1166

Fax (202) 337-1166

License No. of Bldgs. Reg. No. 1000

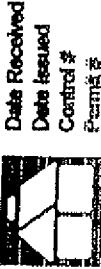
Federal Emp. No. 1000

JOB SUMMARY (Office Use Only)

PLAN REVIEW		INSPECTIONS		Dates (Month/Day)	
☐ No Plans Required	Date Initial	Type:	Failure	Failure	Initial
☒ All		Footings			
☐ Footings		Foundation			
☐ Foundation		Slab			
☐ Frame		Frame			
☐ Other		Barrier-Free			
Joint Plan Review Required:		Insulation			
☐ Elec.	☐ Plumb.	Finishes			
SUBCODE APPROVAL		Energy			
☐ CO	☐ COO	Mechanical			
Date:		TCO			
Approved by:		Other			
		Final			
		Barrier-Free			

B. BUILDING CHARACTERISTICS

Use Group	Present	Proposed	Est. Cost of Bldg. Work:
Const. Class	Present	Proposed	1. New Bldg. \$
No. of Stories			2. Alteration \$
Height of Structure			3. Total (1+2) \$
Area — Largest Floor			
New Bldg. Area/All Floors			
Volume of New Structure			
Total Land Area Disturbed			



G. CERTIFICATION IN LIEU OF OATH

I hereby certify that I am the (agent of) owner of record and am authorized to make this application.

Signature

D. TECHNICAL SITE DATA

DESCRIPTION OF WORK

1000 East 1st St.
1000 East 1st St.

TYPE OF WORK

☐ New Building	☐ Roofing	☐ Fence	☐ Sign	☐ Pool	☐ Asbestos Abatement Subchapter 8	☐ Lead Haz. Abatement NJAC 5:17	☐ Other	☒ Demolition
☐ Addition	☐ Siding	☐ Height (exceeds 6')	☐ Sq. Ft.					
☐ Alteration								

FEE (Office Use Only)

\$	
\$	
\$	
\$	
\$	
\$	
\$	
\$	
\$	
\$	

Administrative Surcharge	\$
Minimum Fee	\$
DCA Training Fee	\$
TOTAL FEE	\$

U.C.C. F110
(rev. 3/00)

1 White = Inspector Copy
2 Pink = Office Copy
3 Gold = Applicant Copy
4 Gold = Applicant Copy

CONSTRUCTION PERMIT

Date Issued
Control #
Permit #

IDENTIFICATION Block Lot
Work Site Location 15
Owner in Fee 300 RIVER DRIVE
Address 4500 300 RIVER DRIVE
Tel. 973 472 1100 Lic. No. or Bids Reg. No. 23 1992140

I hereby granted permission to perform the following work:

- ☒ BUILDING ☐ PLUMBING ☐ LEAD HAZARD ABATEMENT
 - ☐ ELECTRICAL ☐ FIRE PROTECTION ☐ DEMOLITION
 - ☐ ELEVATOR DEVICES ☐ ASBESTOS ABATEMENT ☐ OTHER
- (Subchapter 8 only)

DESCRIPTION OF WORK:

Water Tank Removal

NOTE: If construction does not commence within one (1) year of date of issuance, or if construction ceases for a period of six (6) months, this permit is void.

Estimated Cost of Work \$

Construction Official

U.C.C. F170
(rev 5/2K)

1 WHITE—INSPECTOR

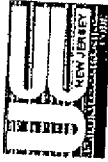
2 CANARY—OFFICE

3 PINK—TAX ASSESSOR

4 GOLD—APPLICANT

(see reverse side)

PAYMENTS (Office Use Only)	
Building	<u>25</u>
Electrical	
Plumbing	
Fire Protection	
Elevator Devices	
Other	
DCA Training Fee	
Cert. of Occupancy	
Other	
Total	<u>25</u>
Check No.	<u>2588</u>
Cash	
Collected by	



For Information Call:

Permit No. 00668

APPROVAL FOR BUILDING

	Date	Inspector
☐ Footing		
☐ Foundation		
☐ Frame		
☐ Insulation		
☐ Mechanical		
☐ Other <u>Final</u>	<u>11/2/00</u>	<u>[Signature]</u>
☐ Other		
☐ Final		

U.C.C. F221
(rev. 3/95)



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 2

Tank Contents Waste Disposal Documentation



(973) 696-3122

5612

PA/AH/0564/ CT-HW-658

DEP # - S50015/JA-389

EPA # - NJD 986588630



All State O.R.C. Inc.
473 Hamburg Tpke.
West Milford, N.J. 0748

CUSTOMER'S ORDER NO.		PHONE		DATE	
				10-11-01	
NAME JOE ROYCE					
ADDRESS 410 FALMOUTH AVE.					
ELMWOOD PARK N.J.					
YARD TIME OUT			YARD TIME IN		
IN	OUT	DESCRIPTION	CHECK #	AMOUNT DUE	
		GENERATOR #			
		MANIFEST #			
		DRIVER NAME RON			
		#103			
		#104 VAC TR.			
		#105 RACK TRUCK			
		#107			
10:20		#108			
		#110 VACTOR			
		#111 CARRIER			
		#112 CARRIER			
		WASTE DESCRIPTION #2 oil Ballast - H₂O			
		GALLONS # 174			
		VACUUM out 17" in Ground Tank			
		2" water			
RECEIVED BY [Signature]			TAX		
			TOTAL		

W. SAUNDERS PRINTING - 232-1304
CEDAR GROVE, NJ 07015-0101
CLC 4076-S

SM 1100

Thank You

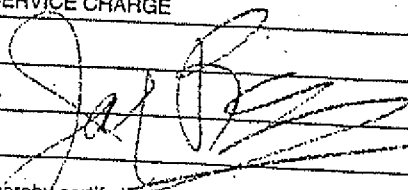
(973) 696-3122

PA/AH/0564/ CT-HW-658
DEP # - S50015/JA-389
EPA # - NJD 986588630



3176

All State O.R.C. Inc
473 Hamburg Tpke
West Milford, N.J. 0

CUSTOMER'S ORDER NO.		PHONE		DATE		
NAME		JOE ROYCE		10-2-00		
ADDRESS		416 FAHMOUD AVE		ELMWOOD PARK, N.J.		
SOLD BY	CASH	C.O.D.	CHARGE	ON ACCT.	MDSE. RET'D	PAID OUT
QTY.	DESCRIPTION				PRICE	AMOUNT
	GENERATOR # Vacuum out Bottoms of					
	MANIFEST # 1000 gal TANK					
	X721	196 H2O				
	X722	11:00 - 11:30				
	X723					
	X724	Ron & Ricky				
	X725					
	X726 L.H.					
	FUEL					
	SERVICE CHARGE					
						
I hereby certify that my Hazardous waste streams total less than 220 pounds (100 kg) for this calendar month and that I am not required to obtain an EPA Identification number						
RECEIVED BY					TAX	
					TOTAL	

W. SAUNDERS PRINTING - 223-1306
CEDAR GROVE, NJ 07009-5000
CLC 4070-S

All claims and returned goods
MUST be accompanied by this bill.

Thank You

COPY



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 3

Clean Fill Material Documentation





BRAEN STONE
INDUSTRIES, INC.

P.O. Box 8310 •
HALEDON, NJ 07538
973.595.6250 •
FAX: 973.595.7087
www.braenstone.com

DATE: 10/11/01
TIME: 09:22
PLANT: COMP:
SCALE: 1

P.O. BOX 8310 HALEDON N.J.
973-595-6250 FAX#973-595-7087

TICKET #: 25088
PD #:
FOREMAN #:

CUST: 000091
JOE ROYCE, INC.
JOB: 91-1

TRUCK ID: ROYCE27
LICENSES:
CARRIER: JOE ROYCE INC
LOCATION: JOE ROYCE

PROD: 329
QUARRY PROCESS

WE MAKE DELIVERIES INSIDE CURB LINE AND ON THE LOT
AT CUSTOMER'S RISK ONLY AND ACCEPT NO RESPONSIBILITY
WHATSOEVER FOR DAMAGE RESULTING FROM SUCH
DELIVERIES.

09:22	GROSS: 23980 Lb	11.99 Tn	10.875 T
09:18	TARE: 14960 Lb	7.48 Tn	06.785 T
	NET: 9020 Lb	4.51 Tn	04.091 T

JOB TOTALS	LOADS	POUNDS	TONS	MB	
9999	0001	9020	4.51	4.090	PRICE/TDN
	0000				SUB-TOTAL
	0000				DELV RATE
	0000				STATE TAX
	0000				DELV CHARGE
	0000				TOTAL AMT

RECEIVED BY:
INSPECTED BY:

TEMPERATURE:
WEIGHMASTER: U.S.A.#1

DELIVERY TICKET
TRUCKER



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 4

UST Disposal Documentation



invoice

TO

Royce

Invoice Date

Shipped To

Your Order No.

Date Shipped

Shipped Via

F.O.B.

Terms

Salesman

QUANTITY	DESCRIPTION	PRICE	AMOUNT
	1-550 GALLON CUT & CLEANED		
	NO VALUE		

Reorder from Business Envelope Manufacturers • Pearl River, N.Y. 10965 • Item No. N

invoice

TO JOE ROYCE

Invoice Date 10.2.00

Shipped To

Salesman:

Reorder from Business Envelope Manufacturers, Pearl River, N.Y. 10963 • Item #

COPY



BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 5

Photographic Log



Photolog

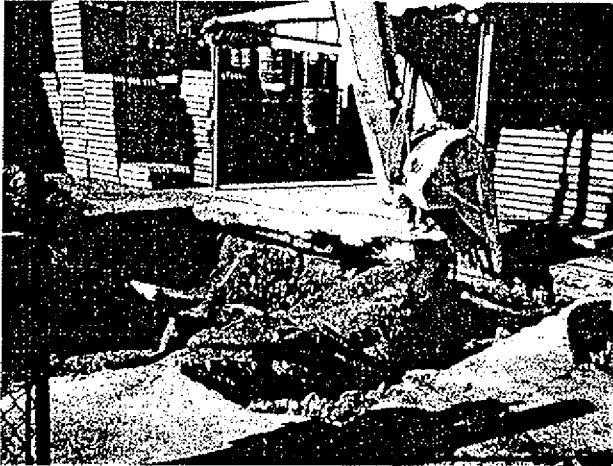


Photo: 9 AOC 19 Discolored concrete



Photo: 9 AOC 19 Discolored concrete

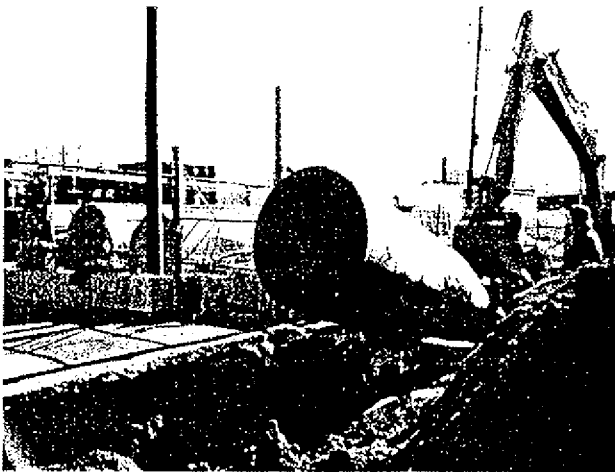


Photo: 9 AOC 19 Discolored concrete

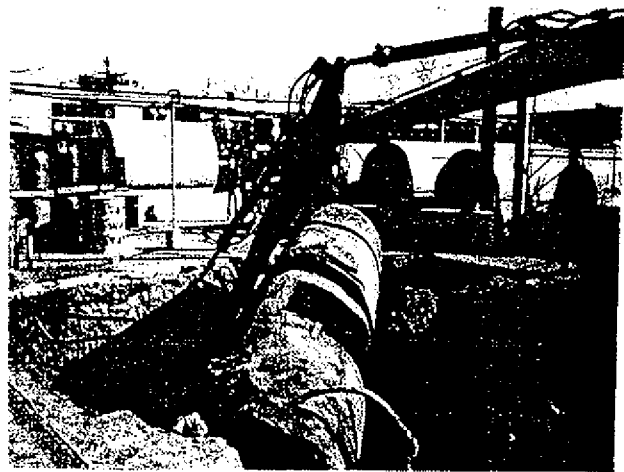


Photo: 9 AOC 19 Discolored concrete



Photolog

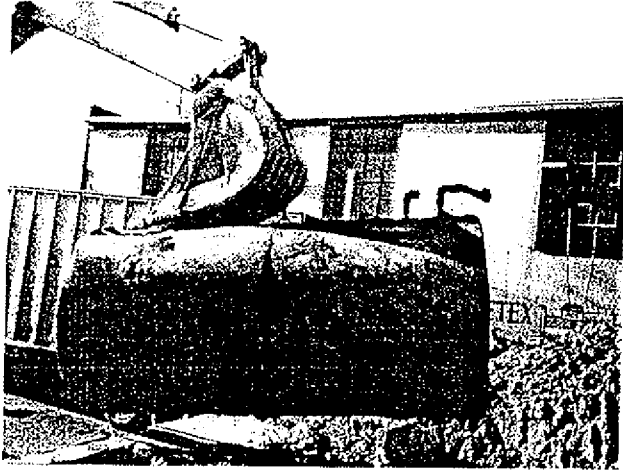


Photo: 9 AOC 19 Discolored concrete

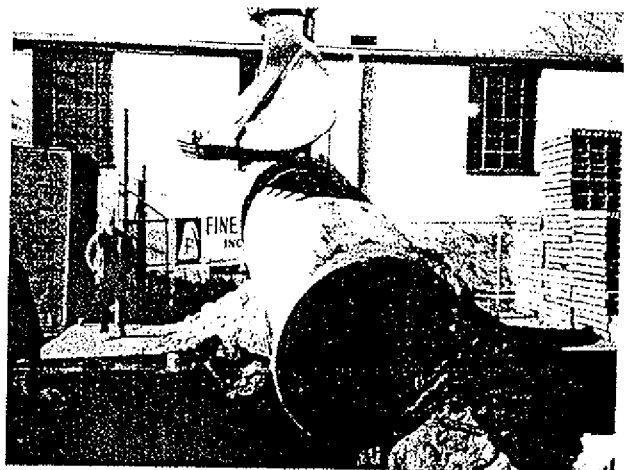


Photo: 9 AOC 19 Discolored concrete

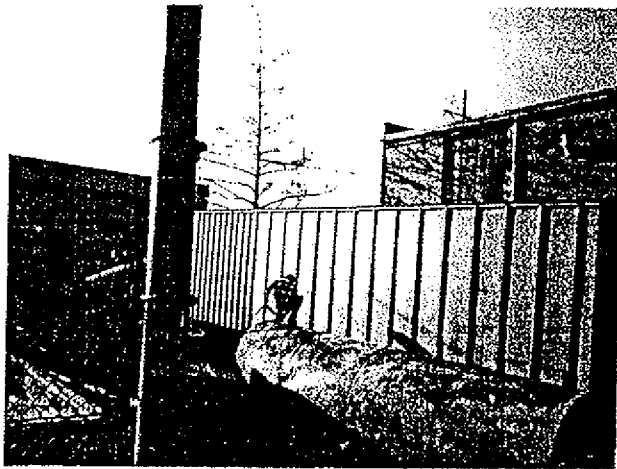


Photo: 9 AOC 19 Discolored concrete



Photo: 9 AOC 19 Discolored concrete





BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 6

BEI's Standard QA/QC Plan



TABLE OF CONTENTS

THE SAMPLING PLAN.....	2
A. <u>SITE HISTORY-EVALUATING EXISTING DATA/FILE INFORMATION</u>	2
B. <u>DEFINING THE PHYSICAL ENVIRONMENT</u>	4
C. <u>SAMPLE LOCATIONS AND NUMBERS</u>	5
D. <u>SAMPLE METHODOLOGY AND MATRIX</u>	6
E. <u>LABORATORY SELECTION</u>	6
F. <u>QUALITY ASSURANCE CONSIDERATIONS</u>	7
G. <u>HEALTH AND SAFETY CONCERNS</u>	7
H. <u>SCHEDULE</u>	8
QUALITY ASSURANCE	8
A. <u>INTRODUCTION</u>	8
B. <u>SAMPLE CONTAINERS</u>	9
1. <i>Reactivity of Container Material with Sample</i>	9
2. <i>Volume of the Container</i>	10
3. <i>Color of Container</i>	10
4. <i>Container Closures</i>	10
5. <i>Decontamination of Sample Containers and Chain of Custody</i>	10
6. <i>Sample Bottle Storage and Transport</i>	11
C. <u>DECONTAMINATION OF SAMPLING EQUIPMENT</u>	11
1. <i>Laboratory Decontamination</i>	11
2. <i>Field Decontamination</i>	12
3. <i>Decontamination of Pumps</i>	14
4. <i>Decontamination of Heavy Equipment</i>	14
5. <i>Decontamination of Backhoes/Drill Rigs</i>	15
6. <i>Decontamination of Monitor Well Casing and Screen</i>	15
7. <i>Cleaning Location</i>	15
8. <i>Disposal of Drill Cuttings</i>	16
9. <i>Disposal of Development, Purge, Pump Test and Decon Waters</i>	17
D. <u>PROCEDURES FOR QUALITY ASSURANCE AND QUALITY CONTROL (QA/QC)</u>	17
1. <i>Non-Aqueous Matrix</i>	18
a. <i>Field Blanks</i>	19
b. <i>Trip Blanks</i>	19
2. <i>Aqueous Matrix</i>	19
a. <i>Field Blanks</i>	19
b. <i>Trip Blanks</i>	20
3. <i>Air Matrix</i>	20
4. <i>Blank Water Quality</i>	20
5. <i>Sample Handling and Holding Times</i>	21
a. <i>Handling Time</i>	21
b. <i>Maximum Holding Time</i>	21
6. <i>Special Analytical Services (SAS)</i>	22
7. <i>Additional QA/QC Samples</i>	22
a. <i>Duplicate Samples Obtained in the Field</i>	22
b. <i>Splitting Samples with Responsible Parties</i>	24
c. <i>Performance Evaluation Samples (PE)</i>	24
d. <i>Matrix Spike/Matrix Spike Duplicate Analyses</i>	25
e. <i>Background Samples</i>	25
f. <i>Dioxin and Furans (PCDDs/PCDFs)</i>	26
E. <u>SAMPLE PRESERVATION REQUIREMENTS</u>	26
TABLE 2-1 <i>AQUEOUS SAMPLING EQUIPMENT DECONTAMINATION</i>	29
TABLE 2-2 <i>NON-AQUEOUS SAMPLING EQUIPMENT DECONTAMINATION</i>	30

THE SAMPLING PLAN

There are a wide variety of reasons for collecting samples and various sampling strategies for different situations. It is important that the purpose of the sampling and associated data quality objectives be identified before field work begins. For example, samples may be collected to determine the existence and/or to define the extent of contamination at a site, to allow waste characterization and classification for disposal or recovery, or to determine compliance with existing regulations. Once the objective is known, decisions about analytical parameter selection, NJ certified laboratory selection, quality control samples, sample location and frequency, etc., can be made more confidently. Defining these sampling and data quality objectives is important to assure that the sampling plan is complete. Environmental sampling is often conducted to gather data that will be the basis for remedial decisions. Because of the potential threat to health and environment and high costs usually associated with site remediation, strict adherence to quality assurance measures are strongly recommended. In such a case, the object of the sampling helps to dictate what should be prescribed in the sampling plan.

An integral part of any sampling program is planning. Before a plan can be written site-specific information must be gathered to insure that the plan is logical, will meet the required objectives, and the course of action laid out is achievable.

The purpose of developing a sampling plan is to detail a "plan of action". The person writing the plan must be very familiar with the site specific conditions and those implementing the plan must be very familiar with the plan's contents. A properly prepared sampling plan that is correctly implemented will allow the sampling objective to be met, help avoid confusion in the field, preserve health and safety, and ultimately save time and money. In the development of the sampling plan other pre-sampling activities must be heavily relied upon. Some factors to be taken into account include the following.

A. Site History-Evaluating Existing Data/File Information

The first step in a site investigation should be the gathering of background information. Information concerning the history of activity at a site (including locations and age of buildings, drainage pathways, contours, building layout, foundations, septic systems, tanks, etc.; processes and materials for manufacture, storage and disposal both past and present) can be extremely useful in planning sampling events. A file search may reveal areas of a site used for specific processes (aerial site history, site plans, area land use may also be useful) and will help in the logical placement of sampling locations. By revealing what materials were handled on site, a file search may provide guidance in choosing which parameters to include for analysis. Additionally, while caution must still be used, judgments regarding health and safety requirements can be made. When no information is available, field personnel must consider that worst case conditions may exist and take proper precautions to insure safety.

The following is a list of federal, state, local and other agencies or sources where additional information regarding site history may be obtained. Addresses and phone numbers can be found in the Blue Pages of Governmental Listings provided in New Jersey Bell's yellow pages.

U.S. Government

U.S. Department of Justice
U.S. Geological Survey
U.S. DOA - Soil Conservation Service
U.S. DOA - Forest Service
U.S. DOI - Fish and Wildlife Agencies
U.S. Army Corps of Engineers
Federal Emergency Management Agency
National Oceanic and Atmospheric Administration
U.S. Environmental Protection Agency

State of New Jersey

NJ State Library
NJ State Attorney General Office
NJ Geological Survey
NJ Department of Transportation
NJ Department of Agriculture
NJ Department of Health
NJ Department of Environmental Protection:
 Regional Enforcement (Northern, Central, Metro, and Southern)
 Bureau of Federal Case Management
 Bureau of State Case Management
 Bureau of Community Relations
 Bureau of Site Management
 Bureau of Air Pollution Control
 Bureau of Emergency Response
 Bureau of Environmental Evaluation and Responsibility Assessment (ECRA)
 Bureau of Environmental Evaluation and Risk Assessment
 Bureau of Site Assessment
 Bureau of Environmental Measurements and Quality Assurance
 Information Resource Center
 Office of Environmental Analysis (Aerial Maps)
 Radiation Protection Programs
 Pesticide Control Program

Local Government

County Health Department
Local Health Department
Tax Assessors Office
Economic Development Officer
Local Planning Board
Town Engineer
Local Chamber of Commerce
Local Airport
Local Library
Local Well Drillers

Other Sources

Facility Records
Employee Records
Citizens residing nearby
Local and regional waste haulers and generators

B. Defining The Physical Environment

Equal in importance to finding out what may be on site is determining where it is most likely to be located. A pre-sampling site visit should be conducted to gather additional background information. Labels and DOT numbers on drums and tanks may be useful. Files found on-site may include information about materials that were manufactured, stored or disposed of on-site. Product names may be determined from shipping labels or manifests. Any and all information will be useful in sampling plan preparation, and in formulating a site-specific Health and Safety Plan (see Chapters 6, 15 and 16 of this Manual).

The fate of environmental contaminants is dictated by the source, the characteristics of the contaminant itself, (i.e., persistency and toxicity) and perhaps most importantly, by the physical environmental system into which it is released. Contaminants move at varying rates and to varying degrees when released into different kinds of matrices. Defining what kind of environmental system the site is a part of is extremely important to the success of achieving the sampling objectives. An investigation into the local geology, hydrology (including flow rates of nearby surface waters, average depth to ground water and flow direction, identification of areas of recharge, etc.), and climatology is necessary. The biological system should also be assessed. The flora and fauna of the area (including identification of sensitive environments and/or species, stressed vegetation, potential for bioaccumulation and biotransformation in the plant and animal life, especially agricultural) are definite factors to be taken into account. Stressed vegetation may serve as an indicator for contaminant migration to a particular area. Overall, by defining the physical environment, the fate of contaminants can be predicted. Migration pathways should also be identified assuring that samples will be collected in the most appropriate area.

The factors addressed above offer an overview of considerations that must be evaluated for a sampling plan to be complete. The more information that is obtained, the more that will be known about the source, movement, and concentrations of contaminants in the media to be sampled. With this knowledge, it will be easier to write a complete, site specific sampling plan.

Along with the historical and physical information needed prior to sampling plan development, the following topical areas of basic information are necessary components for an inclusive sampling plan.

C. Sample Locations And Numbers

The objective of the sampling event is important when choosing the location of sampling points. Samples are sometimes collected to characterize a site for which limited background information is available and/or obvious contaminated areas do not exist. In such a case, a random sampling scheme may be useful. Random sampling depends on the theory of random chance probabilities to choose the most representative sample.

This process is utilized when there are numerous available sampling locations and there are no satisfactory reasons for choosing one location over another.

Tables of random numbers are readily available from many sources and should be used to eliminate any possible bias generated by those collecting the sample, assuming a random approach is used.

Also important when choosing sample locations is consideration of the site's physical environmental setting and how these factors can influence the concentration and movement of the material of concern. Sampling at hazardous waste sites is usually conducted in an attempt to discover contamination and to define its extent and variability. With such an objective, it is most logical to choose sample locations that will yield the most information about site conditions. Here, judgment (or biased) sampling should be employed. Biased samples are those collected at locations that were chosen based on historical information, knowledge about the location and behavior of the contaminant(s), and/or knowledge about the effects of the physical system on the contaminants' fate.

Both biased and random sampling techniques can be used together to thoroughly address an entire site. Some samples may be biased to potentially contaminated areas (e.g., stained soil, former process or disposal areas) or potentially impacted areas (e.g., areas of stressed vegetation, sediment downstream from discharge pipe). In areas less likely to be contaminated or areas with little available background information, random samples may be used to allow adequate assessment of the entire site.

There are six factors in determining the number of samples required for site characterization and these are:

1. exposure pathways
2. statistical performance objectives
3. quality assurance objectives
4. background samples
5. sampling objectives
6. site specific conditions.

For example, if the objective of the event is to determine whether the site is contaminated, a limited number of samples, from properly chosen locations, will yield useful information. If, however, the site is known to be contaminated, and delineation of the contamination is the objective, a greater number of samples may be needed. In many cases statistical considerations can be helpful in determining sampling strategy (see Chapter 3. "Statistical Considerations").

D. Sample Methodology and Matrix

Once the appropriate numbers and locations have been chosen, consideration must be given to what collection method will be used to assure that representative samples of site conditions are obtained. The selected sampling methodology will be matrix dependent. In some instances, there may be several acceptable options available for collecting a sample. In other instances, site specific conditions may dictate that only one approach will work, even though that method may not be the preferred method. In all cases, the construction material of the sampling device, its design, decontamination, and proper use are critical factors and should be included in the proposed sampling plan.

Use of a device constructed of undesirable material may compromised sample quality by the material leaching into the sample or absorbing materials from the sample after repeated use and decontamination. Sampler design is also important. For example, a ground water sampling device that aerates the sample during collection may yield a sample that is not representative of actual aquifer conditions. Finally, even the most well designed, constructed and cleaned sampling device will yield a non-representative sample if used improperly. All personnel involved in sample collection must receive training on the use, care and limitations of different sampling equipment.

Further, decontamination of the chose device must be considered. The sampling device must be resistant to the decontamination solutions and should be constructed to allow ease of cleaning and assure thorough decontamination. (See Chapter 2. "Quality Assurance", for decontamination procedures).

E. Laboratory Selection

For regulatory samples collected under the Safe Drinking Water Act, the National Pollution Discharge Elimination System and the Radon Act, the laboratory must have current certification for that contaminant or parameter covered by the "Regulations Governing

Laboratory Certification and Standards of Performance", N.J.A.C. 7:18. When additional certifications for other contaminants and parameters and other matrices are offered under amendments to N.J.A.C. 7:18, the laboratory must have current certified status before performing these analysis.

In addition, the following laboratory capabilities may be applicable depending upon the specific regulatory program that sample analysis is being conducted:

1. For the analysis of aqueous samples, the laboratory shall be a New Jersey certified Laboratory for all parameters of appropriate category of parameters for which analysis will be performed and for which certification exists pursuant to N.J.A.C. 7:18 et seq.;
2. For the analysis of aqueous and non-aqueous samples for applicable organics and inorganics including polychlorinated dibenzo-p-dioxins and polychlorinated dibenzofurans, the laboratory should be a participant in good standing in the EPA Contract Laboratory Program (CLP) for the applicable Statement of Work protocol as described in the version of the EPA CLP "Invitation for Bid" (IFB) contract documents in effect as of the date on which the laboratory is performing the analysis;
3. For the analysis of aqueous and non-aqueous samples for applicable organic and inorganic, the laboratory should be a contract awardee able to perform sample analyses in the applicable analytical task under the version of the Professional Laboratory Analytical Service contract issued by the New Jersey Department of Treasury, Division of Purchase and Property in effect as of the date on which the laboratory is performing the analysis.
4. For the analysis of non-aqueous samples for selected organics and inorganics using methods contained in the EPA Publication SW-846 "Test Methods for Evaluating Solid Waste", in effect as of the date on which the laboratory is performing analysis, the laboratory shall be a N.J.A.C. 7:18 certified laboratory for those parameters.

F. Quality Assurance Considerations

Quality assurance measures must be associated with each sampling and analysis event as an additional measure of control to assure that the sample delivered to the lab for analysis is representative of site conditions. The sampling plan should outline how the representative quality of the samples will be assured. This will include, but not be limited to: laboratory SOP, field SOP, sample bottle preparation, equipment decontamination, trip blanks, field blanks, duplicates, split samples, performance evaluation samples, sample preservation and handling, chain of custody, analysis request, analytical methods, parameters, and deliverables (See Chapter 2. for further quality assurance information).

G. Health And Safety Concerns

Health and safety measures must be incorporated in the sampling plan and may address the following areas: Organizational responsibilities, risk analysis, employee training, personnel protection, medical surveillance, air surveillance, site control, decontamination, site standard operating procedures, contingency planning, confined space operations, and spill

containment. Some of these concern areas may not be applicable for your sampling episode. (See Chapter 15 for more information on Personnel Protection).

H. Schedule

Scheduling information may or may not be applicable for your particular sampling plan development. If it is necessary, the type of information to include would be time frames for various milestone sampling episodes.

QUALITY ASSURANCE

A. Introduction

This section of the manual provides the user with quality assurance requirements and procedures for conducting environmental measurement sampling episodes. In order to generate analytical data of known and defensible quality, adherence to established quality assurance protocol is necessary. Quality assurance measures coupled with a statistically based sampling plan will improve sample collection while maintaining the integrity of the samples prior to analysis. NJDEP has established standard operating procedures to maintain consistency in sample collection and handling. The standard operating procedures may vary among the specific regulatory programs (i.e. CERCLA, RCRA, NJPDES).

To help ensure that environmental monitoring data is of known quality, monitoring projects for the NJPDES and Safe Drinking Water Act Programs are based on an approved Quality Assurance Project Plan (QAPjP) prepared by using a standard reference, "Guidance for Preparation of Combined Work/Quality Assurance Project Plans for Environmental Monitoring" (OWRS QA-1). The above referenced document was developed in conformance with QASMS-005/80, "Interim Guidelines and Specifications for Preparing Quality Assurance Project Plans". All QAPjPs must conform with these guidelines. A QAPjP is a single document incorporating QA/QC controls which encompass both field and laboratory issues associated with the information of NJPDES permit requirements.

Another example of the application of these protocols is the Field Sampling Plan-Quality Assurance Project Plan (FSP-QAPP). To develop reliable site investigation data for NJDEP lead, publicly funded CERCLA (Superfund) sites, the prime consultant/contractor awarded a term contract for the Remedial Investigation/Feasibility Study (RI/FS) or Engineering Design work has the responsibility to develop and implement a FSP-QAPP. This document must present the organization, functional activities and specific QA/QC activities needed to attain specific project goals and data quality objectives.

The core components of a FSP-QAPP are the Field Sampling Plan and the Quality Assurance Plan. Other essential elements included in the document are the project description, project organization/responsibilities, and schedules. Regardless of a document title or "deliverable"

name (e.g. QAPjP, FSP-QAPP), it is recommended that all sampling and quality assurance protocols associated with a site specific investigation/remediation be contained in a stand alone document.

Quality Assurance Programs

The procedures established to control the collection and handling of samples are an integral part of the Quality Assurance Program operating within NJDEP. The importance of the environmental sample collection process and associated analytical data is demonstrated through integration of this information into the decision making process. All phases of this process rely on the provision of accurate, precise, comparable and complete analytical data.

Sample collection, preservation and handling methods that are required by the code of Federal Regulation must be used for samples covered by the Safe Drinking Water Act and Water Pollution Control Act. Those requirements are published annually in 40 CFR Parts 141 and 136, respectively.

The following quality assurance requirements have been established to maintain sample integrity. Their prime objectives are to maintain the physical form and chemical composition of the sample and to prevent contamination from other sources or changes in contaminant concentration. To meet these objectives, there must be a measure of control over all sample handling procedures beginning with sample container cleaning procedures and ending with laboratory analysis. This chapter focuses on the first half of the control process: the procedures leading up to and ending with sample packaging and transport to the laboratory.

B. Sample Containers

Prior to the collection of a sample, consideration must be given to the type of container that will be used to store and transport the sample. The party requesting the analysis is responsible for requesting the proper sample containers. Their selection is based on the sample matrix, potential contaminants to be encountered, analytical methods requested, and the laboratory's internal quality assurance requirements. Selection of appropriate sample containers should also be based upon review of the criteria listed below, as well as the information provided in Appendix 2-1.

1. Reactivity of Container Material with Sample

Choosing the proper composition of sample containers will help to ensure that the chemical and physical integrity of the sample is maintained. For sampling potentially hazardous material, glass is the recommended container type because it is chemically inert to most substances. Plastic containers are not recommended for most hazardous wastes because the potential exists for contaminants to adsorb to the surface of the plastic or for the plasticizers to leach into the sample.

In some instances, the sample characteristics or analytes of interest may dictate that plastic containers be used instead of glass. Because some metals species will adhere to the sides of glass. Because some metals species will adhere to the sides of glass containers in an aqueous matrix, plastic bottles must be used for samples collected for metals analysis. In the case of a strong alkali waste or hydrofluoric solution, plastic containers may be more suitable because glass containers may be etched by these compounds creating adsorptive site on the container surface.

2. Volume of the Container

The volume of sample to be collected will be dictated by the analytical method and the sample matrix. The laboratory must supply bottles of sufficient volume to perform the required analysis. Appendix 2-1 of this chapter indicates the container volumes required for the various parameters. In most cases, the methodology dictates the volume of sample material required to complete the analysis. However, individual labs may provide larger volume containers for various analytes to ensure sufficient quantities for replicates or other quality control checks.

3. Color of Container

Whenever possible, amber glass containers should be used to prevent photodegradation of the sample, except when samples are being collected for metals analysis. If amber containers are not available, containers holding samples should be protected from light. However, 40ml clear glass vials are often provided by laboratories for VOA/aqueous analysis and are acceptable for use.

4. Container Closures

Container closures should form a leak-proof seal (i.e., screw caps or ground glass stoppers). Closures must be constructed of a material which is inert with respect to the sampled material, such as PTFE (e.g., Teflon[®]). Alternately, the closure may be separated from the sample by a closure liner that is inert to the sample material such as PTFE liner or septum.

5. Decontamination of Sample Containers and Chain of Custody

Sample containers must be laboratory cleaned, preferably by the laboratory performing the analysis. The cleaning procedure is dictated by the specific analysis to be performed on the sample (see Appendix 2-1 of this Chapter). The sample bottles should be prepared for shipment accompanied by a chain of custody and the cooler or shuttle containing them should be custody sealed. The chain of custody must also accompany the bottles during transportation to the field, sample collection, transportation back to the lab, during analysis

and identify final disposal of the sample container. When collecting a sample, sampling personnel should record the seal number associated with each sample shuttle or cooler and record whether the seal was intact upon arrival in the field. This assures that the sample containers were not tampered with in the time between their preparation and their arrival in the field. After sample collection, the bottles again should be sealed into the shuttle or cooler and the seal number should be recorded in the field log book. Upon arrival at the lab, the person receiving the sample should note the number and condition of the custody seal. Refer to Chapter 13, Documentation, for additional information on Chain of Custody.

6. Sample Bottle Storage and Transport

No matter where the sample bottles are, whether at the lab waiting to be packed for shipment or in the field waiting to be packed for shipment or in the field waiting to be filled with sample, care must be taken to avoid contamination. Sample shuttles, and clean sampling equipment must never be stored near solvents, gasoline, or other equipment that is a potential source of contamination. When under chain of custody, sample bottles must be secured in locked vehicles, custody sealed in shuttles or in the presence of authorized personnel.

C. Decontamination of Sampling Equipment

An important aspect of quality control is the decontamination of field sampling equipment. Improperly cleaned and prepared sampling equipment can lead to misinterpretation of environmental data due to interference caused by cross-contamination.

The following sampling equipment cleaning procedures (ASTM D-5088-90, "Decontamination of Equipment") are standard requirements of the NJDEP. Also included are procedures for cleaning heavy equipment and disposal of decontamination fluids and drill cuttings. Exception to the following procedures may be evaluated and approved by NJDEP on a case-by-case basis if justifications to do so involving site specific issues or conditions are presented and verified beforehand.

1. Laboratory Decontamination

In certain instances laboratory decontamination can serve as a viable alternative to field decontamination when sampling a non-aqueous matrix. Some advantages include: 1.) decontamination takes place in a controlled environment; 2.) reduced need to transport, handle or dispose cleaning solvents, acids or wash water; 3.) more attention can be focused upon sampling with field decontamination labor reduced or eliminated; 4.) reduced probability of cross-contamination due to improperly field decontaminated equipment; and 5.) laboratory documentation of cleaning procedures and materials used. Disadvantages may include: 1.) relative cost to scope of sampling event; 2.) constraints meeting demands in emergency situations; and 3.) logistics.

While the option to sue, or not use, laboratory decontaminated field sampling equipment for non-aqueous sampling and certain aqueous sampling equipment exists (i.e., foot check valves, filtering equipment, stainless steel clamps, automatic wastewater composite samplers), there is no option available when sampling ground water with bailers. GROUND WATER SAMPLING DEVICES are required to be laboratory cleaned, packaged and dedicated for exclusive use at one sample location for that day's sampling (see definition of "laboratory cleaned" in the glossary). Field decontamination of bailers elevates the potential of cross-contamination to unacceptable levels. The possibility of contaminating a clean well is also of concern when using improperly cleaned sampling devices.

It is recommended that extra sampling devices be available on-site in the event problems occur which prohibit the use of a particular instrument. When samples are returned to the laboratory, the bailers are also returned so they can be cleaned and prepared for the next sampling event. It is preferred that the lab performing the analysis be the same lab that prepared the sampling equipment. This cannot always be arranged; therefore, make certain the laboratory analyzing the samples will accept samples collected with field equipment not prepared at their laboratory. Also, the cleaning procedure of the laboratory preparing the sampling equipment must be in accordance with NJDEP requirements. The procedure for laboratory decontamination of dedicated field sampling equipment can be found in Table 2-1.

After this procedure has been completed, the sampling device should be wrapped in cleaned and oven baked aluminum foil (see Chapter 2 footnote #4) or equivalent material and custody sealed for identification. A record should be kept of the date and time and this information should be labeled on the sampling device.

Sampling equipment should remain in the wrapped material until it is used in the field. It should be handled as little as possible prior to use and disposable gloves must be worn at all times when handling cleaned sampling equipment. Sampling equipment must never be stored near solvents, gasoline, exhaust emissions, or other equipment and/or materials that may impact the integrity of prepared sampling instruments.

2. Field Decontamination

In the event that laboratory cleaning is not an option or not feasible, a field cleaning procedure must be used in order to reduce the chances of cross-contamination between sample locations. Several criteria are used to judge whether field decontamination procedures are appropriate. These are:

- Large number of sample locations may be scheduled in a one-day sampling event.
- Matrix of sample: Dedicated sampling equipment is always the preferred option for all matrices, however, it may not be practical for soil sampling equipment such as split spoons and hand augers. (Ground water sampling equipment must be laboratory cleaned and dedicated to a sampling point per day of sampling.)

- Episode logistics: It may be impractical to lab clean and dedicate sampling equipment in a short-term emergency situation because of time and administrative constraints.

Selection of a location for a field equipment decontamination station is also important. It should be located away from any on-site source of contamination so as not to adversely impact the decontamination procedure and close enough to the sampling teams to keep equipment handling to a minimum. The decontamination station must also be set up in such a way as to not adversely impact a clean environment.

Utilization of several units of one type of sampler will allow decontamination teams to rotate equipment more effectively. Tables 2-1 and 2-2 detail the procedures for field decontamination of aqueous and non-aqueous sampling equipment respectively.

The decontamination procedures (Tables 2-1 and 2-2) developed by the NJDEP, have the concurrence of USEPA, Region II Monitoring Management Branch and follow ASTM D-5088-90 methods. The first step, a detergent and water wash, is to remove all visible particulate matter and residual oils and grease. (This may be preceded by a steam or high pressure water wash to facilitate residual removal). This is followed by a generous tap water rinse and a distilled and deionized water rinse to remove the detergent. If aqueous sampling is to be performed, the following additional steps must be completed. An acid rinse, included if metals samples are to be collected, provides a low pH media for trace metals removal. It is followed by another distilled and deionized water rinse. If the sample is not to be analyzed for metals, the acid rinse and water rinse can be omitted. Next, a high purity solvent rinse is designated for trace organics removal. Acetone has been chosen because it is an excellent solvent, miscible in water and is not a targeted analyte in Priority Pollutant Analysis. If acetone is known to be a contaminant at a given site or Target Compound List analysis is to be performed, Methanol or another solvent may be substituted on a case by case basis with approval from NJDEP. Note, methanol can not be used when sampling gasoline and its by products. The solvent must be allowed to evaporate and then a final distilled and deionized water rinse is performed. This rinse removes any residual traces of the solvent.

All equipment utilized for sampling use must be decontaminated using distilled and deionized water. Through distillation, all ionized solids and a broad range of organic constituents will be removed, thus making it an ideal solvent for use when sampling for organic parameters. Deionized water is water that has been effectively freed from any existing ionic impurities. Deionized water should be used when sampling for inorganic parameters. The use of distilled and deionized water, commonly available from commercial vendors, is acceptable provided that the lot number and the associated analysis is available upon request to the NJDEP.

Whenever sampling, regardless of how equipment has been cleaned, always start sampling in the area of the site with the lowest contaminant probability and proceed to the areas of highest known or suspected contamination. Following this procedure will add another measure of quality control keeping cross contamination interference to a minimum.

3. Decontamination of Pumps

When submersible pumps are used to evacuate stagnant ground water in the well casing, they must be cleaned and flushed prior to and between each use. This cleaning process consists of an external laboratory grade glassware detergent wash and tap water rinse, or steam cleaning of pump casing, hose and cables followed by a 20 gallon flush of potable water through the pump. This flushing can be accomplished by the use of a clean plastic overpack drum or a plastic garbage can filled with potable water. This must be followed by a distilled and deionized rinse of the outside of the pump. For submersible pumps smaller than four inches in diameter, the recommended number of gallons required for flushing may be proportionately reduced (i.e. three inch - 15 gallons, two inch - 10 gallons). If the evacuation hose is not changed between locations, it must also be decontaminated in the same manner as the pump. Exercise caution to avoid contact with the pump casing and water in the drum while the pump is running (do not use metal drums or garbage cans) to avoid electric shock. Always disconnect the pump from power source before handling. Surface pumps (centrifugal and diaphragm) used for well evacuation need not be cleaned between well locations only if a check valve is used. New ASTM drinking water grade polyethylene tubing must be used for each well and discarded after use. The submersible pump and tubing should always be placed on clean polyethylene sheeting to avoid contact with the ground surface. All tubing must be rinsed/wiped with distilled and deionized water to remove any residual material before entering the well (Refer to ASTM D-5088-90, "Decontamination of Equipment").

4. Decontamination of Heavy Equipment

Other equipment and material associated with sampling episodes are required to be cleaned prior to usage. Items such as drill rigs, well casing, auger flights, and backhoes all present potential sources of interference to environmental samples. These items may come in contact with the materials adjacent to the matrix being sampled or may be attached to actual sampling equipment which has been cleaned in accordance with procedures set forth in Table 2-2. Heavy equipment may potentially retain contaminants from other sources such as roadways or storage areas or have soil material from previous job sites that have not been removed. For these reasons it is most important that these items be cleaned prior to usage during a site investigation (Refer to ASTM D-5088-90).

Two options are available to accomplish cleaning of heavy equipment: steam cleaning and manual scrubbing. The use of a steam generator can remove visible debris and has several advantages. Steam generators using potable water provide a high pressure medium which is very effective for residuals removal. They are also efficient in terms of ease of handling and generate low volumes of wash solutions.

Potential disadvantages include the need for a fixed or portable power source and they may not be cost effective for use on small pieces of equipment or for one day sampling events.

A second option involves manual scrubbing of equipment using a laboratory grade glassware detergent solution followed by a thorough water rinse. This procedure can be as effective as steam cleaning or preferred in situations where steam cleaning fails to remove visible materials. Disadvantages to manual scrubbing are that it is labor intensive and it generates large volumes of wash and rinse solutions.

The following requirements for cleaning heavy equipment should be incorporated into Field Sampling - Quality Assurance Project Plans where applicable.

5. Decontamination of Backhoes/Drill Rigs

These items should be thoroughly steam cleaned or manually scrubbed upon initial arrival on-site and between drilling or excavation locations. After the initial washing, cleaning may be reduced to those areas that are in close proximity to materials being sampled. The backhoe bucket and extension arm should be cleaned between each individual sample depth and between each test pit excavation. Drill rig items such as auger flights, drill rods, and drill bits must be cleaned between sample locations. Even when Shelby tubes or split spoons are used to collect samples that will not undergo chemical analysis, they must also be steam cleaned or scrubbed between each sample point (Refer to ASTM D-5088-90).

6. Decontamination of Monitor Well Casing and Screen

Before installation, field cleaning of well casing must consist of a manual scrubbing to remove foreign material and steam cleaning, inside and out, until all traces of oil and grease are removed. Special attention to threaded joints may be necessary to remove cutting oil or weld burn residues. The casing should then be handled and stored in such a manner so as to prevent cross contamination prior to installation.

7. Cleaning Location

It is preferred, given site specific conditions, that cleaning of all heavy equipment take place in one central location on site. A designated area or decontamination pad should be established to conduct all cleaning and provisions for containment of wash solutions must be made. All equipment such as drill rigs, backhoes, and other mobile equipment should receive an initial cleaning prior to use at a site. The frequency of subsequent cleanings while on-site will depend on how the equipment is actually used in relation to taking environmental samples. Unless otherwise specified and approved, all wash/rinse solutions should be collected and contained on-site. The actual fate of this material will be determined after review of analytical data generated from samples and on site discharge impacts have been evaluated.

8. Disposal of Drill Cuttings

During the routine course of site investigation, where materials are known (via field instrumentation or observation) or suspected (historic information) to be contaminated, sampling activity (i.e., soil boring or installation of monitoring wells) will produce waste intrinsic to the site. The disposition of this material must be carried out in a manner such as not to contribute further environmental degradation or pose a threat to public health or safety.

This material may be disposed of on-site provided: 1) the surface soils in areas of disposal are known or expected to be contaminated at levels above NJDEP soil standards in effect at that time; 2) the disposed soil/water will not erode/flow either off-site or on-site onto uncontaminated areas; 3) no potential to contaminate an uncontaminated aquifer exists, and; 4) the potential to create a health hazard to adjoining property owners through airborne exposure is non-existent. In addition, the requirements of the New Jersey Pollution Discharge Elimination System (NJPDES) must be followed for discharges to ground or surface waters.

If any of the above conditions cannot be met on-site, the materials must be placed in containers (drums, rolloffs, etc.) and stored in a secure area of the site (fenced or access by unauthorized persons prevented) or transported to a central secured location. The need to perform analyses of the secured material will be determined by NJDEP. The material will be retained for remediation or disposal in accordance with regulations as part of the selected site remedy.

When test pits are utilized for investigation, material excavated may be returned to the hole, but, excavation below the water table is not allowed. Holes produced from soil borings are to be grouted in accordance with NJDEP's Water Monitoring Management Program regulations unless NJDEP determines the material can be returned to the hole. Holes not grouted will be filled with sufficient quantities of material to make up for amount of soil sampled and account for settling thus allowing the hole to return to natural grade.

When materials of a noncontaminated nature are to be disposed of on-site, the following guidelines must be considered: 1.) disposed cuttings, soil or water will not erode or flow off-site; 2) disposed water will not flow through an area of contamination and thereby spread it to a clean area; or 3) NJDEP approves the disposal procedures.

Finally, at off-site (i.e. background) locations where no contamination is expected, the primary consideration is the wish of the property owner. If acceptable to the property owner, drill cuttings and muds from well installation may be raked into adjacent soils. If the property owner requests the uncontaminated material be removed from the site, it is to be properly contained and removed to the site under investigation and disposed of or stored per decision of the NJDEP. If drill cuttings and/or development water are expected to be contaminated, they are to be removed from the off-site location to a secure on-site location and retained for remediation or disposed of per applicable regulations.

9. Disposal of Development, Purge, Pump Test and Decon Waters

Similar to drill cuttings, an initial determination whether these waste waters should be considered contaminated should be made by evaluating field instrumentation readings or by previous analytical information. Additional field tests to assist in that determination (e.g. pH, color, other physical or chemical characterizations) must be utilized to the maximum extent possible.

Essentially, water generated that is not considered to be contaminated may be re-applied directly to the ground surface and permitted to percolate back into the groundwater system. Care should be taken, however, to avoid nuisance situations where the discharge may cause undue concern on the part of property owners or the community. In such cases, it is advisable to dispose of the water into a local stormwater or sanitary sewer system, or collect and discharge the water slowly to avoid such a condition. Please note that all discharges to surface water and/or the sanitary sewer are subject to the permit requirements contained in the NJPDES regulations.

Where the water is considered to be contaminated, the water generated may be re-applied to the ground surface provided all the following conditions are met:

1. The water is not permitted to migrate off-site.
2. There is no potential for contaminating a previously uncontaminated aquifer (for example, the discharge will not be permissible if a lower aquifer is being tested and is contaminated while the upper aquifer is not).
3. The discharge will not cause an increase to ground surface soil contamination.

If the above conditions cannot be met, the water shall be collected and secured at a single location (preferably the primary site under investigation). Collected water may be subsequently re-applied to ground surface only if, based on analytical results, there are indications that the above conditions can be met. If not, arrangements for proper disposal must be accomplished prior to the event.

In addition to the above considerations, the requirements of the NJPDES must be followed for all discharge to ground water and stormwater. The NJPDES Program requires the issuance of a permit for these discharges.

It is preferable to complete discharges of development, purge, and decon waters at a single, known contaminated area on-site. This area will be selected by the NJDEP. In cases where such an area cannot be located, as with contaminated well field projects, discharges will occur as close to the well or sampling location as reasonably possible.

D. *Procedures for Quality Assurance and Quality Control (QA/QC)*

QA/QC samples are intended to provide control over the collection of environmental measurements and subsequent validation, review, and interpretation of generated analytical

data. The various types of blank samples currently used by the NJDEP and related QA/QC concerns such as packaging, handling, preparation and actual procurement of samples from field locations are discussed below.

The trip blank is used exclusively for volatile organic analysis (aqueous sampling only) and its purpose is to measure possible cross contamination of samples during shipping to and from the site. The trip blank is never opened and travels to the site with the empty sample bottles and back from the site with the collected samples in an effort to simulate sampling handling conditions. Contaminated trip blanks may also indicate inadequate bottle cleaning or blank water of questionable quality.

The primary purpose of this type of blank is to detect additional sources of contamination that might potentially influence contaminant values reported in actual samples both quantitatively and qualitatively. The following have been identified as potential sources of contamination.

- Laboratory reagent water
- Sample containers
- Cross contamination in shipment, bottle handling and storage
- Ambient air or contact with analytical instrumentation during preparation and analysis at the laboratory
- Laboratory reagents used in analytical procedures

The purpose of a field blank is to place a mechanism of control on sample equipment handling, preparation, storage, and shipment. The field blank travels and is stored with the sample bottles, and is also representative of bottle shipment effects on sample quality. By being opened in the field and transferred over a cleaned sampling device (where applicable), the field blank is indicative of ambient conditions and/or equipment conditions that may potentially affect the quality of the associated samples.

The primary purpose of this type of blank is to provide an additional check on possible sources of contamination beyond that which is intended for trip blanks. A field blank serves a similar purpose as a trip blank regarding blank water quality and sample bottle preparation. However, it is primarily used to indicate potential contamination from ambient air as well as from sampling instruments used to collect and transfer samples from point of collection into sample containers (it may also be referred to as the field rinsate blank).

The following is a breakdown by matrix of blank sample requirements:

1. **Non-Aqueous Matrix**

a. Field Blanks

- i. Description - The performance of field blanks requires two (2) sets of identical bottles; one set filled with demonstrated analyte free water provided by the laboratory performing the sample analysis, and one empty set of bottles. The bottles should also be identical to those provided for aqueous sample collection. Note: Since field blanks are aqueous, the lab must provide water for volatile analysis in 40 ml septum vials. Although for soil VOA sample collection the lab may provide 4 oz wide mouth bottles. At the field location, in an area suspected to be contaminated, the water is passed from the full set of bottles through the dedicated or field decontaminated sampling device(s) and into the empty set of bottles. This will constitute identical bottle to bottle transfer. Field blanks must be preserved in the same manner as samples and only need to be collected and analyzed for volatile organics when volatile organics constitute a parameter being investigated. On a site specific basis, QA parameter requirements may be amended at the discretion of NJDEP. Note: for logistical purposes it is recommended that the lab provide at least one extra full 40 ml vial to perform the field blank.
- ii. Frequency - For sampling events lasting more than one day, field blanks for the non-aqueous matrix should be performed at a rate of 10% of the total number of samples collected throughout the event. If, for example, 40 samples were to be collected over a six day period, then only four field blanks would be required. For one-day sampling events, with the total number of samples collected being less than 10, it is required that one field blank be collected. On a site specific basis, QA frequency requirements may be amended at the discretion of NJDEP. However, it is not necessary to collect more than one field blank per day.

b. Trip Blanks

TRIP BLANKS ARE NOT REQUIRED FOR THE NON-AQUEOUS MATRIX unless specifically requested for Special Analytical Services (SAS) by NJDEP.

2. **Aqueous Matrix**

a. Field Blanks

- i. Description - Same as a.1. above with one exception: Field blanks must be analyzed for all the same parameters as samples collected that day.
- ii. Field Blanks are generally not required for potable well sampling events or when a sample is collected directly from a source into a sampling container.

- iii. Field Blanks may be required to detect cross contamination from ambient air during potable sampling events if known sources of contamination are within close proximity or monitoring instruments indicate the presence of contamination above background levels.
- iv. Frequency - Field blanks for the aqueous matrix must be performed at a rate of one per day.

b. Trip Blanks

- i. Description - Trip blanks are required for aqueous sampling events. They consist of a set of sample bottles filled at the laboratory with laboratory demonstrated analyte free water. These samples then accompany the bottles that are prepared at the lab into the field and back to the laboratory, along with the collected samples for analysis. These bottles are never opened in the field. Trip blanks must return to the lab with the same set of bottles they accompanied to the field. At a minimum trip blanks must be analyzed for volatile organic parameters. The inclusion of additional parameters or amendments to the requirements for trip blanks and associated samples shall not be held on site for more than two (2) calendar days.
- ii. Frequency - Trip blanks must be included at a rate of one per sample shipment (not to exceed two (2) consecutive field days).

3. Air Matrix

Trip and field blank procedures and frequencies for the various air sampling methods available should follow the specifications of the individual analytical method utilized. QA sample requirements may be amended at the discretion of NJDEP.

4. Blank Water Quality

The demonstrated analyte free water used in the field and trip blanks must originate from one common source and physical location within the laboratory and must be the same as the method blank water used by the laboratory performing the specific analysis. The use of commercially prepared water or water not originating from the laboratory analyzing the samples is not permitted. An exception to this requirement is allowable if:

- 1. it is the same water used for method blank analysis,
- 2. the laboratory has analyzed that water and generated data from a specific batch/lot of containers
- 3. the blank sample is drawn from an unopened container from the same batch/lot thus documenting the water is free of contaminants (demonstrated analyte free).

The laboratory performing the analysis may be required to provide documentation that trip and field blank water was demonstrated analyte free if contamination is detected in blanks, or at NJDEP's discretion. This would be verified by analytical results of method blanks run by the laboratory on the day of trip and field blank preparation and shipment. This does not, however, change requirements for the analysis of method blanks on the day of sample analysis at the laboratory.

A method blank is blank water which is carried through the entire sample preparation procedure and analysis at the laboratory. It is utilized as a check on laboratory procedures as well as possible contamination from laboratory equipment (i.e. reagents, glassware, etc.). The use of solid method blanks for volatiles and extractable organics is unacceptable to both USEPA, Region II and NJDEP. Method blanks associated with non-aqueous samples should consist of laboratory demonstrated analyte free water (documentation available upon request) which is prepared and analyzed in the same manner as the samples.

5. Sample Handling and Holding Times

a. Handling Time

Field and trip blank samples must travel with sample containers and must arrive on-site within one day of their preparation in the lab. Blanks and their associated samples may be held on-site for no longer than two calendar days, and must arrive back in the lab within one day of shipment from the field. This constitutes a maximum of a four (4) day handling time. Blanks and all samples must be maintained at 4°C while stored on-site and during shipment. Sample bottles and blanks must be handled in the same manner prior to their return to the laboratory.

The only acceptable exception to handling time requirements is when sampling stormwater runoff. The spontaneity of storm conditions precludes any possibility for preplanning sample bottle shipment. Therefore, due to these obvious logistical constraints, trip and field blanks are not normally required.

While the exception is understandable, the storage of these sample bottles must be carefully controlled to ensure the possibility of cross contamination is kept to an absolute minimum.

b. Maximum Holding Time

The clock governing holding times for samples and blanks analyzed by Contract Laboratory Program (CLP) methodologies begins when the sample is received in the laboratory as documented on the lab's chain of custody form verified time of sample receipt (VTSR). Holding times for individual parameters are dictated by the specific analytical method being used. The holding time clock for samples and blanks analyzed by SW-846 or 40 CFR, Parts

136 and 141, begins at the time of sample collection. For NJPDES, the holding times are those cited in 40 CFR 136.3.

6. Special Analytical Services (SAS)

It is important to note that both trip and field blanks are only capable of determining that contamination of samples may have occurred from additional sources other than the actual environmental matrix being investigated and cannot identify, but may suggest, possible sources of additional contaminant contribution to the reported analytical values. NJDEP may also implement additional types of QA/QC blanks (Special Analytical Services - SAS) when initial sampling episodes produce blank contamination that cause the generated data to become suspect. SAS has been developed to help identify the specific source(s) of blank contamination in a particular analytical fraction to aid in the assessment of reported analytical values. To date its primary purpose has been to confirm or eliminate laboratory contaminant sources in blank samples.

An example of special analytical services may be the inclusion of additional blank samples that are prepared at the same time and in the same manner as the trip and field blanks, but are designated for placement in laboratory storage areas, sample preparation areas or perhaps at ambient air ventilators or other field locations. These additional blanks are then subject to the same analysis as the samples to determine if location specific cross contamination during handling/storage may be occurring. SAS may also include the use of alternate analytical methodologies for unique, site specific parameters of concern.

7. Additional QA/QC Samples

Additional parameter blanks may be required in specific cases. NJDEP may make such a determination during review of the site specific sampling portion of a project plan.

a. Duplicate Samples Obtained in the Field

Collection of duplicate samples provides for the evaluation of the laboratory's performance by comparing analytical results of two samples from the same location. Duplicate samples are to be included for each matrix at a minimum rate of one for every twenty samples (5% of total) and be submitted to the lab as "blind" samples. If less than twenty samples are collected during a particular sampling episode, one duplicate should be performed. Duplicate requirements may be waived or expanded depending on the particular regulatory program or remedial phase involved.

i. Aqueous Matrix Duplicates

Duplicates of water samples (potable well, monitor well, surface water) should be obtained by alternately filling sample containers from the same sampling device for

each parameter. Samples for volatile organics analysis from monitor wells should be filled from the same bailer full of water whenever possible and be the first set of containers filled. When other sampling devices are used, the vials for volatile organics should be alternately filled. If heterogeneity is suspected, separate samples of each phase should be collected as the nature of phased liquids precludes homogenization. It is generally not necessary to homogenize groundwater or surface water samples.

ii. Non-Aqueous Matrix Duplicates

Obtaining duplicate samples in a soil or sediment matrix requires homogenization of the sample aliquot prior to filling sample containers. Regardless, volatile organic samples must always be taken from discrete locations or intervals without compositing or mixing. This practice is necessary to prevent loss of volatile constituents and to preserve, to the extent practicable, the physical integrity of the volatile fraction (see Chapter 7. "Sample Collection", Section B. "Soils" for further information on the procedures for collecting volatile organics). Homogenization of the sample for remaining parameters is necessary to generate two equally representative samples. Note that enough samples must be collected at one time in order to fill all the necessary sample containers. It may be necessary to co-locate or depth integrate collection so enough sample volume is available. A description of this process should be provided in the sampling plan. Moisture content, particle size, and adsorption properties of various oils, sediments, and waste materials may inhibit the ability to achieve complete mixing prior to filling sample containers.

Homogenization should be accomplished by filling a properly decontaminated stainless steel tray or bowl with the sample and mixing it with a decontaminated stainless steel or Teflon^R instrument. The extent of mixing required will depend on the nature of the sample and should be done to achieve a consistent physical appearance prior to filling sample containers.

Once mixing is completed the sample should be divided in half and containers should be filled by scooping sample material alternately from each half.

Several laboratory methodologies for compositing samples published by the American Society for Testing and Materials (ASTM) have been suggested for use in the field; however, they were not specifically designed for homogenization of known or suspected hazardous materials and often must be "modified" to be useful. They tend to assume a uniform sample exists to begin with and their intent may be to calculate average grain size, predict weight to volume ratios, or to reduce the size of a sample to one more convenient for handling and analysis. They also tend to assume a much larger volume of material will be subject to the particular methodology. Therefore, these methods are not recommended for generating duplicate samples in the field.

b. *Splitting Samples with Responsible Parties*

When various sites are under investigation, property owners and other interested parties may desire to obtain samples for analysis which are duplicates of those obtained by NJDEP personnel or contractors. If this becomes necessary, procedures for obtaining duplicate samples described above should be followed.

In order to maintain the integrity of any sample "split" between interested parties, the following procedures shall be followed:

- i. Personnel authorized by NJDEP (e.g. contractors or treatment facilities) using approved NJDEP sampling methods shall be permitted to obtain all sample aliquots.
- ii. Other interested parties must provide their own sample containers, blank samples, preservatives, sample shuttles, chain of custody forms, etc.
- iii. Sampling procedures shall be witnessed by NJDEP personnel to verify consistent handling and packaging of each set of samples.
- iv. Analytical data generated by responsible parties which is submitted for purposes of challenging NJDEP results or for informational purposes only shall first be subject to standard NJDEP "Quality Assurance Data and Validation of Analytical Sample Deliverables" procedures prior to being evaluated and considered for inclusion in the site evaluation process.
- v. Duplicate samples, trip blanks and field blanks must be included as part of those samples which are split between the two or more parties involved..
- vi. All interested parties desiring to obtain split samples during planned sampling episodes must provide the Department with a minimum of two weeks notice. This is essential for planning purposes and to avoid confusion or delays in the field.

c. *Performance Evaluation Samples (PE)*

In certain instances when a laboratory's quality assurance performance is in question, splitting samples may not prove as useful as providing blind performance evaluation samples to a laboratory since analytical performance and accuracy differs from laboratory to laboratory. One laboratory cannot be considered a "referee" whose performance can be considered the standard against which another's can be measured. Performance evaluation samples provide information on a laboratory's performance based upon analysis of that sample which contains parameters of a known and defined concentration. Therefore, at the discretion of NJDEP, PE samples may be required as a QA/QC check on laboratory performance for a particular sampling event or site investigation.

PE samples can be employed in two ways. First, the PE sample can be used to pre-qualify a laboratory. Since the current NJDEP Laboratory Certification procedures apply only to aqueous samples, a soil PE sample is useful in pre-qualifying a laboratory for soil or sediment evaluation projects. Second, a PE sample submitted blind with a sample lot to a contract laboratory is an available method for evaluating the quality of the analytical data.

PE samples consist of pre-measured, pre-determined samples of known origin and concentration which are submitted for analysis along with a sample shipment from the field. The PE samples are provided by NJDEP, and the contractor is responsible for their delivery to the laboratory performing the analysis along with other samples submitted for analysis that day. Data generated from the analysis of a PE sample is then compared to known concentrations as a measure of laboratory performance. Deviations from known concentrations may indicate improper calibration or other laboratory error that may have influenced the results reported for those samples collected in the field.

NJDEP maintains a bank of PE samples that may be drawn upon for individual sampling events. Dioxin PE samples are the most frequently utilized. Others include PCBs, BNAs, VOAs and Pesticides in soil Metals in sediment; and EPA prepared PE samples in an aqueous matrix. The particular concentration and species as well as assigned code numbers remain confidential until data is reviewed and validated.

d. Matrix Spike/Matrix Spike Duplicate Analyses

When performing Contract Laboratory Program (CLP) organic extractable analysis, the laboratory must be supplied with triple sample volume for each Sample Delivery Group (SDG) in order to perform matrix spike and matrix spike duplicate analyses. This does not include field or trip blanks. Blanks do not require separate matrix spike or duplicate analyses regardless of their matrix.

As stated in the CLP Statement of Work (SOW), the limits on an SDG are:

- each case of field samples, or
- each 20 field samples within a Case, or
- each 14 calendar day period during which field samples in a Case are received (said period beginning with the receipt of the first sample in the SDG), whichever comes first.

e. Background Samples

When background samples are required for comparison of site conditions to the surrounding environment they should be collected and handled in the same manner as all other samples. Requirements for inclusion of background samples are determined on a program specific and/or case by case basis.

f. Dioxin and Furans (PCDDS/PCDFs)

When dioxin or dioxin/furan sampling is performed, quality control samples may be required to accompany environmental samples regardless of the laboratory's status in the Contract Laboratory Program.

The quality control requirements for:

- each case of field samples received or
- each 20 field samples within a Case or
- each 14 calendar day period during which field samples in a case are received (beginning with the receipt of the first sample),

whichever is the most frequent, are usually defined by the method of analysis chosen.

The NJDEP reserves the right to choose one or more of the following quality control samples for incorporation into a sampling event. The quality control samples may be fortified field blanks, field duplicates, matrix spike/matrix spike duplicates, a fortified sample for each matrix, and/or a Performance Evaluation sample (PE).

A PE sample may be:

- a sample obtained by the NJDEP from the Environmental Measurement Soil Laboratory - Las Vegas (EMSL - LV) fortified with a known amount of native 2,3,7,8-substituted PCDDs/PCDFs or other PCDD/PCDF congeners (concentration unknown to the laboratory) or
- an uncontaminated sample obtained by the NJDEP from EMSL - LV which is to be analyzed by the laboratory for the appropriate PCDDs/PCDFs (concentration unknown to the laboratory) or
- a known blank obtained by the NJDEP from EMSL - LV that is to be fortified with a known amount of native material (Typically, a resultant concentration of 1 ug/Kg is used for soils/sediments and 10 ng/L is used for water).¹

E. Sample Preservation Requirements

Certain analytical methodologies for specific analytes require chemical additives in order to stabilize and maintain sample integrity. Generally this is accomplished under two scenarios:

- Preservative may be added to the sampling bottles by the laboratory prior to shipment into the field or,
- Preservatives are added in the field immediately after the samples are collected.

Many laboratories provide pre-preserved bottles as a matter of convenience and to help ensure that samples will be preserved immediately upon collection. A problem associated with this method arises if not enough sample is collected, resulting in too much preservative

TABLE 2-1 AQUEOUS SAMPLING EQUIPMENT DECONTAMINATION
(LABORATORY AND FIELD)

A. All field sampling equipment must be laboratory cleaned, wrapped, and dedicated to a particular sampling point or location during a sampling episode. In instances where this is not feasible, permission for field cleaning must be obtained from the NJDEP prior to the collection of any samples and be referenced in the approved site specific field sampling quality assurance project plan.

B. The field sampling equipment cleaning and decontamination procedures are as follows:

1. Laboratory grade glassware detergent plus tap water wash
2. Generous tap water rinse
3. Distilled and deionized (ASTM Type II) water rinse
4. 10% nitric acid rinse* (trace metal or higher grade HNO₃ diluted with Distilled and deionized (ASTM Type II) H₂O)
5. Distilled and deionized (ASTM Type II) water rinse*
6. Acetone (pesticide grade) rinse**
7. Total air dry or pure nitrogen blow out**
8. Distilled and deionized (ASTM Type II) water rinse**

* Only if sample is to be analyzed for metals.

** Only if sample is to be analyzed for organics.

C. Wherever possible, sampling devices should be numbered in a manner that will not affect their integrity and wrapped in a material (e.g., aluminum foil) that has been cleaned and oven baked at 105° C or cleaned in the same manner as the equipment (see footnote #4, Appendix 2-1). Equipment should be custody sealed and information concerning decontamination methodology, date, time, and personnel should be recorded in the field log book.

D. The use of distilled and deionized water commonly available from commercial vendors may be acceptable for sampling equipment decontamination. NJDEP may require specific lot numbers from containers or analytical verification that the distilled and deionized water meets ASTM Type II specifications.

E. Hexane is not a necessary solvent for dioxin, PCB, or other chlorinated organic sampling. The cleaning procedure outlined above is adequate for all sampling episodes.

F. In those instances where acetone is a parameter of concern, another solvent may be used.

G. Decontamination should be carried out over a container and the material properly disposed off-site. Decontamination wastes must be disposed of properly.

TABLE 2-2 NON-AQUEOUS SAMPLING EQUIPMENT DECONTAMINATION
(Laboratory and Field)

A. While it is preferred that all non-aqueous field sampling equipment be laboratory cleaned, wrapped, and dedicated to a particular sampling point or location during a sampling episode, field cleaning may be more practical.

B. The field sampling equipment cleaning and decontamination procedures are as follows:

1. Laboratory grade glassware detergent and tap water scrub to remove visual contamination
2. Generous tap water rinse
3. Distilled and deionized (ASTM Type II) water rinse

NOTE: If visual contamination persists, or gross contamination is suspected, the full eight step decontamination procedure found in Table 2-1 is recommended.

C. Wherever possible, sampling devices should be numbered in a manner that will not affect their integrity and wrapped in material (e.g., aluminum foil) that has been cleaned and oven baked at 105°C or cleaned in the same manner as the equipment (see footnote #4), Appendix 2-1). Equipment should be custody sealed and information concerning decontamination methodology, date, time, and personnel should be recorded in the field log book.

D. The use of distilled and deionized water commonly available from commercial vendors may be acceptable for sampling equipment decontamination. NJDEP may require specific lot numbers from containers or analytical verification that the distilled and deionized water meets ASTM Type II specifications.

E. Decontamination should be carried out over a container and the residual liquid material must be properly disposed. Decontamination wastes must be disposed in accordance with current NJDEP policy (see Chapter 2, Section C.9).

F. When analysis for metals is required, it may be necessary to use carbon steel split spoon sampling devices instead of stainless steel. If this is the case and it is necessary to utilize the acid rinse for removal of visible contamination, the nitric acid rinse may be lowered to a concentration of 1% instead of 10% so as to reduce the possibility of leaching metals from the spoon itself.

F:\Company Shared Folders\Brennan Env\TEMPLATE\BEIQAqc.doc

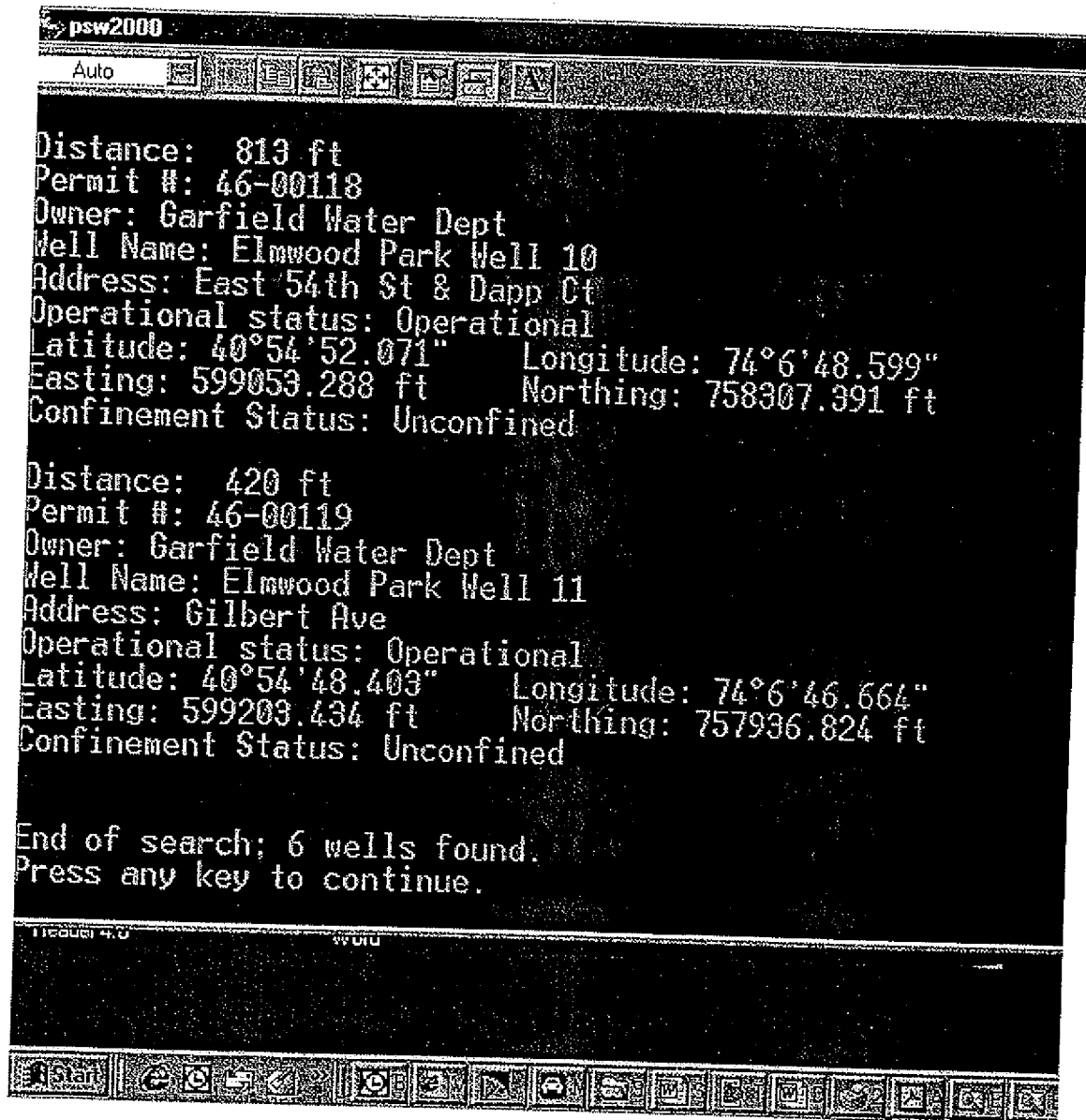


BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 7

PSW 2000







BRENNAN ENVIRONMENTAL, INC.
ENVIRONMENTAL CONSULTING SERVICES

APPENDIX 8

Laboratory Analytical Data Report





Integrated Analytical Laboratories, LLC.

273 Franklin Road
Randolph, N.J. 07869

Phone: 973 361-4252
Fax: 973 989-5288

ANALYTICAL DATA REPORT

for

Brennan Environmental, Inc.
8 Great Meadow Lane
Unit D
East Hanover, NJ 07936

Project: 99704 FINETEX

Lab Case Number: E01-7040

Date Report Prepared: October 17, 2001

<u>CLIENT SAMPLE ID</u>	<u>LABORATORY SAMPLE ID</u>
5-S1	7040-001
5-S2	7040-002
5-S3	7040-003
6-S1	7040-004
6-S2	7040-005
6-P1	7040-006
TB	7040-007
5-P1	7040-008

All required protocols were followed during analyses. These data have been reviewed and accepted by

A handwritten signature in cursive script, reading 'Michael H. Leftin', is written over a horizontal line.

Michael H. Leftin, Ph.D.
Laboratory Director

The liability of Integrated Analytical Laboratories, LLC. is limited to the actual cost of the analyses performed.



INTEGRATED ANALYTICAL LABORATORIES, LLC.

TABLE OF CONTENTS

	<u>Page</u>
Qualifiers	1
Conformance / NonConformance Summary	2
Laboratory Deliverables Check List	3
GC/MS NonConformance Summary	4
PHC NonConformance Summary	5
Summary Report	6
Analytical Results	
Volatiles	7
General Analytical	
Total Petroleum Hydrocarbons	10
Methodology Summary	11
Quality Control	
Volatiles	12
Tuning Results Summary	
Method Blank Results Summary	
Calibration Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Internal Standard Summary	
Chromatograms	
Petroleum Hydrocarbons	46
Blank Results Summary	
Spike Sample Results Summary	
Duplicate Sample Results Summary	
Calibration Curve	
Sample Tracking	
Chains of Custody	50
Laboratory Chronicle	54

INTEGRATED ANALYTICAL LABORATORIES, LLC.

MATRIX QUALIFIERS

- A - Indicates the sample is an Aqueous matrix.
- O - Indicates the sample is an Oil matrix.
- S - Indicates the sample is a Soil, Sludge or Sediment matrix.
- X - Indicates the sample is an Other matrix as indicated by Client Chain of Custody.

DATA QUALIFIERS

- B - Indicates the analyte was found in the Blank and in the sample. It indicates possible sample contamination and warns the data user to use caution when applying the results of the analyte.
- C - Common Laboratory Contaminant.
- D - The compound was reported from the Diluted analysis.
- D.F. - Dilution Factor.
- E - Estimated concentration, reported results are outside the calibrated range of the instrument.
- J - Indicates an estimated value. The compound was detected at a value below the method detection limit but greater than zero. For GC/MS procedures, the mass spectral data meets the criteria required to identify the target compound.
- MDL - Method Detection Limit.
- MI - Indicates compound concentration could not be determined due to Matrix Interferences.
- NA - Not Applicable.
- ND - Indicates the compound was analyzed for but Not Detected at the MDL.

REPORT QUALIFIERS

All solid sample analyses are reported on a dry weight basis.

All solid sample values are corrected for original sample size and percent solids.

INTEGRATED ANALYTICAL LABORATORIES, LLC.

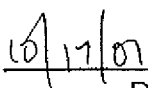
CONFORMANCE / NONCONFORMANCE SUMMARY

Integrated Analytical Laboratories, LLC. received eight (8) soil sample(s) from Brennan Environmental, Inc. (Project: 99704 FINETEX) on October 11, 2001 for the analysis of:

- (1) Volatiles + 10
- (1) Volatiles
- (7) Total Petroleum Hydrocarbons

A review of the QA/QC measures for the analysis of the sample(s) contained in this report has been performed by:


Reviewed by


Date

0002

INTEGRATED ANALYTICAL LABORATORIES, LLC.

LABORATORY DELIVERABLES CHECK LIST

Lab Case Number: E01-7040

	Check If Complete
1. Cover Page, Title Page listing Lab Certification #, facility name & address and date of report preparation.	<u>✓</u>
2. Table of Contents.	<u>✓</u>
3. Summary Sheets listing analytical results for all targeted and non-targeted compounds.	<u>✓</u>
4. Summary Table cross-referencing Field ID's vs. Lab ID's.	<u>✓</u>
5. Document bound, paginated and legible.	<u>✓</u>
6. Chain of Custody.	<u>✓</u>
7. Methodology Summary.	<u>✓</u>
8. Laboratory Chronicle and Holding Time Check.	<u>✓</u>
9. Results submitted on a dry weight basis (if applicable).	<u>✓</u>
10. Method Detection Limits.	<u>✓</u>
11. Lab certified by NJDEP for parameters or appropriate category of parameters or a member of the USEPA CLP.	<u>✓</u>
12. NonConformance Summary.	<u>✓</u>


QC Reviewed by

6/17/07
Date

0003

INTEGRATED ANALYTICAL LABORATORIES
CONFORMANCE/NONCONFORMANCE SUMMARY
GC/MS VOLATILE ANALYSIS

Lab Case Number: E01 - 2040

	No	Yes
1. Chromatograms Labeled/Compounds Identified (Field Samples and Method Blanks).	☐	☒
2. GC/MS Tuning Specifications:		☒
a. BFB Passed		
3. GC/MS Tuning Frequency - Performed every 24 hours for 600 series, 12 hours for 8000 series and 8 hours for 500 series.		☒
4. GC/MS Calibration - Initial calibration performed within 30 days before sample analysis and continuing calibration performed within 24 hours before sample analysis for 600 series, 12 hours for 8000 series		☒
5. GC/MS Calibration Requirements:		
a. Calibration Check Compounds		☒
b. System Performance Check Compounds		☒
6. Blank Contamination - If yes, list compounds and concentrations in each blank:	☒	
7. Surrogate Recoveries Meet Criteria (If not met, list those compounds and their recoveries which fall outside the acceptable range)		☒
If not met, were the calculations checked and the results qualified as "estimated"?		<u>na</u>
8. Matrix Spike/Matrix Spike Duplicate meet criteria (if not, list those compounds and their recoveries/% differences which fall outside the acceptable range)		☒
9. Internal Standard Area/Retention Time Shift meet criteria		☒
10. Extraction Holding Time Met		☒
If not met, list number of days exceeded for each sample:		<u>2/10</u>
11. Analysis Holding Time Met		☒
If not met, list number of days exceeded for each sample:		
12. Sample Dilution Performed		
High Target Compounds	High Nontarget Compounds	Matrix Interference
☐	☐	☐
☐	☐	☐

13. Comments:


Organics Manager

10/19/01
Date

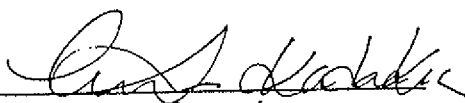
INTEGRATED ANALYTICAL LABORATORIES

PHC ANALYSIS CONFORMANCE/NONCONFORMANCE SUMMARY

Lab Case Number: E01 - 7040

- | | No | Yes |
|--|---------------|---------------|
| 1. Blank contamination | <u>✓</u> | <u> </u> |
| If yes, list the sample and the concentration in each blank: | | |
| <u> </u> | | |
| <u> </u> | | |
| 2. Matrix Spike/Matrix Spike Duplicate Recoveries Meet Criteria (If not met, list the samples and corresponding recovery which falls outside the acceptable range) | <u> </u> | <u>✓</u> |
| <u> </u> | | |
| <u> </u> | | |
| 3. IR spectra submitted for all standards, blanks & samples (as required). | <u> </u> | <u>✓</u> |
| <u> </u> | | |
| <u> </u> | | |
| 4. Chromatograms submitted for all standards, blanks & samples if GC fingerprinting was conducted. | <u> </u> | <u>NA</u> |
| <u> </u> | | |
| 5. Extraction Holding Time Met | <u> </u> | <u>✓</u> |
| If not met, list number of days exceeded for each sample: | | |
| <u> </u> | | |
| <u> </u> | | |
| 6. Analysis Holding Time Met | <u> </u> | <u>✓</u> |
| If not met, list number of days exceeded for each sample: | | |
| <u> </u> | | |
| <u> </u> | | |

Additional Comments:


 Dept. Supervisor

10/15/01
 Date

SUMMARY REPORT

Client: Brennan Environmental, Inc.

Project: 99704 FINETEX

Lab Case No.: E01-7040

Lab ID:	7040-001	7040-002	7040-003	7040-004
Client ID:	5-S1	5-S2	5-S3	6-S1
Matrix:	Soil	Soil	Soil	Soil
Sampled Date:	10/11/2001	10/11/2001	10/11/2001	10/11/2001
PARAMETER(Units)	Conc Q MDL	Conc Q MDL	Conc Q MDL	Conc Q MDL
General Analytical				
Total Petroleum Hydrocarbons (ppm)	516 25.0	ND 25.0	ND 25.0	168 25.0
Lab ID:	7040-005	7040-006	7040-007	7040-008
Client ID:	6-S2	6-P1	TB	5-P1
Matrix:	Soil	Soil	Soil	Soil
Sampled Date:	10/11/2001	10/11/2001	10/11/2001	10/11/2001
PARAMETER(Units)	Conc Q MDL	Conc Q MDL	Conc Q MDL	Conc Q MDL
Volatiles (ppb)				
Total Xylenes	863 J 1560	~ ~	ND 625	~ ~
TOTAL VO's:	863 J	~	ND	~
TOTAL TIC's:	21070	~ ~	~	~
TOTAL VO's & TIC's:	21933 J	~ ~	NA	~ ~
General Analytical				
Total Petroleum Hydrocarbons (ppm)	3820 119	170 25.0	~ ~	ND 25.0

~ = Sample not analyzed for

ND = Analyzed for but Not Detected at the MDL

J = The concentration was detected at a value below the MDL

All qualifiers on individual Volatiles are carried down through summation.

0006

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7040-005

Client ID: 6-S2

Date Received: 10/11/2001

Date Analyzed: 10/17/2001

Data file: I1953.D

GC/MS Column: DB-624

Sample wt/vol: 0.019g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 2

% Moisture: 15.8

Compound	Concentration	Q	MDL
Chloromethane	ND		1560
Vinyl Chloride	ND		1560
Bromomethane	ND		1560
Chloroethane	ND		1560
Trichlorofluoromethane	ND		1560
Acrolein	ND		3130
1,1-Dichloroethene	ND		1560
Methylene Chloride	ND		1560
Acrylonitrile	ND		3130
trans-1,2-Dichloroethene	ND		1560
1,1-Dichloroethane	ND		1560
Chloroform	ND		1560
1,1,1-Trichloroethane	ND		1560
Carbon Tetrachloride	ND		1560
1,2-Dichloroethane(EDC)	ND		1560
Benzene	ND		1560
Trichloroethene	ND		1560
1,2-Dichloropropane	ND		1560
Bromodichloromethane	ND		1560
2-Chloroethylvinyl Ether	ND		1560
cis-1,3-Dichloropropene	ND		1560
Toluene	ND		1560
trans-1,3-Dichloropropene	ND		1560
1,1,2-Trichloroethane	ND		1560
Tetrachloroethene	ND		1560
Dibromochloromethane	ND		1560
Chlorobenzene	ND		1560
Ethylbenzene	ND		1560
Total Xylenes	863	J	1560
Bromoform	ND		1560
1,1,2,2-Tetrachloroethane	ND		1560
1,3-Dichlorobenzene	ND		1560
1,4-Dichlorobenzene	ND		1560
1,2-Dichlorobenzene	ND		1560

Total Target Compounds: 863 J

0007

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Tentatively Identified Compounds

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7040-005

Client ID: 6-S2

Date Received: 10/11/2001

Date Analyzed: 10/17/2001

Date File: I1953.D

GC/MS Column: DB-624

Sample wt/vol: 0.019g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 2

% Moisture: 15.8

CAS #	Compound	Estimated Concentration	Retention Time
	Unknown alkane	5500	18.76
	Unknown alkane	1660	19.31
	Substituted benzene	1880	19.44
001120-21-4	Undecane	6910	20.96
	Unknown cyclic hydrocarbon	1840	22.10
000112-40-3	Dodecane	3280	23.01

Total TICs = 21070

0008

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7040-007

Client ID: TB

Date Received: 10/11/2001

Date Analyzed: 10/17/2001

Data file: I1950.D

GC/MS Column: DB-624

Sample wt/vol: 0.040g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 0

Compound	Concentration	Q	MDL
Chloromethane	ND		625
Vinyl Chloride	ND		625
Bromomethane	ND		625
Chloroethane	ND		625
Trichlorofluoromethane	ND		625
Acrolein	ND		1250
1,1-Dichloroethene	ND		625
Methylene Chloride	ND		625
Acrylonitrile	ND		1250
trans-1,2-Dichloroethene	ND		625
1,1-Dichloroethane	ND		625
Chloroform	ND		625
1,1,1-Trichloroethane	ND		625
Carbon Tetrachloride	ND		625
1,2-Dichloroethane(EDC)	ND		625
Benzene	ND		625
Trichloroethene	ND		625
1,2-Dichloropropane	ND		625
Bromodichloromethane	ND		625
2-Chloroethylvinyl Ether	ND		625
cis-1,3-Dichloropropene	ND		625
Toluene	ND		625
trans-1,3-Dichloropropene	ND		625
1,1,2-Trichloroethane	ND		625
Tetrachloroethene	ND		625
Dibromochloromethane	ND		625
Chlorobenzene	ND		625
Ethylbenzene	ND		625
Total Xylenes	ND		625
Bromoform	ND		625
1,1,2,2-Tetrachloroethane	ND		625
1,3-Dichlorobenzene	ND		625
1,4-Dichlorobenzene	ND		625
1,2-Dichlorobenzene	ND		625

Total Target Compounds: 0

0005

INTEGRATED ANALYTICAL LABORATORIES, LLC.

TOTAL PETROLEUM HYDROCARBONS

Client/Project: BRENNAN/99704 FINETEX

Date Received: 10/11/01

Lab ID	Client ID	Result	Q	DF	Matrix- Units	MDL	% Solid	Date Analyzed
7040-001	5-S1	516		1	Soil-mg/kg	25.0	84.9	10/15/2001
7040-002	5-S2	ND		1	Soil-mg/kg	25.0	82.3	10/15/2001
7040-003	5-S3	ND		1	Soil-mg/kg	25.0	81.3	10/15/2001
7040-004	6-S1	168		1	Soil-mg/kg	25.0	84.8	10/15/2001
7040-005	6-S2	3820		5	Soil-mg/kg	119	84.2	10/15/2001
7040-006	6-P1	170		1	Soil-mg/kg	25.0	85.4	10/15/2001
7040-008	5-P1	ND		1	Soil-mg/kg	25.0	88.5	10/15/2001

0010

TIERRA-B-010301

INTEGRATED ANALYTICAL LABORATORIES, LLC.

METHODOLOGY SUMMARY

<u>Analytical Parameter</u>	<u>Aqueous Samples</u> <u>Method #</u>	<u>Soil/Other Samples</u> <u>Method #</u>
Volatiles	NA	EPA 8260B
Total Petroleum Hydrocarbons	NA	EPA 418.1 M

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1746.D

BFB Injection Date: 10/10/2001

Inst ID: MSD I

BFB Injection Time: 9:22

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.5
75	30.0 - 60.0% of mass 95	57.3
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	6.4
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	69.3
175	5.0 - 9.0% of mass 174	5.9 (8.5)1
176	95.0 - 101.0% of mass 174	68.7 (99.1)1
177	5.0 - 9.0% of mass 176	4.5 (6.5)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
100	100_PPB_STD	I1747.D	10/10/2001	9:34
200	200_PPB_STD	I1748.D	10/10/2001	10:08
150	150_PPB_STD	I1749.D	10/10/2001	10:48
50	50_PPB_STD	I1750.D	10/10/2001	11:22
20	20_PPB_STD	I1751.D	10/10/2001	12:01
N/A	METHOD_BLK	I1752.D	10/10/2001	12:34
LCS	BLK_SPK	I1753.D	10/10/2001	13:07
TB	6805-017	I1754.D	10/10/2001	13:41
9B-2-V_3-3.5'	6805-002	I1755.D	10/10/2001	14:14
9B-2-VB_15.5-16'	6805-003	I1756.D	10/10/2001	14:48
9B-3-V_1.5-2'	6805-005	I1757.D	10/10/2001	15:22
9B-4_0-0.5'	6805-006	I1758.D	10/10/2001	15:56
9B-4-B_19.5-20'	6805-007	I1759.D	10/10/2001	16:29
9B-5-V_8.5-9'	6805-009	I1760.D	10/10/2001	17:03
9B-5B_11.5-12'	6805-010	I1761.D	10/10/2001	17:36
9B-1-V_9-9.5'	6805-012	I1762.D	10/10/2001	18:10
9B-1-B_13.5-14'	6805-013	I1763.D	10/10/2001	18:43
9B-6-V_8.5-9'	6805-015	I1764.D	10/10/2001	19:17
9B-6-B_11.5-12'	6805-016	I1765.D	10/10/2001	19:50
MS	6805-016MS	I1766.D	10/10/2001	20:24

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1746.D

BFB Injection Date : 10/10/200

Inst ID: MSD I

BFB Injection Time: 9:22

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.5
75	30.0 - 60.0% of mass 95	57.3
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	6.4
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	69.3
175	5.0 - 9.0% of mass 174	5.9 (8.5)1
176	95.0 - 101.0% of mass 174	68.7 (99.1)1
177	5.0 - 9.0% of mass 176	4.5 (6.5)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
MSD	6805-016MSD	I1767.D	10/10/2001	20:57

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1944.D

BFB Injection Date: 10/17/2001

Inst ID: MSD I

BFB Injection Time: 1:55

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.7
75	30.0 - 60.0% of mass 95	59.1
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	8.0
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	81.8
175	5.0 - 9.0% of mass 174	4.1 (5.0)1
176	95.0 - 101.0% of mass 174	80.0 (97.8)1
177	5.0 - 9.0% of mass 176	5.8 (7.2)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
100	100_PPb_STD	I1946.D	10/17/2001	2:56
N/A	METHOD_BLK	I1948.D	10/17/2001	4:02
LCS	BLK_SPK	I1949.D	10/17/2001	4:35
TB	7040-007	I1950.D	10/17/2001	5:09
TRIP_BLANK	6996-010	I1951.D	10/17/2001	5:42
31-E-FILL_MAT	06996-017	I1952.D	10/17/2001	6:15
6-S2	7040-005	I1953.D	10/17/2001	6:48
26D-1-B/2H-C-2-B	7090-003	I1955.D	10/17/2001	7:55
5R-3_0-0.5'	7090-005	I1956.D	10/17/2001	8:28
23A-3_0-0.5'	7090-008	I1957.D	10/17/2001	9:01
23A-4_0-0.5'	7090-009	I1958.D	10/17/2001	9:35
7B-1-V_1.5-2'	7090-010	I1959.D	10/17/2001	10:09
22G-3_0-0.5'	7090-012	I1960.D	10/17/2001	10:43
MS	7090-012MS	I1961.D	10/17/2001	11:17
MSD	7090-012MSD	I1962.D	10/17/2001	11:50
SB-43B_5-5.5	7085-005	I1963.D	10/17/2001	12:23
SB-43C_13.5-14	7085-006	I1964.D	10/17/2001	12:57

VOLATILE METHOD BLANK SUMMARY

Lab File ID: I1948.D

Instrument ID: MSD I

Date Analyzed: 10/17/2001

Time Analyzed: 04:02

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
LCS	BLK_SPK	10/17/2001	4:35
TB	7040-007	10/17/2001	5:09
TRIP_BLANK	6996-010	10/17/2001	5:42
31-E-FILL_MAT_0-9	6996-017	10/17/2001	6:15
6-S2	7040-005	10/17/2001	6:48
26D-1-B/2H-C-2-B	7090-003	10/17/2001	7:55
5R-3_0-0.5'	7090-005	10/17/2001	8:28
23A-3_0-0.5'	7090-008	10/17/2001	9:01
23A-4_0-0.5'	7090-009	10/17/2001	9:35
7B-1-V_1.5-2'	7090-010	10/17/2001	10:09
22G-3_0-0.5'	7090-012	10/17/2001	10:43
MS	7090-012MS	10/17/2001	11:17
MSD	7090-012MSD	10/17/2001	11:50
SB-43B_5-5.5	7085-005	10/17/2001	12:23
SB-43C_13.5-14	7085-006	10/17/2001	12:57

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/17/2001

Data file: I1948.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

Compound	Concentration	Q	MDL
Chloromethane	ND		250
Vinyl Chloride	ND		250
Bromomethane	ND		250
Chloroethane	ND		250
Trichlorofluoromethane	ND		250
Acrolein	ND		500
1,1-Dichloroethene	ND		250
Methylene Chloride	ND		250
Acrylonitrile	ND		500
trans-1,2-Dichloroethene	ND		250
1,1-Dichloroethane	ND		250
Chloroform	ND		250
1,1,1-Trichloroethane	ND		250
Carbon Tetrachloride	ND		250
1,2-Dichloroethane(EDC)	ND		250
Benzene	ND		250
Trichloroethene	ND		250
1,2-Dichloropropane	ND		250
Bromodichloromethane	ND		250
2-Chloroethylvinyl Ether	ND		250
cis-1,3-Dichloropropene	ND		250
Toluene	ND		250
trans-1,3-Dichloropropene	ND		250
1,1,2-Trichloroethane	ND		250
Tetrachloroethene	ND		250
Dibromochloromethane	ND		250
Chlorobenzene	ND		250
Ethylbenzene	ND		250
Total Xylenes	ND		250
Styrene	ND		250
Bromoform	ND		250
1,1,2,2-Tetrachloroethane	ND		250
1,3-Dichlorobenzene	ND		250
1,4-Dichlorobenzene	ND		250
1,2-Dichlorobenzene	ND		250
Naphthalene	ND		250

Total Target Compounds: 0

0016

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Tentatively Identified Compounds

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/17/2001

Data file: I1948.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

CAS #	Compound	Estimated Concentration	Retention Time
-------	----------	----------------------------	-------------------

No peaks detected

Total TICs = 0

0017

Response Factor Report 5970 - In

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Last Update : Wed Oct 10 12:41:09 2001
 Response via : Initial Calibration

Calibration Files

20 =I1751.D 50 =I1750.D 100 =I1747.D
 150 =I1749.D 200 =I1748.D

	Compound	20	50	100	150	200	Avg	%RSD
1) I	Pentafluorobenzene	-----ISTD-----						
2) T	Dichlorodifluorometha	0.315	0.311	0.294	0.284	0.290	0.299	4.58
3) P	Chloromethane	0.229	0.244	0.248	0.228	0.231	0.236	3.91
4) C	Vinyl Chloride	0.209	0.216	0.219	0.208	0.209	0.212	2.36
5) T	Bromomethane	0.289	0.265	0.233	0.219	0.223	0.246	12.24
6) T	Chloroethane	0.163	0.164	0.168	0.159	0.161	0.163	1.99
7) T	Trichlorofluoromethan	0.866	0.828	0.778	0.784	0.769	0.805	5.11
8) T	Acrolein	0.009	0.009	0.007	0.007	0.009	0.008	14.00
9) MC	1,1-Dichloroethene	0.274	0.278	0.279	0.260	0.263	0.271	3.26
10) T	Acetone	0.065	0.087	0.093	0.070	0.065	0.076	17.35
11) T	Carbon Disulfide	0.789	0.756	0.783	0.770	0.754	0.770	2.04
12) T	Vinyl Acetate	0.997	1.052	0.915	0.935	0.877	0.955	7.24
13) T	Methylene Chloride	0.310	0.316	0.301	0.300	0.284	0.302	3.95
14) T	Acrylonitrile	0.032	0.035	0.034	0.032	0.036	0.034	5.40
15) T	t-Butyl Alcohol (TBA)	0.018	0.016	0.019	0.020	0.016	0.018	9.22
16) T	trans-1,2-Dichloroeth	0.332	0.340	0.346	0.326	0.315	0.332	3.65
17) T	Methyl-t-Butyl Ether(	0.833	0.872	0.737	0.799	0.717	0.792	8.17
18) P	1,1-Dichloroethane	0.765	0.790	0.758	0.738	0.719	0.754	3.57
19) T	Diisopropyl Ether(DIP	1.467	1.485	1.351	1.382	1.319	1.401	5.18
20) T	cis-1,2-Dichloroethen	0.360	0.358	0.363	0.349	0.340	0.354	2.62
21) T	2,2-Dichloropropane	0.646	0.646	0.643	0.629	0.614	0.636	2.23
22) T	2-Butanone (MEK)	0.108	0.106	0.073	0.083	0.073	0.089	19.37
23) T	Bromochloromethane	0.229	0.236	0.220	0.221	0.208	0.223	4.72
25) C	Chloroform	1.040	1.014	1.000	0.975	0.959	0.998	3.21
26) T	1,1,1-Trichloroethane	0.950	0.950	0.911	0.920	0.885	0.923	3.02
27) T	Carbon Tetrachloride	0.708	0.775	0.666	0.734	0.685	0.714	5.97
28) T	1,1-Dichloropropene	0.542	0.544	0.519	0.524	0.509	0.527	2.83
29) T	1,2-Dichloroethane (ED	0.717	0.742	0.665	0.692	0.634	0.690	6.15
30) S	1,2-Dichloroethane-d4	0.592	0.649	0.607	0.660	0.651	0.632	4.77
31) I	1,4-Difluorobenzene	-----ISTD-----						
32) M	Benzene	0.698	0.703	0.706	0.675	0.648	0.686	3.54
33) M	Trichloroethene	0.476	0.460	0.454	0.430	0.414	0.447	5.48
34) C	1,2-Dichloropropane	0.353	0.342	0.332	0.322	0.314	0.333	4.69
35) T	Dibromomethane	0.304	0.319	0.282	0.278	0.256	0.288	8.45
37) T	Bromodichloromethane	0.809	0.807	0.755	0.788	0.708	0.773	5.47
38) T	2-Chloroethylvinyl Et	0.145	0.160	0.139	0.149	0.136	0.146	6.60
39) T	cis-1,3-Dichloroprope	0.478	0.503	0.476	0.461	0.435	0.471	5.32
40) T	4-Methyl-2-pentanone(	0.226	0.240	0.177	0.205	0.178	0.205	13.74
41) S	Toluene-d8	0.780	0.788	0.783	0.770	0.796	0.784	1.21
42) MC	Toluene	0.476	0.489	0.462	0.444	0.436	0.461	4.77
43) T	trans-1,3-Dichloropro	0.435	0.448	0.399	0.403	0.376	0.412	7.08
44) T	1,1,2-Trichloroethane	0.212	0.223	0.189	0.205	0.185	0.203	7.75
45) T	Tetrachloroethene	0.390	0.375	0.357	0.344	0.334	0.360	6.29
46) T	1,3-Dichloropropane	0.395	0.407	0.360	0.367	0.332	0.372	7.97
47) T	2-Hexanone	0.158	0.163	0.123	0.137	0.120	0.140	14.11
48) T	Dibromochloromethane	0.492	0.521	0.435	0.456	0.423	0.465	8.78
49) T	1,2-Dibromoethane (EDB	0.350	0.335	0.297	0.308	0.289	0.316	8.22

0018

		-----ISTD-----						
50) I	Chlorobenzene-d5							
51) MP	Chlorobenzene	0.822	0.827	0.782	0.761	0.760	0.790	4.09
52) T	1,1,1,2-Tetrachloroet	0.532	0.545	0.480	0.524	0.491	0.514	5.37
53) C	Ethylbenzene	1.286	1.301	1.200	1.217	1.170	1.235	4.56
54) T	m,p-Xylene	0.436	0.437	0.413	0.415	0.398	0.420	3.97
55) T	o-Xylene	0.461	0.440	0.397	0.405	0.398	0.420	6.94
56) T	Styrene	0.697	0.737	0.654	0.705	0.680	0.695	4.47
57) P	Bromoform	0.371	0.399	0.342	0.382	0.346	0.368	6.52
58) T	Isopropylbenzene	1.141	1.076	1.078	1.069	1.036	1.080	3.55
59) S	Bromofluorobenzene	0.895	0.905	0.847	0.881	0.883	0.882	2.51
60) P	1,1,2,2-Tetrachloroet	0.337	0.349	0.276	0.314	0.287	0.313	10.06
61) T	Bromobenzene	0.452	0.403	0.382	0.400	0.390	0.405	6.72
62) T	1,2,3-Trichloropropan	0.422	0.388	0.322	0.334	0.318	0.357	12.89
63) T	n-Propylbenzene	1.332	1.269	1.275	1.195	1.207	1.255	4.45
64) T	2-Chlorotoluene	1.039	1.006	0.931	0.991	0.942	0.982	4.57
65) T	1,3,5-Trimethylbenzen	0.991	0.938	0.880	0.926	0.897	0.926	4.59
66) T	4-Chlorotoluene	1.215	1.200	1.120	1.178	1.085	1.160	4.76
67) T	tert-Butylbenzene	0.884	0.843	0.835	0.792	0.822	0.835	4.02
68) T	1,2,4-Trimethylbenzen	0.995	0.986	0.946	0.971	0.911	0.962	3.51
69) T	sec-Butylbenzene	0.994	0.958	0.906	0.925	0.900	0.937	4.18
70) T	1,3-Dichlorobenzene	0.658	0.683	0.573	0.616	0.580	0.622	7.73
71) T	4-Isopropyltoluene	0.879	0.824	0.817	0.769	0.777	0.813	5.42
72) T	1,4-Dichlorobenzene	0.658	0.683	0.573	0.616	0.580	0.622	7.73
73) T	n-Butylbenzene	0.850	0.805	0.796	0.754	0.757	0.792	4.95
74) T	1,2-Dichlorobenzene	0.586	0.569	0.477	0.516	0.468	0.523	10.18
75) T	1,2-Dibromo-3-chlorop	0.109	0.104	0.063	0.080	0.069	0.085	24.49
76) T	1,2,4-Trichlorobenzen	0.194	0.188	0.131	0.229	0.173	0.183	19.49
77) T	Hexachlorobutadiene	0.176	0.127	0.135	0.112	0.117	0.133	19.12
78) T	Naphthalene	0.249	0.315	0.194	0.361	0.241	0.272	24.19
79) T	1,2,3-Trichlorobenzen	0.167	0.196	0.094	0.171	0.115	0.149	28.61
80) T	1,1,2-Trichloro-1,2,2	0.489	0.460	0.415	0.402	0.409	0.435	8.68
81) T	Methyl Acetate	0.173	0.175	0.134	0.159	0.155	0.159	10.39
82) T	Cyclohexane	0.323	0.295	0.266	0.265	0.264	0.283	9.19
83) T	Methylcyclohexane	0.212	0.208	0.180	0.178	0.184	0.192	8.39

(#) = Out of Range

IME1010.M

Wed Oct 10 12:41:37 2001 MSD_I

0019

TIERRA-B-010310

Instrument ID: MSD_I
Method ID: SOIL1003.M
Date: 10/03/2001

Average %RSD = 10.22

Refer to SW846 Method 8000B Section 7.5.1.

0020

Evaluate Continuing Calibration Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1946.D

Vial: 29

Acq On : 17 Oct 2001 2:56

Operator: DP

Sample : 100,100_PP8_STD,W,5mL,100

Inst : 5970 - In

Misc : N/A,N/A,N/A

Multiplr: 1.00

MS Integration Params: rteint.p

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Multiple Level Calibration

Min. RRF : 0.000 Min. Rel. Area : 50% Max. R.T. Dev 0.50min

Max. RRF Dev : 30% Max. Rel. Area : 200%

	Compound	AvgRF	CCRF	%Dev	Area%	Dev(min)
1 I	Pentafluorobenzene	1.000	1.000	0.0	167	-0.03
2 T	Dichlorodifluoromethane	0.299	0.337	-12.7	192	-0.02
3 P	Chloromethane	0.236	0.187	20.8	127	-0.01
4 C	Vinyl Chloride	0.212	0.181	14.6	139	-0.03
5 T	Bromomethane	0.246	0.249	-1.2	179	-0.02
6 T	Chloroethane	0.163	0.152	6.7	152	-0.02
7 T	Trichlorofluoromethane	0.805	0.616	23.5	132	-0.04
8 T	Acrolein	0.008	0.007	12.5	183	-0.03
9 MC	1,1-Dichloroethene	0.271	0.266	1.8	160	-0.03
10 T	Acetone	0.076	0.071	6.6	128	-0.05
11 T	Carbon Disulfide	0.770	0.720	6.5	154	-0.02
12 T	Vinyl Acetate	0.955	0.956	-0.1	175	-0.04
13 T	Methylene Chloride	0.302	0.304	-0.7	169	-0.03
14 T	Acrylonitrile	0.034	0.031	8.8	154	-0.05
15 T	t-Butyl Alcohol (TBA)	0.018	0.014	22.2	122	-0.05
16 T	trans-1,2-Dichloroethene	0.332	0.340	-2.4	164	-0.05
17 T	Methyl-t-Butyl Ether (MTBE)	0.792	0.734	7.3	167	-0.04
18 P	1,1-Dichloroethane	0.754	0.713	5.4	158	-0.03
19 T	Diisopropyl Ether (DIPE)	1.401	1.309	6.6	162	-0.03
20 T	cis-1,2-Dichloroethene	0.354	0.361	-2.0	166	-0.04
21 T	2,2-Dichloropropane	0.636	0.519	18.4	135	-0.04
22 T	2-Butanone (MEK)	0.089	0.077	13.5	176	-0.04
23 T	Bromochloromethane	0.223	0.220	1.3	167	-0.03
25 C	Chloroform	0.998	0.902	9.6	151	-0.04
26 T	1,1,1-Trichloroethane	0.923	0.748	19.0	137	-0.03
27 T	Carbon Tetrachloride	0.714	0.577	19.2	145	-0.02
28 T	1,1-Dichloropropene	0.527	0.498	5.5	161	-0.03
29 T	1,2-Dichloroethane (EDC)	0.690	0.594	13.9	149	-0.04
30 S	1,2-Dichloroethane-d4	0.632	0.536	15.2	148	-0.02
31 I	1,4-Difluorobenzene	1.000	1.000	0.0	177	-0.04
32 M	Benzene	0.686	0.698	-1.7	175	-0.04
33 M	Trichloroethene	0.447	0.400	10.5	156	-0.03
34 C	1,2-Dichloropropane	0.333	0.305	8.4	163	-0.03
35 T	Dibromomethane	0.288	0.235	18.4	148	-0.03
37 T	Bromodichloromethane	0.773	0.647	16.3	152	-0.04
38 T	2-Chloroethylvinyl Ether	0.146	0.121	17.1	154	-0.03
39 T	cis-1,3-Dichloropropene	0.471	0.423	10.2	157	-0.03
40 T	4-Methyl-2-pentanone (MIBK)	0.205	0.170	17.1	170	-0.02
41 S	Toluene-d8	0.784	0.795	-1.4	180	-0.03
42 MC	Toluene	0.461	0.462	-0.2	177	-0.03
43 T	trans-1,3-Dichloropropene	0.412	0.348	15.5	155	-0.03
44 T	1,1,2-Trichloroethane	0.203	0.192	5.4	180	-0.03
45 T	Tetrachloroethene	0.360	0.318	11.7	158	-0.02
46 T	1,3-Dichloropropane	0.372	0.328	11.8	161	-0.02

47	T	2-Hexanone	0.140	0.126	10.0	182	-0.02
48	T	Dibromochloromethane	0.465	0.381	18.1	155	-0.03
49	T	1,2-Dibromoethane (EDB)	0.316	0.271	14.2	162	-0.02
50	I	Chlorobenzene-d5	1.000	1.000	0.0	183	-0.03
51	MP	Chlorobenzene	0.790	0.722	8.6	169	-0.02
52	T	1,1,1,2-Tetrachloroethane	0.514	0.417	18.9	159	-0.03
53	C	Ethylbenzene	1.235	1.099	11.0	167	-0.03
54	T	m,p-Xylene	0.420	0.394	6.2	174	-0.02
55	T	o-Xylene	0.420	0.380	9.5	175	-0.02
56	T	Styrene	0.695	0.637	8.3	178	-0.02
57	P	Bromoform	0.368	0.277	24.7	148	-0.03
58	T	Isopropylbenzene	1.080	0.919	14.9	156	0.00
59	S	Bromofluorobenzene	0.882	0.809	8.3	175	-0.01
60	P	1,1,2,2-Tetrachloroethane	0.313	0.257	17.9	171	-0.02
61	T	Bromobenzene	0.405	0.346	14.6	165	-0.02
62	T	1,2,3-Trichloropropane	0.357	0.253	29.1	144	0.00
63	T	n-Propylbenzene	1.255	1.022	18.6	147	-0.03
64	T	2-Chlorotoluene	0.982	0.840	14.5	165	-0.03
65	T	1,3,5-Trimethylbenzene	0.926	0.783	15.4	163	0.00
66	T	4-Chlorotoluene	1.160	0.947	18.4	155	-0.02
67	T	tert-Butylbenzene	0.835	0.686	17.8	150	-0.03
68	T	1,2,4-Trimethylbenzene	0.962	0.799	16.9	155	-0.01
69	T	sec-Butylbenzene	0.937	0.809	13.7	163	0.00
70	T	1,3-Dichlorobenzene	0.622	0.503	19.1	161	-0.01
71	T	4-Isopropyltoluene	0.813	0.643	20.9	144	-0.01
72	T	1,4-Dichlorobenzene	0.622	0.503	19.1	161	-0.01
73	T	n-Butylbenzene	0.792	0.617	22.1	142	-0.01
74	T	1,2-Dichlorobenzene	0.523	0.402	23.1	154	-0.03
75	T	1,2-Dibromo-3-chloropropane	0.085	0.065	23.5	190	0.00
76	T	1,2,4-Trichlorobenzene	0.183	0.146	20.2	205	-0.02
77	T	Hexachlorobutadiene	0.133	0.077	42.1	105	0.00
78	T	Naphthalene	0.272	0.210	22.8	198	0.00
79	T	1,2,3-Trichlorobenzene	0.149	0.098	34.2	191	-0.02
80	T	1,1,2-Trichloro-1,2,2-trifl	0.435	0.430	1.1	190	-0.03
81	T	Methyl Acetate	0.159	0.132	17.0	180	-0.05
82	T	Cyclohexane	0.283	0.237	16.3	163	-0.03
83	T	Methylcyclohexane	0.192	0.154	19.8	156	-0.03

(#) = Out of Range
I1747.D IME1010.M

SPCC's out = 0 CCC's out = 0
Thu Oct 18 13:35:20 2001 MSD_I

VOLATILE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/17/2001

Lab Sample ID	Matrix	File ID	SMC1 #	SMC2 #	SMC3 #
METHOD_BLK	SOIL	I1948.D	81	100	91
BLK_SPK	SOIL	I1949.D	76	100	87
7040-007	MEOH	I1950.D	85	97	77
6996-010	MEOH	I1951.D	83	91	76
6996-017	MEOH	I1952.D	81	77	72
7040-005	MEOH	I1953.D	84	98	74
7090-003	MEOH	I1955.D	71	73	61
7090-005	MEOH	I1956.D	74	80	65
7090-008	MEOH	I1957.D	79	84	66
7090-009	MEOH	I1958.D	77	88	66
7090-010	MEOH	I1959.D	78	77	63
7090-012	MEOH	I1960.D	79	82	66
7090-012MS	MEOH	I1961.D	86	88	75
7090-012MSD	MEOH	I1962.D	82	86	69
7085-005	MEOH	I1963.D	100	100	100
7085-006	MEOH	I1964.D	80	90	70

	Concentration	Aqueous/Meoh	Soil
SMC1 = 1,2-Dichloroethane-d4	50 ppb	48-125	51-118
SMC2 = Toluene-d8	50 ppb	54-123	89-109
SMC3 = Bromofluorobenzene	50 ppb	46-118	80-111

Column to be used to flag recovery values

MEOH VOLATILE MATRIX SPIKE/SPIKE DUPLICATE RECOVERY

Matrix spike Lab sample ID: 7090-012

Batch No.: IM0233

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
1,1-Dichloroethene	50.0	0.0	48.6	97	48 - 149
Benzene	50.0	0.0	47.4	95	59 - 149
Trichloroethene	50.0	0.0	39.9	80	55 - 143
Toluene	50.0	0.0	45.5	91	54 - 150
Chlorobenzene	50.0	0.0	47.3	95	55 - 144

Compound	SAMPLE CONC. (ug/Kg)	MSD CONC. (ug/Kg)	MSD % # REC	% RPD #	QC LIMITS	
					RPD	REC.
1,1-Dichloroethene	0.0	49.8	100	3	17	48 - 149
Benzene	0.0	46.3	93	2	15	59 - 149
Trichloroethene	0.0	38.5	77	4	15	55 - 143
Toluene	0.0	45.6	91	0	16	54 - 150
Chlorobenzene	0.0	48.4	97	2	15	55 - 144

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

RPD: 0 out of 5 outside limits

Spike Recovery: 0 out of 10 outside limits

0024

SOIL VOLATILE LABORATORY CONTROL SAMPLE RECOVERY

Matrix spike Lab sample ID: BLK SPK

Batch No.: IM0233

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
1,1-Dichloroethene	50.0	0.0	55.8	112	70 - 130
Benzene	50.0	0.0	51.5	103	70 - 130
Trichloroethene	50.0	0.0	43.7	87	70 - 130
Toluene	50.0	0.0	52.0	104	70 - 130
Chlorobenzene	50.0	0.0	47.3	95	70 - 130

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

Spike Recovery: 0 out of 5 outside limits

0025

VOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): 11747.D

Date Analyzed: 10/10/2001

Instrument ID: MSD_I

Time Analyzed: 9:34

	50UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
	12 HOUR STD	653387	9.73	784077	10.95	634368	15.90
	UPPER LIMIT	1306774	10.23	1568154	11.45	1268736	16.40
	LOWER LIMIT	326693.5	9.23	392038.5	10.45	317184	15.40
	LAB SAMPLE ID						
01	200_PPb_STD	647856	9.70	798050	10.90	615497	15.84
02	150_PPb_STD	656854	9.65	814005	10.87	640682	15.82
03	50_PPb_STD	675342	9.65	821122	10.84	635952	15.80
04	20_PPb_STD	674167	9.65	799529	10.87	623610	15.82
05	METHOD_BLK	648350	9.64	792486	10.86	668256	15.81
06	BLK_SPK	656491	9.64	789960	10.86	643953	15.82
07	6805-017	694488	9.63	832579	10.85	592975	15.82
08	6805-002	700505	9.65	828125	10.87	586715	15.83
09	6805-003	708272	9.65	845648	10.88	603174	15.83
10	6805-005	770218	9.65	860282	10.85	626482	15.82
11	6805-006	758939	9.63	860569	10.85	628899	15.82
12	6805-007	779707	9.64	875607	10.86	633294	15.82
13	6805-009	734189	9.65	892746	10.87	624207	15.82
14	6805-010	775090	9.66	914234	10.86	684844	15.81
15	6805-012	787746	9.63	907557	10.85	657037	15.80
16	6805-013	788950	9.63	913993	10.85	642244	15.80
17	6805-015	795434	9.62	941670	10.85	663946	15.81
18	6805-016	803676	9.63	945405	10.86	671447	15.81
19	6805-016MS	813939	9.65	941658	10.86	677527	15.81
20	6805-016MSD	795946	9.66	950448	10.87	668293	15.83
21							
22							

IS1 = PENTAFLUOROBENZENE

IS2 = 1,4-DIFLUOROBENZENE

IS3 = CHLOROBENZENE-D5

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

VOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): I1946.D

Date Analyzed: 10/17/2001

Instrument ID: MSD_I

Time Analyzed: 2:56

50UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
12 HOUR STD	1093433	9.70	1389694	10.92	1160240	15.86
UPPER LIMIT	2186866	10.20	2779388	11.42	2320480	16.36
LOWER LIMIT	546716.5	9.20	694847	10.42	580120	15.36
LAB SAMPLE ID						
01 METHOD_BLK	1047735	9.71	1387504	10.92	1182634	15.87
02 BLK_SPK	1069109	9.71	1381684	10.92	1165754	15.88
03 7040-007	1063650	9.70	1430560	10.92	1160991	15.87
04 6996-010	1092890	9.71	1407551	10.92	1169632	15.88
05 6996-017	1174540	9.71	1498070	10.91	1071222	15.88
06 7040-005	1186341	9.71	1443196	10.93	1087576	15.88
07 7090-003	1216331	9.72	1542878	10.93	1105934	15.88
08 7090-005	1235988	9.71	1497297	10.91	1101262	15.88
09 7090-008	1188178	9.68	1480973	10.90	1116937	15.85
10 7090-009	1208938	9.67	1461196	10.90	1146334	15.84
11 7090-010	1185468	9.67	1434970	10.87	1111598	15.83
12 7090-012	1211880	9.64	1493263	10.86	1110437	15.80
13 7090-012MS	1176326	9.61	1476249	10.83	1089950	15.78
14 7090-012MSD	1161925	9.61	1477952	10.82	1100599	15.78
15 7085-005	1168648	9.63	1391800	10.85	1070052	15.82
16 7085-006	1155584	9.62	1387096	10.84	1071641	15.79
17						
18						
19						
20						
21						
22						

IS1 = PENTAFLUOROBENZENE

IS2 = 1,4-DIFLUOROBENZENE

IS3 = CHLOROBENZENE-D5

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

FORM 8

0027

VOLATILE METHANOL PRESERVATION CONTAINER SUMMARY

Sample ID	Client ID	Bottle ID	Date Prepared	Initial Wgt.(g)	Final Wgt.(g)	Sample Wgt.(g)	MeOH Lot #	Surrogate Lot #
7040-005	6-S2	111765	10/9/01	106.6	116.2	9.6	cc226	ls224
7040-007	TB	111772	10/9/01	107.3	117.3	10.0	cc226	ls224

Limits : Sample Weight = 8 to 12 grams, * = Outside limits

Comments : _____

Sample weight is reported on a wet weight basis

0028

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D Vial: 46
Acq On : 17 Oct 2001 6:48 Operator: DP
Sample : 6-S2,7040-005,M,0.019g Inst : 5970 - In
Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
MS Integration Params: rteint.p
Quant Time: Oct 17 14:31 2001 Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Last Update : Thu Oct 11 09:23:20 2001
Response via : Initial Calibration
DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev (Min)
1) Pentafluorobenzene	9.71	168	1186341	50.00	UG	-0.02
31) 1,4-Difluorobenzene	10.93	114	1443196	50.00	UG	-0.03
50) Chlorobenzene-d5	15.88	117	1087576	50.00	UG	-0.02

System Monitoring Compounds

30) 1,2-Dichloroethane-d4	10.19	65	314976	21.01	UG	-0.02
Spiked Amount	50.000	Range	36 - 115	Recovery	=	42.02%
41) Toluene-d8	13.40	98	551061	24.36	UG	-0.02
Spiked Amount	50.000	Range	35 - 117	Recovery	=	48.72%
59) Bromofluorobenzene	17.96	95	352943	18.39	UG	0.00
Spiked Amount	50.000	Range	47 - 115	Recovery	=	36.78%#

Target Compounds

54) m,p-Xylene	16.32	106	14795	1.62	UG	Qvalue 88
55) o-Xylene	17.03	106	10447m	1.14	UG	

(#) = qualifier out of range (m) = manual integration

I1953.D IME1010.M

Wed Oct 17 14:31:35 2001

MSD_I

Page 1

0029

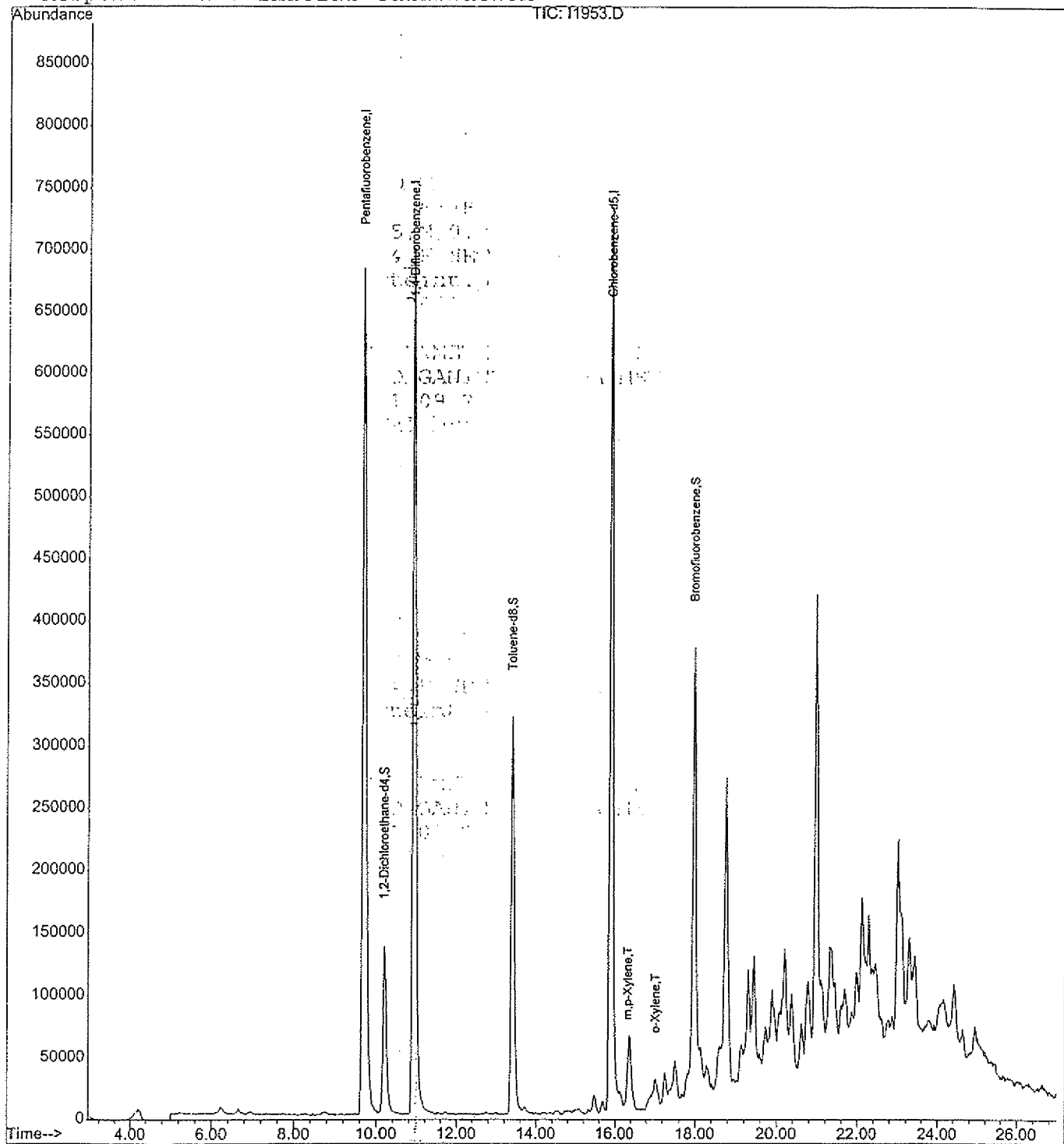
Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D
 Acq On : 17 Oct 2001 6:48
 Sample : 6-S2,7040-005,M,0.019g
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01,
 MS Integration Params: rteint.p
 Quant Time: Oct 17 14:31 2001

Vial: 46
 Operator: DP
 Inst : 5970 - In
 Multiplr: 1.00

Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Last Update : Thu Oct 11 09:23:20 2001
 Response via : Initial Calibration



LSC Area Percent Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D Vial: 46
 Acq On : 17 Oct 2001 6:48 Operator: DP
 Sample : 6-S2,7040-005,M,0.019g Inst : 5970 - In
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
 MS Integration Params: LSCINT.P

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Smoothing : ON Filtering: 5
 Sampling : 1 Min Area: 3 % of largest Peak
 Start Thrs: 0.2 Max Peaks: 100
 Stop Thrs : 0.15 Peak Location: TOP

If leading or trailing edge < 100 prefer < Baseline drop else tangent >
 Peak separation: 5

Signal : TIC

peak #	R.T. min	first scan	max scan	last scan	PK TY	peak height	corr. area	corr. % max.	% of total
1	9.698	438	446	463	rBV	681776	4339268	100.00%	16.185%
2	10.188	471	478	491	rBV	134567	823820	18.99%	3.073%
3	10.926	518	526	542	rBV	697178	4186998	96.49%	15.617%
4	13.400	679	687	699	rBV2	319109	1782140	41.07%	6.647%
5	15.466	815	821	830	rVB5	14520	87314	2.01%	0.326%
6	15.697	830	836	842	rBV3	9980	52320	1.21%	0.195%
7	15.881	842	848	860	rBV	728690	4075980	93.93%	15.203%
8	16.367	872	880	894	rVB3	60290	445524	10.27%	1.662%
9	16.922	907	916	917	rBV6	13230	77089	1.78%	0.288%
10	17.029	920	923	931	rVB6	20578	114776	2.65%	0.428%
11	17.243	933	937	943	rBV3	25195	130999	3.02%	0.489%
12	17.488	947	953	963	rVB5	31120	201165	4.64%	0.750%
13	17.810	968	974	976	rBV6	19414	97677	2.25%	0.364%
14	17.962	976	984	993	rBV	342767	1908436	43.98%	7.118%
15	18.613	1019	1027	1029	rBV3	33394	207365	4.78%	0.773%
16	18.764	1031	1037	1047	rVB	245392	1434870	33.07%	5.352%
17	19.142	1057	1062	1065	rBV5	30777	174136	4.01%	0.650%
18	19.307	1068	1073	1077	rVV	82436	431277	9.94%	1.609%
19	19.441	1077	1082	1091	rVB2	84813	487951	11.25%	1.820%
20	19.740	1096	1102	1105	rBV7	30911	162173	3.74%	0.605%
21	19.890	1108	1112	1120	rVB5	43467	233650	5.38%	0.872%
22	20.056	1120	1123	1125	rBV3	24393	86702	2.00%	0.323%
23	20.205	1129	1133	1140	rVB3	69543	329940	7.60%	1.231%
24	20.372	1140	1144	1154	rVB4	60817	346971	8.00%	1.294%
25	20.615	1154	1160	1164	rBV3	37210	198870	4.58%	0.742%
26	20.766	1164	1170	1177	rBV6	58921	399895	9.22%	1.492%
27	20.962	1177	1183	1190	rBV	356337	1805487	41.61%	6.734%
28	21.307	1201	1206	1207	rBV3	60930	190535	4.39%	0.711%
29	21.607	1221	1226	1227	rBV4	23906	89607	2.07%	0.334%
30	21.965	1247	1250	1255	rBV4	36153	171851	3.96%	0.641%
31	22.099	1255	1259	1266	rVV3	84872	482775	11.13%	1.801%

0031

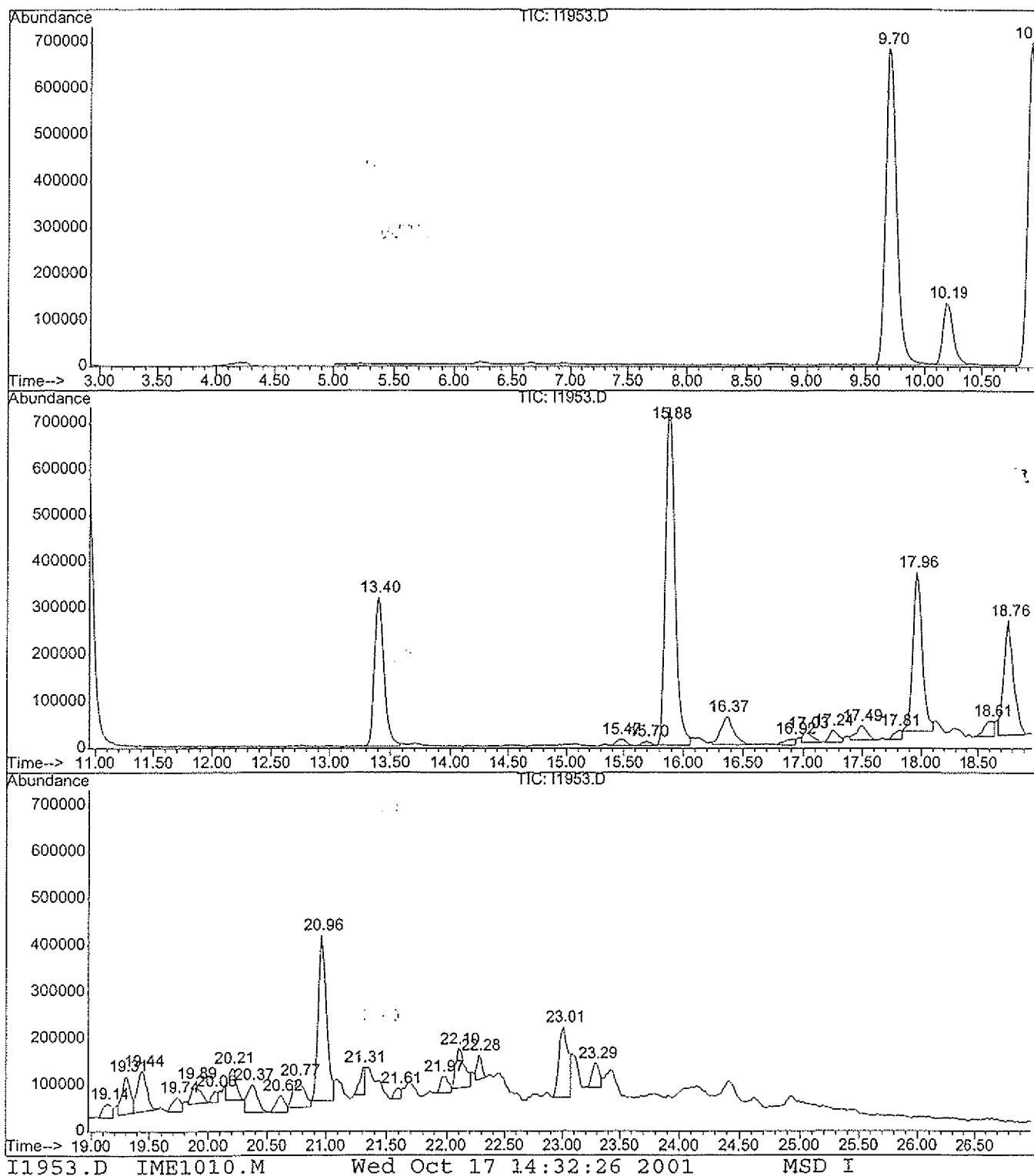
32	22.278	1269	1271	1275	rVB	49273	146348	3.37%	0.546%
33	23.009	1314	1320	1324	rBV	152892	858336	19.78%	3.202%
34	23.293	1334	1339	1342	rBV3	53554	247812	5.71%	0.924%

Sum of corrected areas: 26810057

I1953.D IME1010.M Wed Oct 17 14:32:25 2001 MSD_I

LSC Report - Integrated Chromatogram

File : C:\HPCHEM\1\DATA\10-16-01\I1953.D
 Operator : DP
 Acquired : 17 Oct 2001 6:48 using AcqMethod VOA1
 Instrument : 5970 - In
 Sample Name: 6-S2,7040-005,M,0.019g
 Misc Info : BRENNAN/99704_FINETEX,10/11/01,10/11/01,2
 Vial Number: 46
 Quant File :IME1010.RES (RTE Integrator)



I1953.D IME1010.M Wed Oct 17 14:32:26 2001 MSD_I

0033

Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D

Vial: 46

Acq On : 17 Oct 2001 6:48

Operator: DP

Sample : 6-S2,7040-005,M,0.019g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

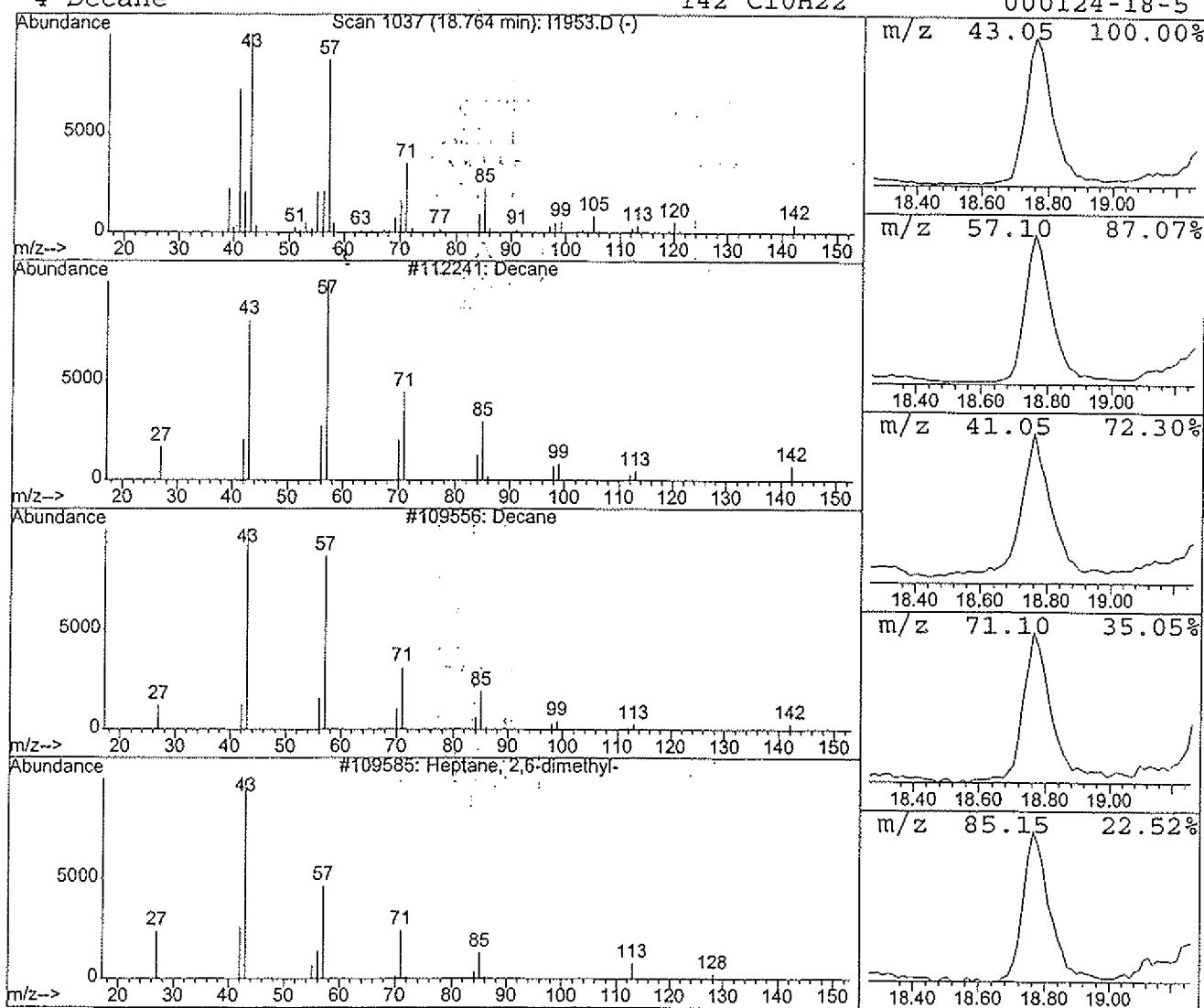
Library : C:\DATABASE\NIST98.L

Peak Number 1 Unknown alkane

Concentration Rank 2

R.T.	EstConc	Area	Relative to ISTD	R.T.
18.76	17.60 UG	1434870	Chlorobenzene-d5	15.88

Hit#	of	Tentative ID	MW	MolForm	CAS#	Qual
1	5	Decane	142	C10H22	000124-18-5	95
2		Decane	142	C10H22	000124-18-5	87
3		Heptane, 2,6-dimethyl-	128	C9H20	001072-05-5	80
4		Decane	142	C10H22	000124-18-5	74



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D

Vial: 46

Acq On : 17 Oct 2001 6:48

Operator: DP

Sample : 6-S2,7040-005,M,0.019g

Inst : 5970 - In

Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

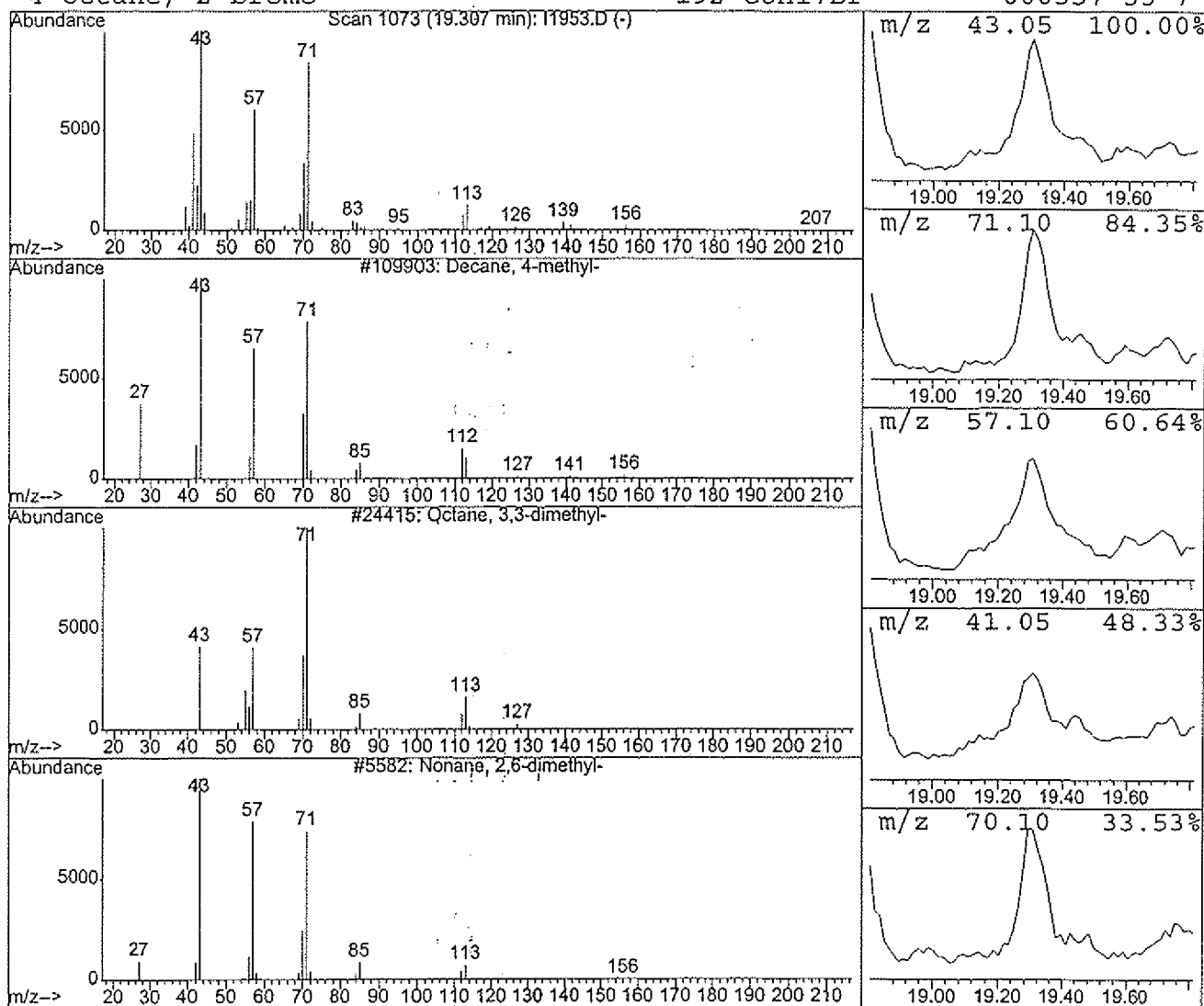
Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 2 Unknown alkane Concentration Rank 6

R.T.	EstConc	Area	Relative to ISTD	R.T.
19.31	5.29 UG	431277	Chlorobenzene-d5	15.88

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1			Decane, 4-methyl-	156	C11H24	002847-72-5	90
2			Octane, 3,3-dimethyl-	142	C10H22	004110-44-5	72
3			Nonane, 2,6-dimethyl-	156	C11H24	017302-28-2	72
4			Octane, 2-bromo-	192	C8H17Br	000557-35-7	59



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D

Vial: 46

Acq On : 17 Oct 2001 6:48

Operator: DP

Sample : 6-S2,7040-005,M,0.019g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

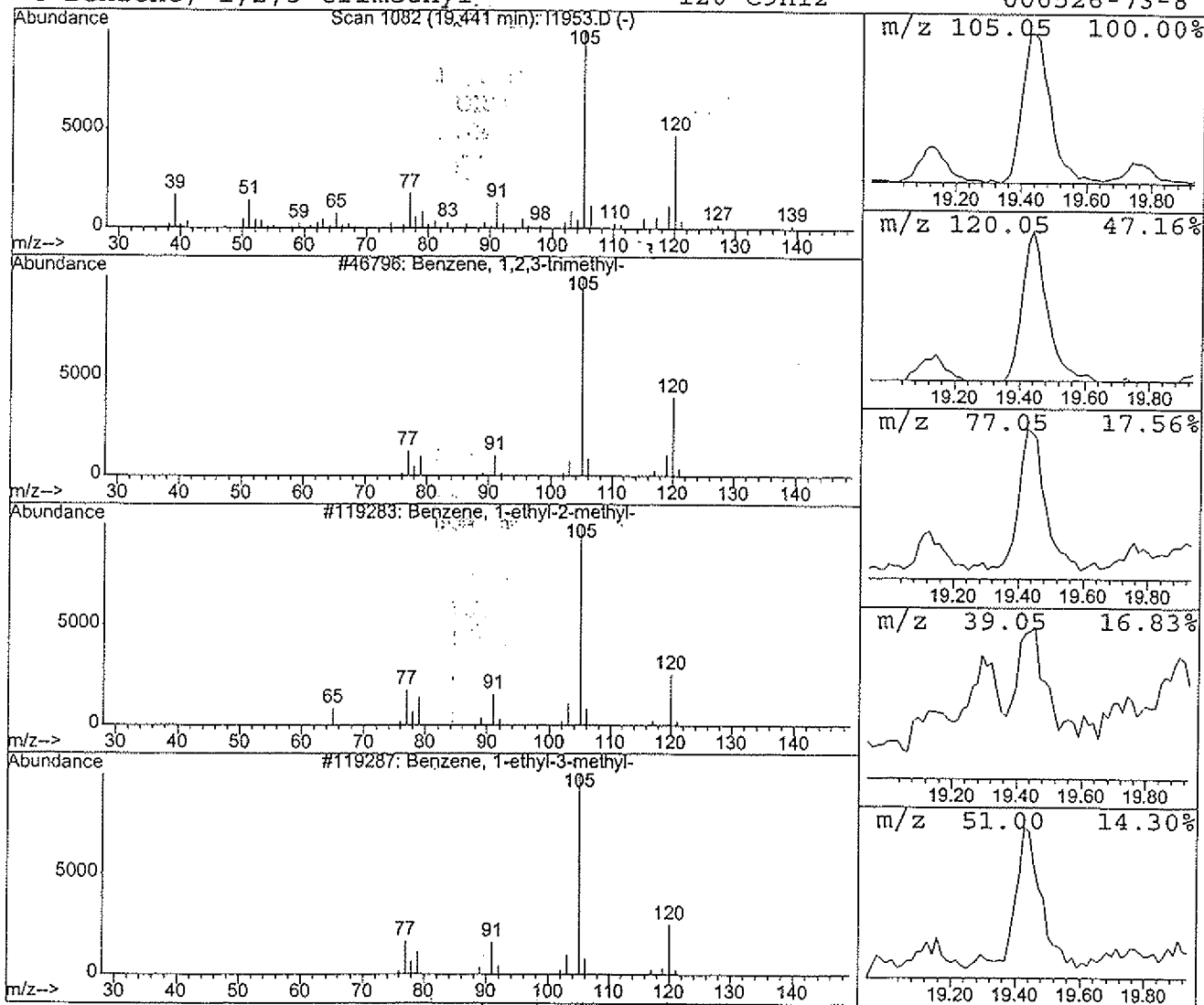
Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 3 Substituted benzene Concentration Rank 4

R.T.	EstConc	Area	Relative to ISTD	R.T.
19.44	5.99 UG	487951	Chlorobenzene-d5	15.88

Hit#	of	Tentative ID	MW	MolForm	CAS#	Qual
1	5	Benzene, 1,2,3-trimethyl-	120	C9H12	000526-73-8	94
2		Benzene, 1-ethyl-2-methyl-	120	C9H12	000611-14-3	94
3		Benzene, 1-ethyl-3-methyl-	120	C9H12	000620-14-4	94
4		Benzene, 1,2,3-trimethyl-	120	C9H12	000526-73-8	93



Library Search Compound Report

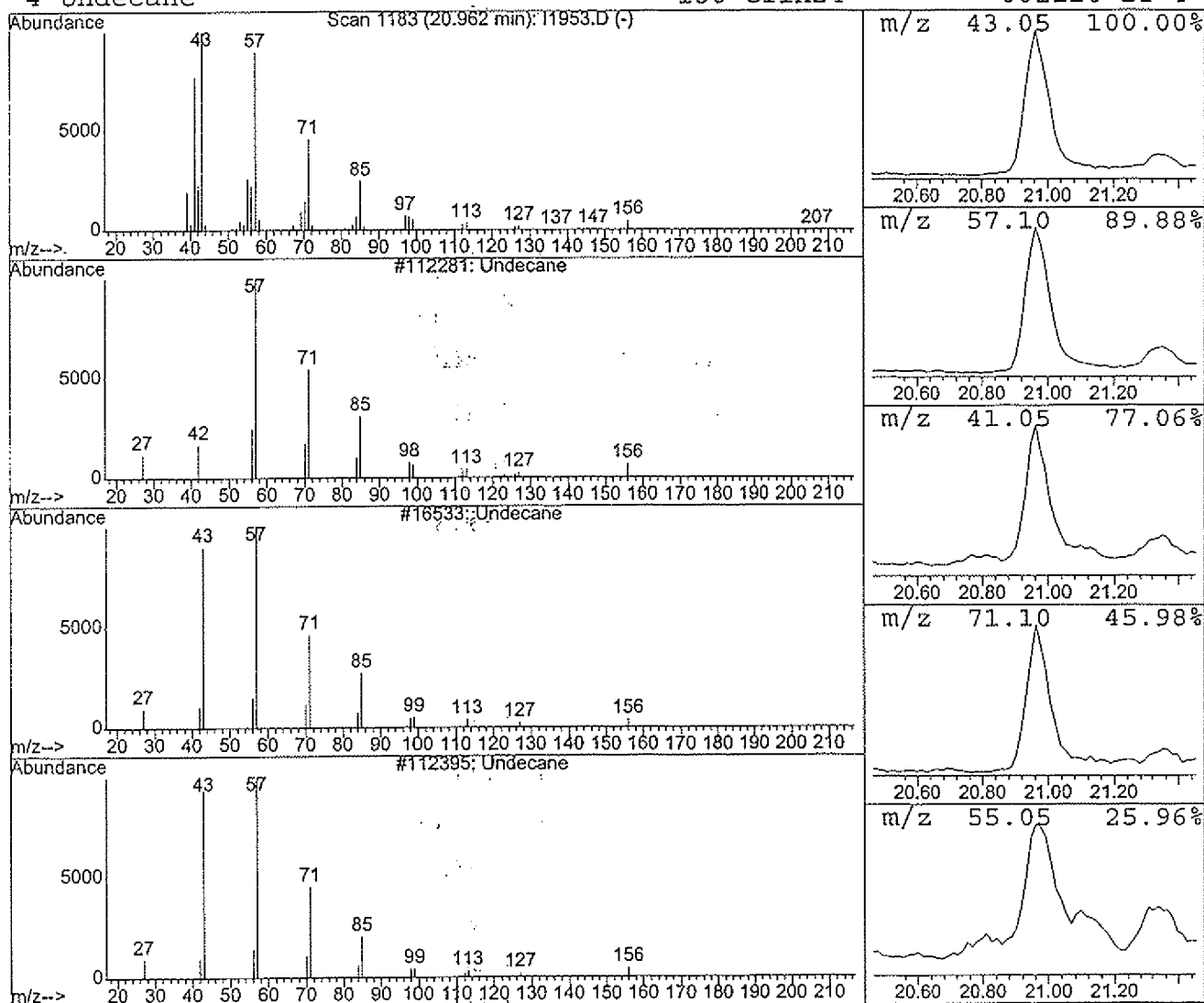
Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D
Acq On : 17 Oct 2001 6:48
Sample : 6-S2,7040-005,M,0.019g
Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00
MS Integration Params: LSCINT.P

Vial: 46
Operator: DP
Inst : 5970 - In

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Library : C:\DATABASE\NIST98.L

Peak Number 4 Undecane Concentration Rank 1

R.T.	EstConc	Area	Relative to ISTD	R.T.	
20.96	22.15 UG	1805490	Chlorobenzene-d5	15.88	
Hit# of 5	Tentative ID	MW	MolForm	CAS#	Qual
1	Undecane	156	C11H24	001120-21-4	94
2	Undecane	156	C11H24	001120-21-4	93
3	Undecane	156	C11H24	001120-21-4	81
4	Undecane	156	C11H24	001120-21-4	81



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D

Vial: 46

Acq On : 17 Oct 2001 6:48

Operator: DP

Sample : 6-S2,7040-005,M,0.019g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00

MS Integration Params: LSCINT.P

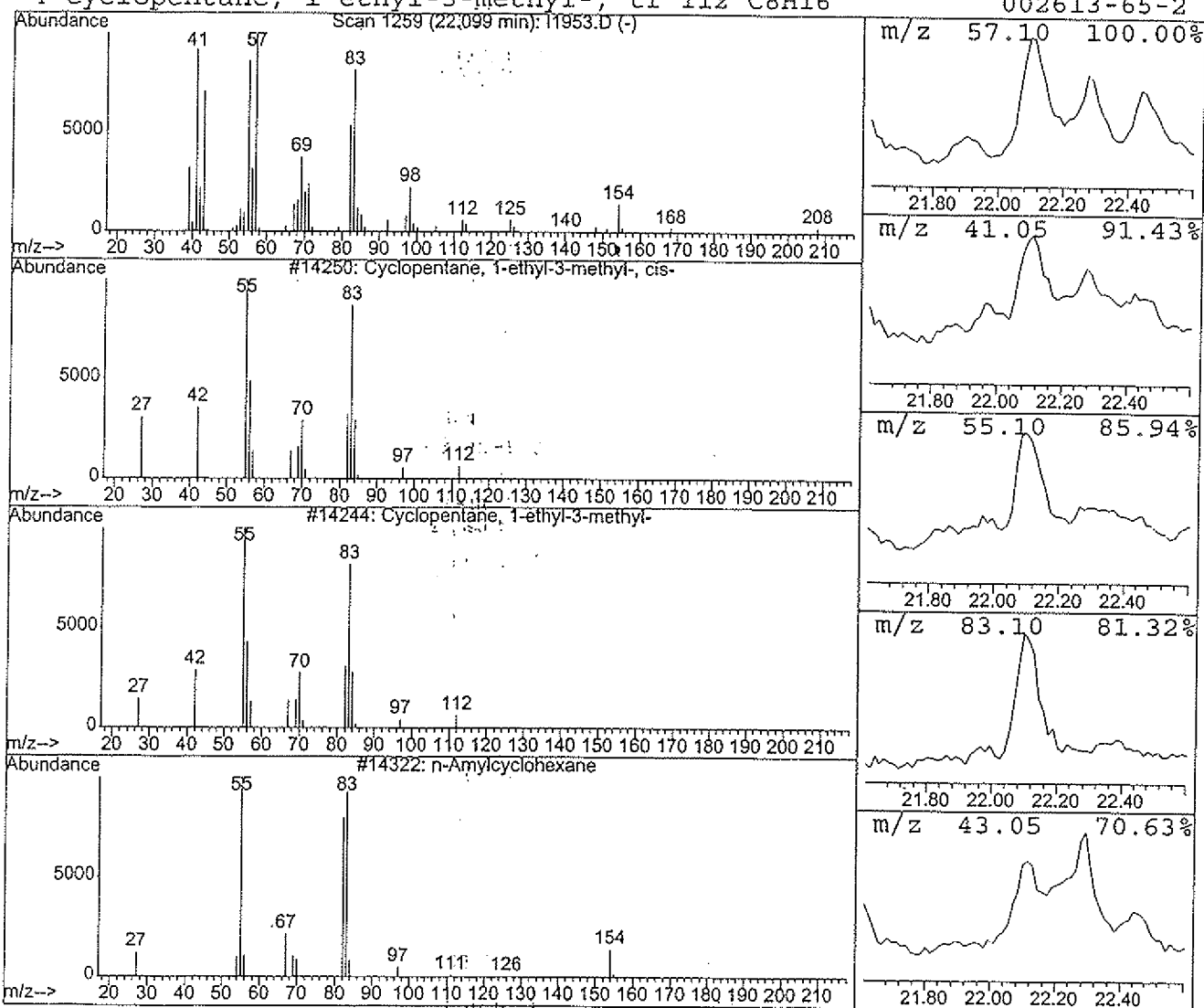
Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 5 Unknown cyclic hydrocarbon Concentration Rank 5

R.T.	EstConc	Area	Relative to ISTD	R.T.	
22.10	5.92 UG	482775	Chlorobenzene-d5	15.88	
Hit# of 5	Tentative ID	MW	MolForm	CAS#	Qual
1	Cyclopentane, 1-ethyl-3-methyl-, ci	112	C8H16	002613-66-3	52
2	Cyclopentane, 1-ethyl-3-methyl-	112	C8H16	003726-47-4	52
3	n-Amylcyclohexane	154	C11H22	029949-27-7	49
4	Cyclopentane, 1-ethyl-3-methyl-, tr	112	C8H16	002613-65-2	49



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1953.D

Vial: 46

Acq On : 17 Oct 2001 6:48

Operator: DP

Sample : 6-S2,7040-005,M,0.019g

Inst : 5970 - In

Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

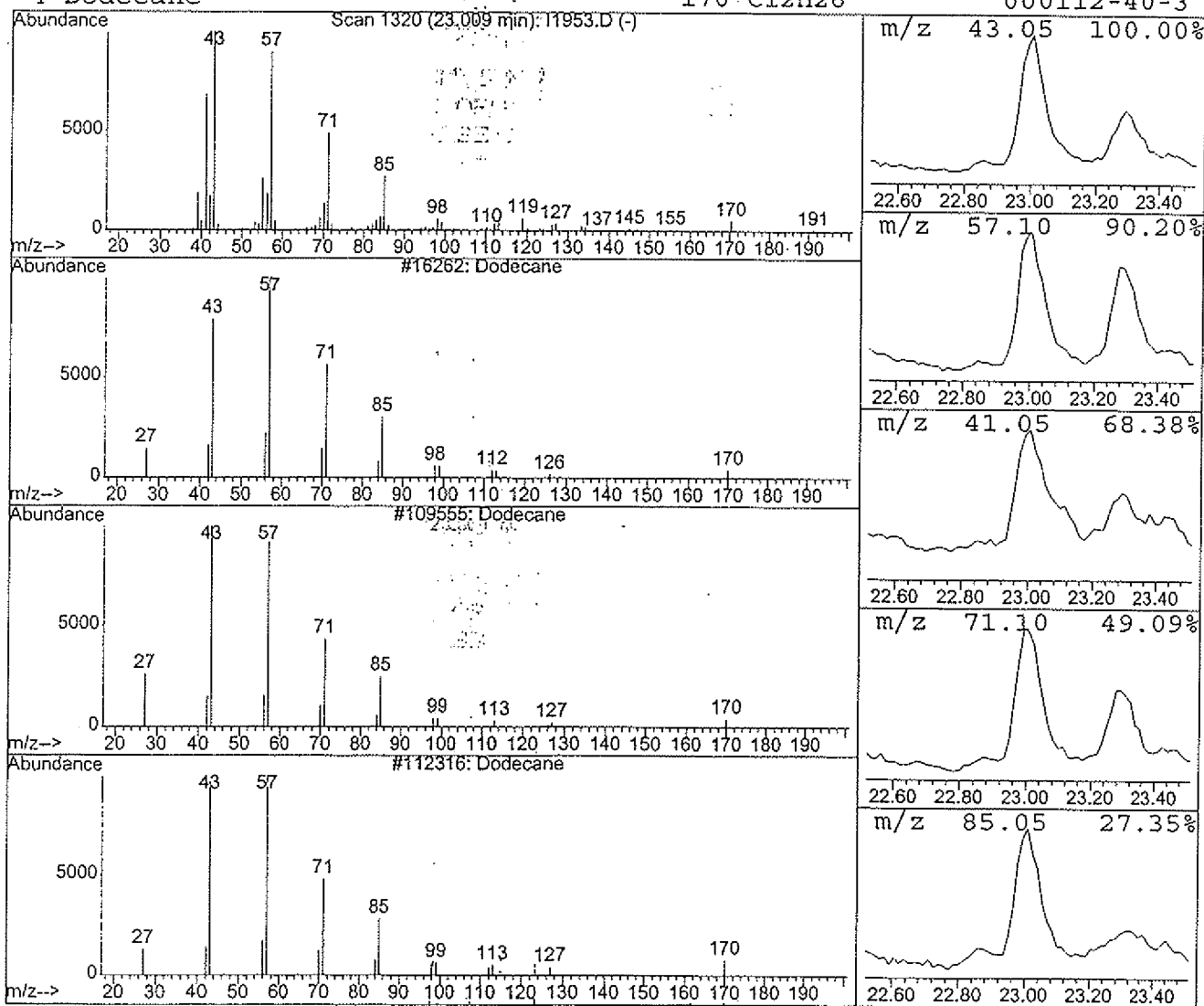
Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 6 Dodecane Concentration Rank 3

R.T.	EstConc	Area	Relative to ISTD	R.T.
23.01	10.53 UG	858336	Chlorobenzene-d5	15.88

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1	Dodecane			170	C12H26	000112-40-3	94
2	Dodecane			170	C12H26	000112-40-3	94
3	Dodecane			170	C12H26	000112-40-3	93
4	Dodecane			170	C12H26	000112-40-3	91



Data File : C:\HPCHEM\1\DATA\10-16-01\I1950.D Vial: 33
Acq On : 17 Oct 2001 5:09 Operator: DP
Sample : TB,7040-007,M,0.040g Inst : 5970 - In
Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01 Multiplr: 1.00
MS Integration Params: rteint.p
Quant Time: Oct 17 14:30 2001 Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Last Update : Thu Oct 11 09:23:20 2001
Response via : Initial Calibration
DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) Pentafluorobenzene	9.70	168	1063650	50.00	UG	-0.04
31) 1,4-Difluorobenzene	10.92	114	1430560	50.00	UG	-0.03
50) Chlorobenzene-d5	15.87	117	1160991	50.00	UG	-0.02

System Monitoring Compounds

30) 1,2-Dichloroethane-d4	10.19	65	1143712	85.09	UG	-0.02
Spiked Amount	50.000	Range	36 - 115	Recovery	=	170.18%#
41) Toluene-d8	13.40	98	2182417	97.34	UG	-0.02
Spiked Amount	50.000	Range	35 - 117	Recovery	=	194.68%#
59) Bromofluorobenzene	17.95	95	1572944	76.78	UG	0.00
Spiked Amount	50.000	Range	47 - 115	Recovery	=	153.56%#

Target Compounds

Qvalue

(#) = qualifier out of range (m) = manual integration

I1950.D IME1010.M

Wed Oct 17 14:30:42 2001

MSD_I

Page 1

0040

Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1950.D

Vial: 33

Acq On : 17 Oct 2001 5:09

Operator: DP

Sample : TB,7040-007,M,0.040g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01

Multiplr: 1.00

MS Integration Params: rteint.p

Quant Time: Oct 17 14:30 2001

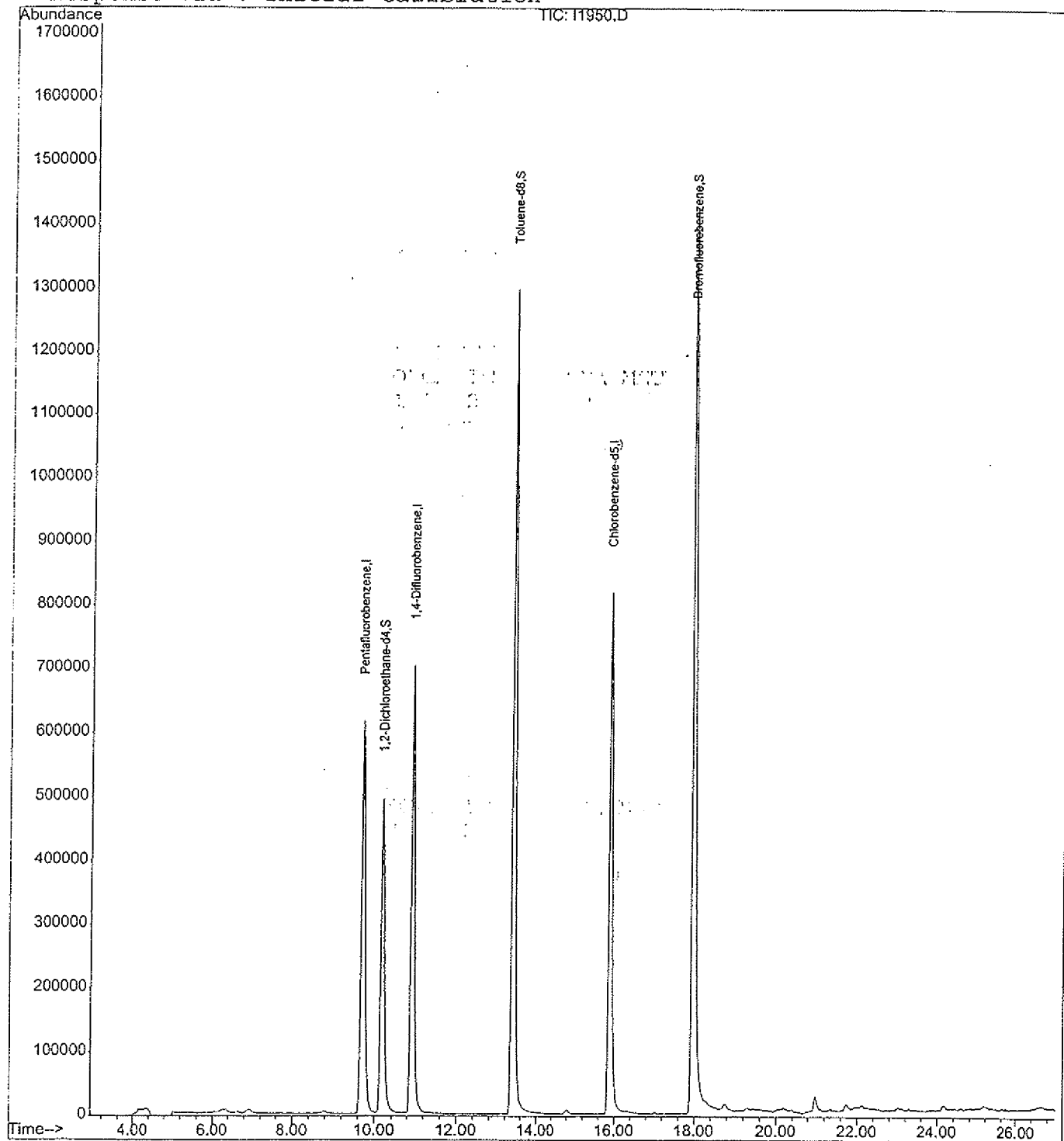
Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Initial Calibration



Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D

Vial: 31

Acq On : 17 Oct 2001 4:02

Operator: DP

Sample : N/A, METHOD_BLK, S, 0.1g, 0.0

Inst : 5970 - In

Misc : N/A, N/A, N/A

Multiplr: 1.00

MS Integration Params: rteint.p

Quant Time: Oct 18 13:36 2001

Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Initial Calibration

DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev (Min)
1) Pentafluorobenzene	9.71	168	1047735	50.00	UG	-0.02
31) 1,4-Difluorobenzene	10.92	114	1387504	50.00	UG	-0.03
50) Chlorobenzene-d5	15.87	117	1182634	50.00	UG	-0.02

System Monitoring Compounds

30) 1,2-Dichloroethane-d4	10.20	65	533676	40.31	UG	0.00
Spiked Amount	50.000	Range	36 - 115	Recovery	=	80.62%
41) Toluene-d8	13.40	98	1090226	50.14	UG	-0.02
Spiked Amount	50.000	Range	35 - 117	Recovery	=	100.28%
59) Bromofluorobenzene	17.96	95	944331	45.25	UG	0.00
Spiked Amount	50.000	Range	47 - 115	Recovery	=	90.50%

Target Compounds

Qvalue

(#) = qualifier out of range (m) = manual integration

I1948.D IME1010.M

Thu Oct 18 13:36:49 2001

MSD_I

Page 1

0042

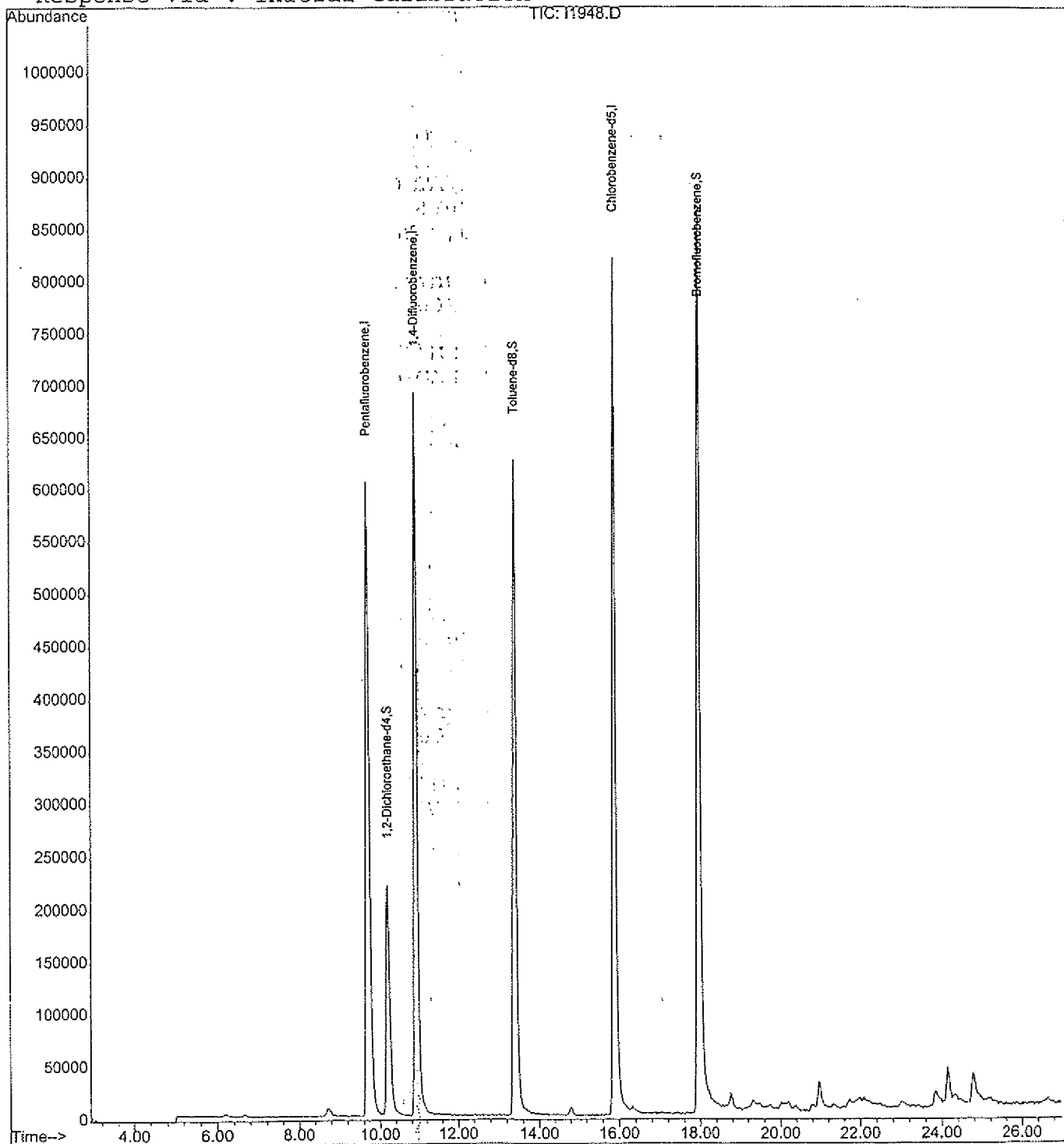
Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
 Acq On : 17 Oct 2001 4:02
 Sample : N/A,METHOD BLK,S,0.1g,0.0
 Misc : N/A,N/A,N/A
 MS Integration Params: rteint.p
 Quant Time: Oct 18 13:36 2001

Vial: 31
 Operator: DP
 Inst : 5970 - In
 Multiplr: 1.00

Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Last Update : Thu Oct 11 09:23:20 2001
 Response via : Initial Calibration



LSC Area Percent Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
Acq On : 17 Oct 2001 4:02
Sample : N/A,METHOD BLK,S,0.1g,0.0
Misc : N/A,N/A,N/A
MS Integration Params: LSCINT.P

Vial: 31
Operator: DP
Inst : 5970 - In
Multiplr: 1.00

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Smoothing : OFF Filtering: 5
Sampling : 1 Min Area: 1 % of largest Peak
Start Thrs: 0.2 Max Peaks: 100
Stop Thrs : 0 Peak Location: TOP

If leading or trailing edge < 100 prefer < Baseline drop else tangent >
Peak separation: 5

Signal : TIC

peak #	R.T. min	first scan	max scan	last scan	PK TY	peak height	corr. area	corr. % max.	% of total
1	9.711	438	447	462	rBV	604591	3830995	75.35%	16.546%
2	10.201	471	479	491	rBV	218025	1363833	26.82%	5.890%
3	10.923	518	526	550	rBV	689956	4065116	79.95%	17.557%
4	13.399	679	687	700	rBV	625464	3560689	70.03%	15.379%
5	15.871	841	848	865	rBV	818788	4556628	89.62%	19.680%
6	17.958	977	984	1000	rBV	866079	5084454	100.00%	21.960%
7	18.751	1032	1036	1041	rVB5	12835	53258	1.05%	0.230%
8	20.969	1174	1180	1189	rVV3	25691	165419	3.25%	0.714%
9	23.851	1361	1367	1373	rBV8	14062	101750	2.00%	0.439%
10	24.143	1380	1386	1393	rBV10	32926	173906	3.42%	0.751%
11	24.769	1420	1427	1434	rBV4	28846	197383	3.88%	0.853%

Sum of corrected areas: 23153431

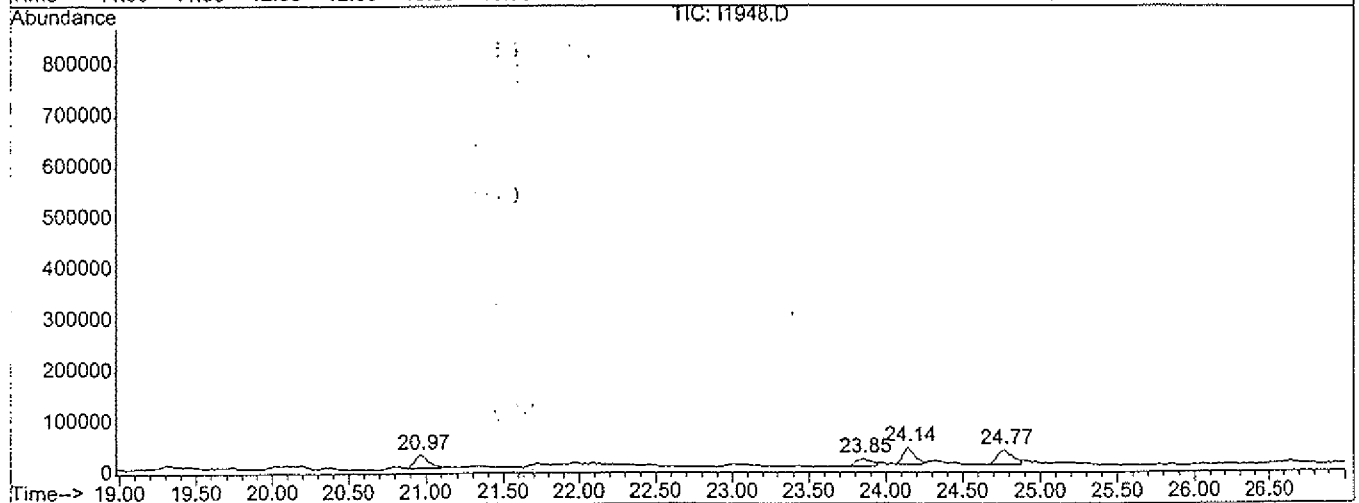
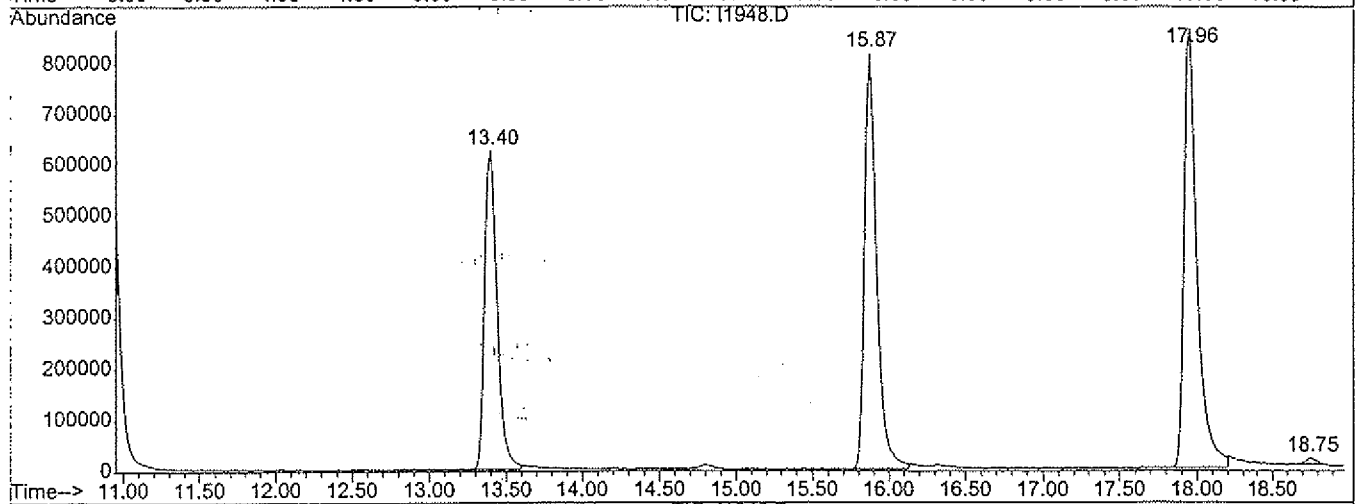
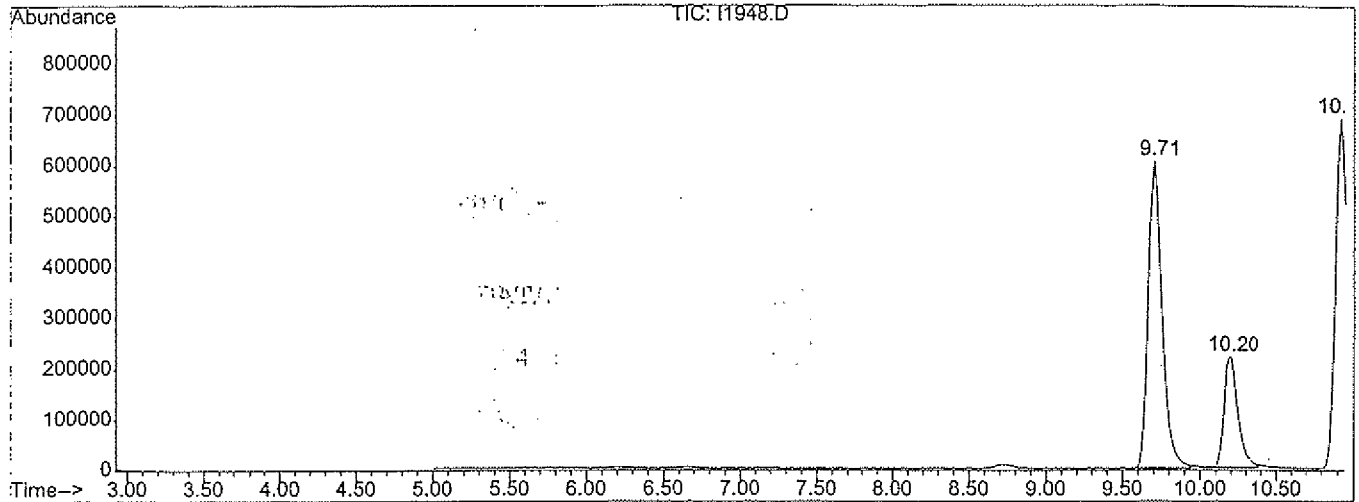
I1948.D IME1010.M

Thu Oct 18 13:36:56 2001 MSD_I

0044

LSC Report - Integrated Chromatogram

File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
 Operator : DP
 Acquired : 17 Oct 2001 4:02 using AcqMethod VOA1
 Instrument : 5970 - In
 Sample Name: N/A, METHOD BLK, S, 0.1g, 0.0
 Misc Info : N/A, N/A, N/A
 Vial Number: 31
 Quant File : IME1010.RES (RTE Integrator)



I1948.D IME1010.M Thu Oct 18 13:36:56 2001 MSD_I

0045

General Chemistry Quality Control

Matrix Soil
Units: S-mg/kg

Batch ID: AP040-0474

Blank

Parameter	Method Blank	MDL	Analysis Date
TPHC	ND	25.0	10/15/01

Duplicate Recovery

Parameter	QC Sample	Result	Duplicate Result	RPD	RPD Limits
TPHC	7048-001	162	164	1	20.0

Spike Recovery

Parameter	QC Sample	Result	Spike Added	Spike Result	%Spike Recovery	%Rec. Limits
TPHC	7048-001	162	533	664	94.2	75 - 125

The above blank results applies to the following samples E01

LCS-001-1015	7048-009
7048-001	7048-010
7048-001D	7048-011
7048-001S	7048-012
7048-002	7040-001
7048-003	7040-002
7048-004	7040-003
7048-005	7040-004
7048-006	7040-005
7048-007	7040-006
7048-008	7040-008
	7027-001

++ = No Flash - Sample boiled at 100C
NA = Not Applicable
ND = Not Detected
NC = Non calculable RPD due to value less than the detection limit
No = Does Not Ignite
Yes = Sample Ignites

©2000 IAL Inc.
All rights reserved

0046

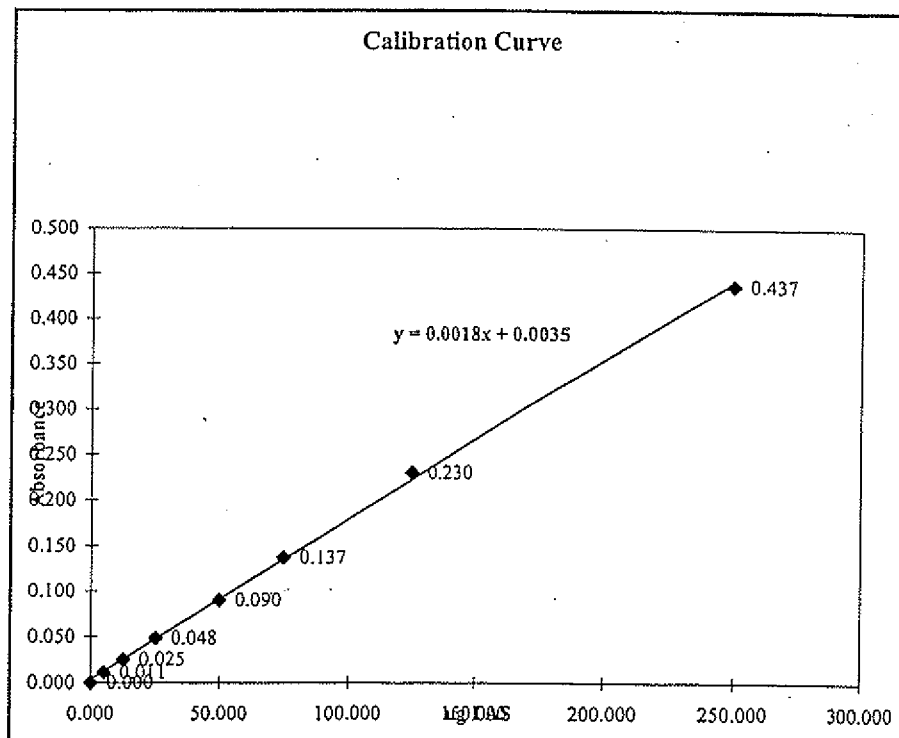
Integrated Analytical Laboratories

TPH

Date:	26-Jul-01
Method:	418.1
Matrix:	S/A
Analyst:	EDC

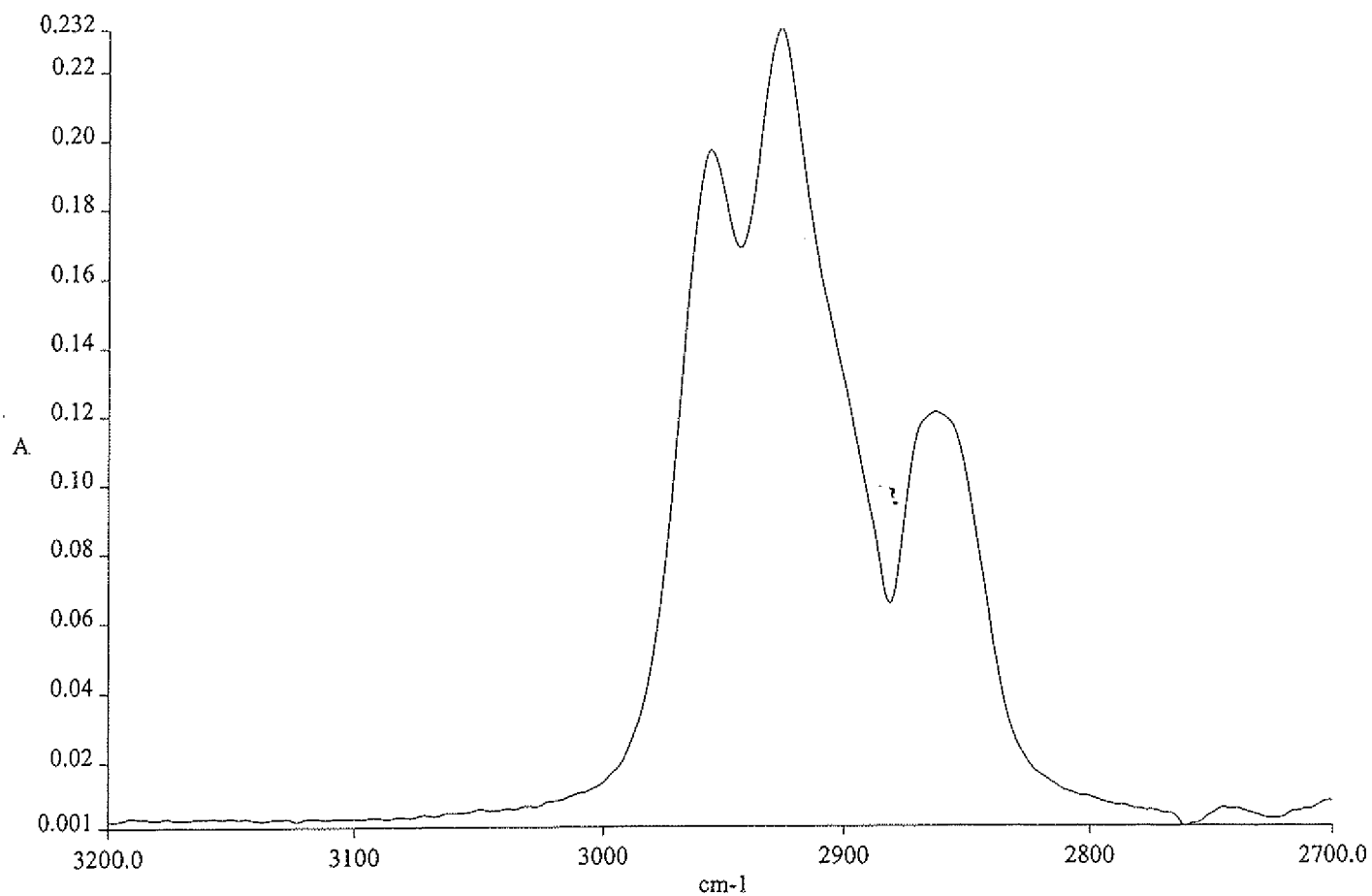
ug	Abs.
0.000	0.000
5.000	0.011
12.500	0.025
25.000	0.048
50.000	0.090
75.000	0.137
125.000	0.230
250.000	0.437

R ² =	0.999686985
Slope=	0.00175
Intercept=	0.00351



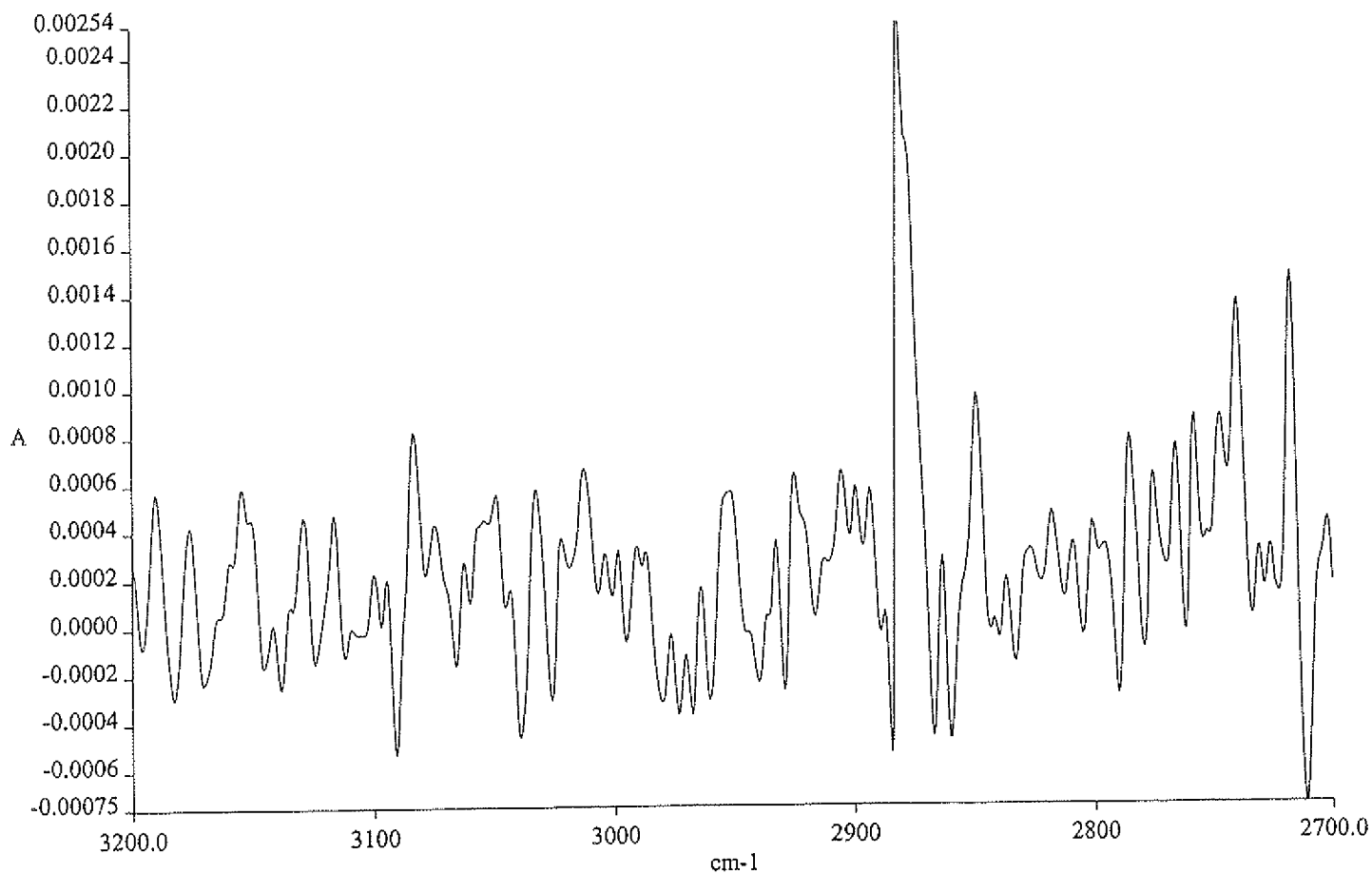
Comments:

0047



Monday, October 15, 2001; 09:32:41 ,
C:\PEL_DATA\SPECTRA\7027-001.SP, 2 scans, 4 cm-1
b-1015-1.sp

0048



Monday, October 15, 2001; 09:04:40 ,
C:\PEL_DATA\SPECTRA\BLANK\B-1015-1.SP, 2 scans, 4 cm-1

0049

INTEGRATED ANALYTICAL LABORATORIES CHAIN OF CUSTODY

CLIENT & PROJECT

Company Name:	Brennan Environmental (BEI)
Address:	8 Great Meadow Lane East Hanover, NJ
Telephone #:	(973) 781-1801
Fax #:	781-1804
Project Name:	99704 Firefly
Project Manager:	John Brennan
Reference ID#:	99704 PO#:

REPORTING

Fax to:	BEI
Fax #:	
Report to:	BEI
Address:	
Invoice to:	BEI
Address:	

Turnaround Time

Conditional/TPHC	48 hr	72 hr	1 wk	NA	Other:	Report Format
24 hr*	48 hr	72 hr	1 wk	NA	Other:	Results Only
Verbal/Fax						Reduced
24 hr*	48 hr*	72 hr*	1 wk*	2 wk	Other:	Regulatory
Hard Copy						SRP Disk** (db) or wkl
72 hr*	1 wk*	2 wk*	3 wk	Other:		Other:

*Prior to sample arrival, Lab notification is required.

ANALYTICAL PARAMETERS / PRESERVATIVES

ANALYTICAL PARAMETERS / PRESERVATIVES		** Circle format required												
123	456	123	456	123	456	123	456	123	456	123	456	123	456	Preservatives
1. HCL	3. BNO ₃													
2. NaOH	4. H ₂ SO ₄													
5. MeOH	6. Other													
COOLER TEMP. °C														
Comments														

SAMPLE INFORMATION

Sample ID	Sample Description	Date	Time	am	pm	Matrix		# of Containers	Lab ID
						W - Waste	SL - Sludge		
5-S1	Soil	10/11/01	9:58	✓		S		2	1
5-S2	Soil	10/11/01	12:49	✓		S		2	2
5-S3	Soil	10/11/01	12:23	✓		S		2	3
6-S1	Soil	10/11/01	10:58	✓		S		2	4
6-S2	Soil	10/11/01	11:08	✓		S		2	5
6-P1	Piping	10/11/01	9:08	✓		S		2	6
TB	trsp blank	10/11/01	9:00	✓		A		1	7
5-P1	piping	10/11/01	1:32	✓		S		1	8

Please print legibly and fill out completely. Samples cannot be processed and the turnaround time will not start until any ambiguities have been resolved.

CUSTODY LOG

Signature	Date	Time	Signature
Robert Dwyer	10/11/01	1800	James Lowell
James Lowell	10/11/01	1910	James Lowell
Relinquished by:			Received by:
Relinquished by:			Received by:
Relinquished by:			Received by:
Relinquished by:			Received by:
Relinquished by:			Received by:

Comments: * Conditional VO+ID analysis
on 25% of the highest
Samples with TPH results 5 min from 1,000 ppm

Lab Case #

70410

CHAIN OF CUSTODY

Case No.: E01-7040

P.O. #: _____

Project : 99704 FINETEX

Client/Project: BRENNAN/99704 FINETEX

Client Address:

Billing Address:

Brennan Environmental, Inc.

Brennan Environmental, Inc.

8 Great Meadow Lane

8 Great Meadow Lane

Unit D

Unit D

East Hanover, NJ 07936

East Hanover, NJ 07936

Date Received: 10/11/01

Verbal Due: Oct 26

Time Received: 19:10

Report Due: Nov 2

Report Format: Reduced

# of Containers	2	2	2	2	2	2
IAL ID #	7040-001	7040-002	7040-003	7040-004	7040-005	7040-006
Client ID #	5-S1	5-S2	5-S3	6-S1	6-S2	6-P1
Matrix	Soil	Soil	Soil	Soil	Soil	Soil
Sample Date	10/11/01	10/11/01	10/11/01	10/11/01	10/11/01	10/11/01
Sample Time	09:58	12:49	13:23	10:58	11:08	09:08
Methanol Kit	✓	✓	✓	✓	✓	✓
VO + 10, PP MP	C	C	C	C	✓	C
% Solids	✓	✓	✓	✓	✓	✓
TPHC-24hr TAT	✓	✓	✓	✓	✓	✓
Electronic Del.	✓					

Comments: NOTE 1: VO DEPENDENT ON TPHC.

REV 01: AS PER BOB DOOLEY 10/16/01, RUN SAMPLES #5 & FOR
VOA+10 AND SAMPLE #7 FOR PP VOA DUE 10/17/01. CANCEL
OTHERS ON HOLD. mrf

0051

CHAIN OF CUSTODY

Case No.: <u>E01-7040</u>	P.O. #: _____
Project : <u>99704 FINETEX</u>	
Client/Project: <u>BRENNAN/99704 FINETEX</u>	
Client Address:	Billing Address:
<u>Brennan Environmental, Inc.</u>	<u>Brennan Environmental, Inc.</u>
<u>8 Great Meadow Lane</u>	<u>8 Great Meadow Lane</u>
<u>Unit D</u>	<u>Unit D</u>
<u>East Hanover, NJ 07936</u>	<u>East Hanover, NJ 07936</u>
Date Received: <u>10/11/01</u>	Verbal Due: <u>Oct 26</u>
Time Received: <u>19:10</u>	Report Due: <u>Nov 2</u>
Report Format: <u>Reduced</u>	

# of Containers	1	1
IAL ID #	7040-007	7040-008
Client ID #	TB	5-P1
Matrix	Soil	Soil
Sample Date	10/11/01	10/11/01
Sample Time	:	13:32
Methanol Kit	✓	
VO MP, PP	✓	
% Solids		✓
TPHC-24hr TAT		✓

Comments: NOTE 1: VO DEPENDENT ON TPHC.
 REV 01: AS PER BOB DOOLEY 10/16/01, RUN SAMPLES #5 & FOR
 VOA+10 AND SAMPLE #7 FOR PP VOA DUE 10/17/01. CANCEL
 OTHERS ON HOLD. mrf

0052

INTEGRATED ANALYTICAL LABORATORIES, LLC

SAMPLE RECEIPT VERIFICATION

CASE NO:

7040

CLIENT:

BET

COOLER TEMPERATURE: 2° - 6°C: ☒

(See Chain of Custody)

CHAIN OF CUSTODY:

COMPLETE / INCOMPLETE

Comments: _____

Sample Bottles Intact:

☒

Comments: _____

Sample Labels Intact/ Correct:

☒

Sufficient Sample Volume:

☒

Correct bottles/ preservative:

☒

Samples received in

holding time/ prep time:

☒

Headspace/ bubbles in voa samples:

☒

Samples to be subcontracted:

☒

Preserved Sample pH checked:

(Excluding voa samples)

☒

KEY

☒ = YES

☒ = NO

☒ = N/A

ADDITIONAL COMMENTS: _____

SAMPLE(S) VERIFIED BY:

INITIAL

RC

DATE

10/11/01

CORRECTIVE ACTION REQUIRED:

YES

☐

(SEE BELOW)

NO

☐

CLIENT NOTIFIED:

YES

☐

Date/ Time: _____

NO

☐

PROJECT CONTACT: _____

SUBCONTRACTED LAB: _____

DATE SHIPPED: _____

ADDITIONAL COMMENTS: _____

VERIFIED/TAKEN BY:

INITIAL

DM

DATE

Integrated Analytical Laboratories, LLC.

Laboratory Custody ChronicleCase No : E01-7040Client : Brennan Environmental, Inc.Project : 99704 FINETEX

	GC/MS V		EXTRACT			ANALYSIS		
			DATE	TIME	INITIAL	DATE	TIME	INITIAL
Methanol Kit	7040-001	S				10/12		DP
	7040-002	S						
	7040-003	S						
	7040-004	S						
	7040-005	S						
	7040-006	S						
	7040-007	S						
VO + 10, PP MP	7040-005	S						
	7040-007	S						
	7040	S						
	7040	S						
	7040	S						
	7040	S						
VO MP, PP	7040	S						

WETCHEM								
% Solids	7040-001	S	10/12		KC			
	7040-002	S						
	7040-003	S						
	7040-004	S						
	7040-005	S						
	7040-006	S						
	7040-008	S						
TPHC-24hr TAT	7040-001	S	10/12	1430	DM	10/15	901	GM
	7040-002	S						
	7040-003	S						
	7040-004	S						
	7040-005	S						
	7040-006	S						
	7040-008	S						

AIR								
Electronic Del.	7040-001	S						

 REVIEW & APPROVAL: Andrew
 REMARKS :

0054



Integrated Analytical Laboratories, LLC.

273 Franklin Road
Randolph, N.J. 07869

Phone: 973 361-4252
Fax: 973 989-5288

ANALYTICAL DATA REPORT

for

Brennan Environmental, Inc.
8 Great Meadow Lane
Unit D
East Hanover, NJ 07936

Project: 99704 FINETEX

Lab Case Number: E01-7041

Date Report Prepared: October 26, 2001

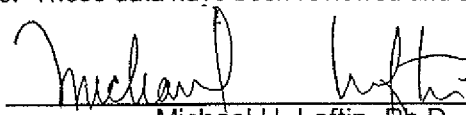
CLIENT
SAMPLE ID

5-F1

LABORATORY
SAMPLE ID

7041-001

All required protocols were followed during analyses. These data have been reviewed and accepted by



Michael H. Leftin, Ph.D.
Laboratory Director

The liability of Integrated Analytical Laboratories, LLC. is limited to the actual cost of the analyses performed.



IAL is a NELAC Certified Lab # 14751 and maintains certification in Connecticut # PH-0699, New York # 11402 and Florida # E87670

TIERRA-B-010346

INTEGRATED ANALYTICAL LABORATORIES, LLC.

TABLE OF CONTENTS

	<u>Page</u>
Qualifiers	1
Conformance / NonConformance Summary	2
Laboratory Deliverables Check List	3
GC/MS NonConformance Summary	4
GC NonConformance Summary	5
Metal NonConformance Summary	6
Summary Report	7
Analytical Results	
Volatiles + 15	10
Semivolatiles - BNA + 25	12
PCB's	15
Pesticides	16
PP Metals	17
General Analytical	
Total Recoverable Phenols	18
Cyanide, Total	19
Methodology Summary	20
Quality Control	
Volatiles + 15	21
Tuning Results Summary	
Method Blank Results Summary	
Calibration Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Internal Standard Summary	
Chromatograms	
Semivolatiles - BNA + 25	65
Tuning Results Summary	
Method Blank Results Summary	
Calibration Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Internal Standard Summary	
Chromatograms	
PCB's	95
Method Blank Results Summary	
Standards Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Retention Time Shift Summary	
Chromatograms	
Pesticides	115
Method Blank Results Summary	
Standards Summary	
Surrogate Compound Recovery Results Summary	
Matrix Spike/Matrix Spike Duplicate Results Summary	
Retention Time Shift Summary	
Chromatograms	
PP Metals	142
Method Blank Results Summary	

INTEGRATED ANALYTICAL LABORATORIES, LLC.

TABLE OF CONTENTS

	<u>Page</u>
Calibration Summary	
Spike Sample Results Summary	
Duplicate Sample Results Summary	
General Analytical	151
Blank Results Summary	
Spike Sample Results Summary	
Duplicate Sample Results Summary	
Sample Tracking	
Chains of Custody	153
Laboratory Chronicle	156

INTEGRATED ANALYTICAL LABORATORIES, LLC.

MATRIX QUALIFIERS

- A -** Indicates the sample is an Aqueous matrix.
- O -** Indicates the sample is an Oil matrix.
- S -** Indicates the sample is a Soil, Sludge or Sediment matrix.
- X -** Indicates the sample is an Other matrix as indicated by Client Chain of Custody.

DATA QUALIFIERS

- B -** Indicates the analyte was found in the Blank and in the sample. It indicates possible sample contamination and warns the data user to use caution when applying the results of the analyte.
- C -** Common Laboratory Contaminant.
- D -** The compound was reported from the Diluted analysis.
- D.F. -** Dilution Factor.
- E -** Estimated concentration, reported results are outside the calibrated range of the instrument.
- J -** Indicates an estimated value. The compound was detected at a value below the method detection limit but greater than zero. For GC/MS procedures, the mass spectral data meets the criteria required to identify the target compound.
- MDL -** Method Detection Limit.
- MI -** Indicates compound concentration could not be determined due to Matrix Interferences.
- NA -** Not Applicable.
- ND -** Indicates the compound was analyzed for but Not Detected at the MDL.

REPORT QUALIFIERS

All solid sample analyses are reported on a dry weight basis.

All solid sample values are corrected for original sample size and percent solids.

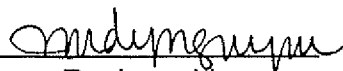
INTEGRATED ANALYTICAL LABORATORIES, LLC.

CONFORMANCE / NONCONFORMANCE SUMMARY

Integrated Analytical Laboratories, LLC. received one (1) soil sample(s) from Brennan Environmental, Inc. (Project: 99704 FINETEX) on October 11, 2001 for the analysis of:

- (1) Volatiles + 15
- (1) Semivolatiles - BNA + 25
- (1) PCB's
- (1) Pesticides
- (1) PP Metals
- (1) Cyanide, Total
- (1) Total Recoverable Phenols

A review of the QA/QC measures for the analysis of the sample(s) contained in this report has been performed by:


Reviewed by

10/26/01
Date

INTEGRATED ANALYTICAL LABORATORIES, LLC.

LABORATORY DELIVERABLES CHECK LIST

Lab Case Number: E01-7041

	Check If Complete
1. Cover Page, Title Page listing Lab Certification #, facility name & address and date of report preparation.	<u>✓</u>
2. Table of Contents.	<u>✓</u>
3. Summary Sheets listing analytical results for all targeted and non-targeted compounds.	<u>✓</u>
4. Summary Table cross-referencing Field ID's vs. Lab ID's.	<u>✓</u>
5. Document bound, paginated and legible.	<u>✓</u>
6. Chain of Custody.	<u>✓</u>
7. Methodology Summary.	<u>✓</u>
8. Laboratory Chronicle and Holding Time Check.	<u>✓</u>
9. Results submitted on a dry weight basis (if applicable).	<u>✓</u>
10. Method Detection Limits.	<u>✓</u>
11. Lab certified by NJDEP for parameters or appropriate category of parameters or a member of the USEPA CLP.	<u>✓</u>
12. NonConformance Summary.	<u>✓</u>


QC Reviewed by

10/26/01
Date

Lab Case Number: E01 - 7041

13. Comments:

Date _____

INTEGRATED ANALYTICAL LABORATORIES
CONFORMANCE/NONCONFORMANCE SUMMARY
GC/MS SEMIVOLATILE ANALYSIS

Lab Case Number: E01 - 7041

	<u>No</u>	<u>Yes</u>
1. Chromatograms Labeled/Compounds Identified (Field Samples and Method Blanks).	_____	✓
2. GC/MS Tuning Specifications:	_____	✓
a. DFTPP Passed	_____	✓
3. GC/MS Tuning Frequency - Performed every 24 hours for 600 series, 12 hours for 8000 series.	_____	✓
4. GC/MS Calibration - Initial calibration performed within 30 days before sample analysis and continuing calibration performed within 24 hours before sample analysis for 600 series.	_____	✓
5. GC/MS Calibration Requirements:		
a. Calibration Check Compounds	_____	✓
b. System Performance Check Compounds	_____	✓
6. Blank Contamination - If yes, list compounds and concentrations in each blank:	/	_____
a. B/N Fraction _____		
b. Acid Fraction _____		
7. Surrogate Recoveries Meet Criteria (If not met, list those compounds and their recoveries which fall outside the acceptable range)	_____	✓
a. B/N Fraction _____		
b. Acid Fraction _____		
If not met, were the calculations checked and the results qualified as "estimated"?	_____	na
8. Matrix Spike/Matrix Spike Duplicate meet criteria (if not, list those compounds and their recoveries/% differences which fall outside the acceptable range)	_____	✓
a. B/N Fraction _____		
b. Acid Fraction _____		
9. Internal Standard Area/Retention Time Shift meet criteria	_____	✓
10. Extraction Holding Time Met	_____	✓
If not met, list number of days exceeded for each sample:		

11. Analysis Holding Time Met	_____	✓
If not met, list number of days exceeded for each sample:		

12. Sample Dilution Performed	/	_____
High Target Compounds High Nontarget Compounds Matrix Interference Other		

13. Comments:

Organics Manager

10/16/01
Date

INTEGRATED ANALYTICAL LABORATORIES
CONFORMANCE/NONCONFORMANCE SUMMARY
GC ANALYSIS - PCB'S, PESTICIDES, HERBICIDES

Lab Case Number: E01 - 9041

	<u>No</u>	<u>Yes</u>
1. Chromatograms Labeled/Compounds Identified (Field Samples and Method Blanks).	<u> </u>	<u> ✓ </u>
2. Standards Summary submitted.	<u> </u>	<u> ✓ </u>
3. Calibration - Initial calibration performed within 30 days before sample analysis and continuing calibration performed within 12 hrs of the sample analysis.	<u> </u>	<u> ✓ </u>
4. Blank Contamination - If yes, list compounds and concentrations in each blank:	<u> ✓ </u>	<u> </u>
a. PCB's		
b. Pesticides		
c. Herbicides		
5. Surrogate Recoveries meet criteria (if applicable). If not met, list those compounds and their recoveries which fall outside the acceptable range:	<u> </u>	<u> ✓ </u>
a. PCB's		
b. Pesticides		
c. Herbicides		
6. Matrix Spike/Matrix Spike Duplicate meet criteria (if not, list those compounds and their recoveries/% differences which fall outside the acceptable range):	<u> </u>	<u> ✓ </u>
a. PCB's		
b. Pesticides		
c. Herbicides		
7. Retention Time Shift Meet Criteria (if applicable).	<u> </u>	<u> ✓ </u>
8. Extraction Holding Time Met. If not met, list number of days exceeded for each sample:	<u> </u>	<u> ✓ </u>

9. Analysis Holding Time Met. If not met, list number of days exceeded for each sample:	<u> </u>	<u> ✓ </u>

Comments:


Organic Manager

10.17. 2001
Date

METAL ANALYSIS CONFORMANCE/NONCONFORMANCE SUMMARY

INTEGRATED ANALYTICAL LABORATORIES, LLC.

SUMMARY REPORT
Client: Brennan Environmental, Inc.
Project: 99704 FINETEX
Lab Case No.: E01-7041

Lab ID:	7041-001
Client ID:	5-F1
Matrix:	Soil
Sampled Date:	10/11/2001
PARAMETER(Units)	Conc Q MDL
Volatiles (ppb)	
TOTAL VO's:	ND 690
TOTAL TIC's:	4430
TOTAL VO's & TIC's:	4430
Semivolatiles - BNA (ppb)	
TOTAL BNA'S:	ND 106
TOTAL TIC's:	ND
TOTAL BNA'S & TIC's:	ND
PCB's (ppb)	
Aroclor-1016	ND 154
Aroclor-1221	ND 154
Aroclor-1232	ND 154
Aroclor-1242	ND 154
Aroclor-1248	ND 154
Aroclor-1254	ND 154
Aroclor-1260	ND 154
Pesticides (ppb)	
alpha-BHC	ND 3.87
beta-BHC	ND 3.87
gamma-BHC	ND 3.87
delta-BHC	ND 3.87
Heptachlor	ND 3.87
Aldrin	ND 3.87
Heptachlor epoxide	ND 3.87
Endosulfan I	ND 3.87
4,4'-DDE	ND 3.87
Dieldrin	ND 3.87
Endrin	ND 3.87
Endosulfan II	ND 3.87
4,4'-DDD	ND 3.87
Endrin aldehyde	ND 3.87
Endosulfan sulfate	ND 3.87
4,4'-DDT	ND 3.87
Chlordane	ND 19.4
Toxaphene	ND 19.4

ND = Analyzed for but Not Detected at the MDL

All qualifiers on individual Volatiles & Semivolatiles are carried down through summation.

INTEGRATED ANALYTICAL LABORATORIES, LLC.

SUMMARY REPORT

Client: Brennan Environmental, Inc.

Project: 99704 FINETEX

Lab Case No.: E01-7041

	Lab ID:	7041-001		
	Client ID:	5-F1		
	Matrix:	Soil		
	Sampled Date:	10/11/2001		
PARAMETER(Units)		Conc	Q	MDL
Metals (ppm)				
Antimony		ND		1.09
Arsenic		1.70		1.09
Beryllium		ND		0.546
Cadmium		ND		0.273
Chromium		9.00		2.18
Copper		13.1		2.18
Lead		14.4		0.546
Mercury		0.0201		0.0138
Nickel		8.48		1.09
Selenium		ND		2.18
Silver		ND		0.546
Thallium		ND		0.109
Zinc		26.1		2.18
General Analytical				
Cyanide, Total (ppm)		ND		1.10
Total Recoverable Phenols (ppm)		ND		2.76

ND = Analyzed for but Not Detected at the MDL

All qualifiers on individual Volatiles & Semivolatiles are carried down through summation.

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Analyzed: 10/18/2001

Data file: I1989.D

GC/MS Column: DB-624

Sample wt/vol: 0.040g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.4

Compound	Concentration	Q	MDL
Chloromethane	ND		690
Vinyl Chloride	ND		690
Bromomethane	ND		690
Chloroethane	ND		690
Trichlorofluoromethane	ND		690
Acrolein	ND		1380
1,1-Dichloroethene	ND		690
Methylene Chloride	ND		690
Acrylonitrile	ND		1380
trans-1,2-Dichloroethene	ND		690
1,1-Dichloroethane	ND		690
Chloroform	ND		690
1,1,1-Trichloroethane	ND		690
Carbon Tetrachloride	ND		690
1,2-Dichloroethane(EDC)	ND		690
Benzene	ND		690
Trichloroethene	ND		690
1,2-Dichloropropane	ND		690
Bromodichloromethane	ND		690
2-Chloroethylvinyl Ether	ND		690
cis-1,3-Dichloropropene	ND		690
Toluene	ND		690
trans-1,3-Dichloropropene	ND		690
1,1,2-Trichloroethane	ND		690
Tetrachloroethene	ND		690
Dibromochloromethane	ND		690
Chlorobenzene	ND		690
Ethylbenzene	ND		690
Total Xylenes	ND		690
Bromoform	ND		690
1,1,2,2-Tetrachloroethane	ND		690
1,3-Dichlorobenzene	ND		690
1,4-Dichlorobenzene	ND		690
1,2-Dichlorobenzene	ND		690

Total Target Compounds: 0

0010

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS Tentatively Identified Compounds

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Analyzed: 10/18/2001

Date File: I1989.D

GC/MS Column: DB-624

Sample wt/vol: 0.040g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.4

CAS #	Compound	Estimated Concentration	Retention Time
	Unknown alkane	869	18.68
001120-21-4	Undecane	1120	20.87
	Unknown alkane	800	22.01
000112-40-3	Dodecane	924	22.90
	Unknown alkane	717	24.81

Total TICs = 4430

INTEGRATED ANALYTICAL LABORATORIES

SEMIVOLATILE ORGANICS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Extracted: 10/15/2001

Date Analyzed: 10/15/2001

Data file: B6861.D

GC/MS Column: DB-5

Sample wt/vol: 10.39g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.40

Compound	Concentration	Q	MDL
N-Nitrosodimethylamine	ND		106
Phenol	ND		106
Aniline	ND		106
bis(2-Chloroethyl)ether	ND		106
2-Chlorophenol	ND		106
1,3-Dichlorobenzene	ND		106
1,4-Dichlorobenzene	ND		106
Benzyl alcohol	ND		106
1,2-Dichlorobenzene	ND		106
2-Methylphenol	ND		106
bis(2-chloroisopropyl)ether	ND		106
4-Methylphenol	ND		106
N-Nitroso-di-n-propylamine	ND		106
Hexachloroethane	ND		106
Nitrobenzene	ND		106
Isophorone	ND		106
2-Nitrophenol	ND		106
2,4-Dimethylphenol	ND		106
bis(2-Chloroethoxy)methane	ND		106
Benzoic acid	ND		106
2,4-Dichlorophenol	ND		106
1,2,4-Trichlorobenzene	ND		106
Naphthalene	ND		106
4-Chloroaniline	ND		106
Hexachlorobutadiene	ND		106
4-Chloro-3-methylphenol	ND		106
2-Methylnaphthalene	ND		106
Hexachlorocyclopentadiene	ND		106
2,4,6-Trichlorophenol	ND		106
2,4,5-Trichlorophenol	ND		106
2-Chloronaphthalene	ND		106
2-Nitroaniline	ND		106
Dimethylphthalate	ND		106
2,6-Dinitrotoluene	ND		106
Acenaphthylene	ND		106

INTEGRATED ANALYTICAL LABORATORIES
SEMIVOLATILE ORGANICS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001
Client ID: 5-F1
Date Received: 10/11/2001
Date Extracted: 10/15/2001
Date Analyzed: 10/15/2001
Data file: B6861.D

GC/MS Column: DB-5
Sample wt/vol: 10.39g
Matrix-Units: Soil-µg/Kg (ppb)
Dilution Factor: 1
% Moisture: 9.40

Compound	Concentration	Q	MDL
3-Nitroaniline	ND		106
Acenaphthene	ND		106
2,4-Dinitrophenol	ND		106
4-Nitrophenol	ND		106
2,4-Dinitrotoluene	ND		106
Dibenzofuran	ND		106
Diethylphthalate	ND		106
Fluorene	ND		106
4-Chlorophenyl-phenylether	ND		106
4-Nitroaniline	ND		106
4,6-Dinitro-2-methylphenol	ND		106
N-Nitrosodiphenylamine	ND		106
1,2-Diphenylhydrazine/Azobenzene	ND		106
4-Bromophenyl-phenylether	ND		106
Hexachlorobenzene	ND		106
Pentachlorophenol	ND		106
Phenanthrene	ND		106
Anthracene	ND		106
Carbazole	ND		106
Di-n-butylphthalate	ND		106
Fluoranthene	ND		106
Benzidine	ND		106
Pyrene	ND		106
3,3'-Dimethylbenzidine	ND		106
Butylbenzylphthalate	ND		106
3,3'-Dichlorobenzidine	ND		106
Benzo[a]anthracene	ND		106
Chrysene	ND		106
bis(2-Ethylhexyl)phthalate	ND		106
Di-n-octylphthalate	ND		106
Benzo[b]fluoranthene	ND		106
Benzo[k]fluoranthene	ND		106
Benzo[a]pyrene	ND		106
Indeno[1,2,3-cd]pyrene	ND		106
Dibenz[a,h]anthracene	ND		106
Benzo[g,h,i]perylene	ND		106

Total Target Compounds: 0

INTEGRATED ANALYTICAL LABORATORIES

SEMIVOLATILE ORGANICS Tentatively Identified Compounds

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Extracted: 10/15/2001

Date Analyzed: 10/15/2001

Data file: B6861.D

GC/MS Column: DB-5

Sample wt/vol: 10.39g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.40

CAS #	Compound	Estimated Concentration	Retention Time
-------	----------	----------------------------	-------------------

No peaks detected

Total TICs = 0

INTEGRATED ANALYTICAL LABORATORIES

PCB's

Client/Project: BRENNAN/99704_FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Extracted: 10/15/2001

Date Analyzed: 10/16/2001

Data file: V6530.D

GC Column: DB-5/DB1701P

Sample wt/vol: 5.72g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.47

Compound	Concentration	Q	MDL
Aroclor-1016	ND		154
Aroclor-1221	ND		154
Aroclor-1232	ND		154
Aroclor-1242	ND		154
Aroclor-1248	ND		154
Aroclor-1254	ND		154
Aroclor-1260	ND		154

INTEGRATED ANALYTICAL LABORATORIES

PESTICIDES

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/2001

Date Extracted: 10/15/2001

Date Analyzed: 10/16/2001

Data file: W8928.D

GC Column: DB-5/DB1701P

Sample wt/vol: 5.7g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 1

% Moisture: 9.4

Compound	Concentration	Q	MDL
alpha-BHC	ND		3.87
beta-BHC	ND		3.87
gamma-BHC	ND		3.87
delta-BHC	ND		3.87
Heptachlor	ND		3.87
Aldrin	ND		3.87
Heptachlor epoxide	ND		3.87
Endosulfan I	ND		3.87
4,4'-DDE	ND		3.87
Dieldrin	ND		3.87
Endrin	ND		3.87
Endosulfan II	ND		3.87
4,4'-DDD	ND		3.87
Endrin aldehyde	ND		3.87
Endosulfan sulfate	ND		3.87
4,4'-DDT	ND		3.87
Chlordane	ND		19.4
Toxaphene	ND		19.4

INTEGRATED ANALYTICAL LABORATORIES, LLC.

METALS

Client/Project: BRENNAN/99704 FINETEX

Lab ID: 7041-001

Client ID: 5-F1

Date Received: 10/11/01

Matrix-Units: Soil mg/Kg (ppm)

% Moisture: 9.4

Batch #: 260

Compound	Result	Q	DF	MDL	Date Analyzed	Method
Antimony	ND		1	1.09	10/15/01	6020
Arsenic	1.70		1	1.09	10/15/01	6020
Beryllium	ND		1	0.546	10/15/01	6020
Cadmium	ND		1	0.273	10/15/01	6020
Chromium	9.00		1	2.18	10/15/01	6020
Copper	13.1		1	2.18	10/15/01	6020
Lead	14.4		1	0.546	10/15/01	6020
Mercury	0.0201		1	0.0138	10/15/01	7471 A
Nickel	8.48		1	1.09	10/15/01	6020
Selenium	ND		1	2.18	10/15/01	6020
Silver	ND		1	0.546	10/15/01	6020
Thallium	ND		1	0.109	10/15/01	6020
Zinc	26.1		1	2.18	10/15/01	6020

INTEGRATED ANALYTICAL LABORATORIES, LLC.

Total Recoverable Phenols

Client/Project: BRENNAN/99704 FINETEX

Date Received: 10/11/01

Lab ID	Client ID	Result	Q	DF	Matrix- Units	MDL	% Solids	Date Analyzed
7041-001	5-F1	ND		1	S-mg/Kg	2.76	90.6	0/12/2001

INTEGRATED ANALYTICAL LABORATORIES, LLC.

Total Cyanide

Client/Project: BRENNAN/99704 FINETEX

Date Received: 10/11/01

Lab ID	Client ID	Result	Q	DF	Matrix- Units	MDL	% Solids	Date Analyzed
7041-001	5-F1	ND		1	S-mg/Kg	1.10	90.6	0/23/2001

INTEGRATED ANALYTICAL LABORATORIES, LLC.

METHODOLOGY SUMMARY

<u>Analytical Parameter</u>	<u>Aqueous Samples</u> <u>Method #</u>	<u>Soil/Other Samples</u> <u>Method #</u>
Volatiles + 15	NA	EPA 8260B
Semivolatiles - BNA + 25	NA	EPA 8270C
PCB's	NA	EPA 8082
Pesticides	NA	EPA 8081A
P.P. Metals	NA	EPA 6020
Metals (Mercury)	NA	EPA 7471A
Cyanide, Total	NA	EPA 9012 A
Total Recoverable Phenols	NA	EPA 9066

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1746.D

BFB Injection Date: 10/10/2001

Inst ID: MSD I

BFB Injection Time: 9:22

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.5
75	30.0 - 60.0% of mass 95	57.3
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	6.4
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	69.3
175	5.0 - 9.0% of mass 174	5.9 (8.5)1
176	95.0 - 101.0% of mass 174	68.7 (99.1)1
177	5.0 - 9.0% of mass 176	4.5 (6.5)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
100	100_PPB_STD	I1747.D	10/10/2001	9:34
200	200_PPB_STD	I1748.D	10/10/2001	10:08
150	150_PPB_STD	I1749.D	10/10/2001	10:48
50	50_PPB_STD	I1750.D	10/10/2001	11:22
20	20_PPB_STD	I1751.D	10/10/2001	12:01
N/A	METHOD_BLK	I1752.D	10/10/2001	12:34
LCS	BLK_SPK	I1753.D	10/10/2001	13:07
TB	6805-017	I1754.D	10/10/2001	13:41
9B-2-V_3-3.5'	6805-002	I1755.D	10/10/2001	14:14
9B-2-VB_15.5-16'	6805-003	I1756.D	10/10/2001	14:48
9B-3-V_1.5-2'	6805-005	I1757.D	10/10/2001	15:22
9B-4_0-0.5'	6805-006	I1758.D	10/10/2001	15:56
9B-4-B_19.5-20'	6805-007	I1759.D	10/10/2001	16:29
9B-5-V_8.5-9'	6805-009	I1760.D	10/10/2001	17:03
9B-5B_11.5-12'	6805-010	I1761.D	10/10/2001	17:36
9B-1-V_9-9.5'	6805-012	I1762.D	10/10/2001	18:10
9B-1-B_13.5-14'	6805-013	I1763.D	10/10/2001	18:43
9B-6-V_8.5-9'	6805-015	I1764.D	10/10/2001	19:17
9B-6-B_11.5-12'	6805-016	I1765.D	10/10/2001	19:50
MS	6805-016MS	I1766.D	10/10/2001	20:24

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1746.D

BFB Injection Date : 10/10/200

Inst ID: MSD I

BFB Injection Time: 9:22

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.5
75	30.0 - 60.0% of mass 95	57.3
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	6.4
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	69.3
175	5.0 - 9.0% of mass 174	5.9 (8.5)1
176	95.0 - 101.0% of mass 174	68.7 (99.1)1
177	5.0 - 9.0% of mass 176	4.5 (6.5)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
MSD	6805-016MSD	I1767.D	10/10/2001	20:57

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1944.D BFB Injection Date: 10/17/2001
 Inst ID: MSD I BFB Injection Time: 1:55

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	31.7
75	30.0 - 60.0% of mass 95	59.1
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	8.0
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	81.8
175	5.0 - 9.0% of mass 174	4.1 (5.0)1
176	95.0 - 101.0% of mass 174	80.0 (97.8)1
177	5.0 - 9.0% of mass 176	5.8 (7.2)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
100	100_PPb_STD	I1946.D	10/17/2001	2:56
N/A	METHOD_BLK	I1948.D	10/17/2001	4:02
LCS	BLK_SPK	I1949.D	10/17/2001	4:35
TB	7040-007	I1950.D	10/17/2001	5:09
TRIP_BLANK	6996-010	I1951.D	10/17/2001	5:42
31-E-FILL_MAT_06	996-017	I1952.D	10/17/2001	6:15
6-S2	7040-005	I1953.D	10/17/2001	6:48
26D-1-B/2H-C-2-B	7090-003	I1955.D	10/17/2001	7:55
5R-3_0-0.5'	7090-005	I1956.D	10/17/2001	8:28
23A-3_0-0.5'	7090-008	I1957.D	10/17/2001	9:01
23A-4_0-0.5'	7090-009	I1958.D	10/17/2001	9:35
7B-1-V_1.5-2'	7090-010	I1959.D	10/17/2001	10:09
22G-3_0-0.5'	7090-012	I1960.D	10/17/2001	10:43
MS	7090-012MS	I1961.D	10/17/2001	11:17
MSD	7090-012MSD	I1962.D	10/17/2001	11:50
SB-43B_5-5.5	7085-005	I1963.D	10/17/2001	12:23
SB-43C_13.5-14	7085-006	I1964.D	10/17/2001	12:57

VOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: I1980.D

BFB Injection Date: 10/18/2001

Inst ID: MSD I

BFB Injection Time: 9:51

m/z	Ion Abundance Criteria	%Relative Abundance
50	15 - 40.0% of mass 95	30.0
75	30.0 - 60.0% of mass 95	57.8
95	Base peak, 100% relative abundance	100.0
96	5.0 - 9.0% of mass 95	8.8
173	Less than 2.0% of mass 174	0.0 (0.0)1
174	Great than 50.0% of mass 95	72.8
175	5.0 - 9.0% of mass 174	5.6 (7.7)1
176	95.0 - 101.0% of mass 174	70.0 (96.2)1
177	5.0 - 9.0% of mass 176	5.7 (8.1)2
	1-Value is % mass 174	2-Value is % mass 176

This check applies to the following: SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
100	100_PPb_STD	I1981.D	10/18/2001	10:04
N/A	METHOD_BLK	I1983.D	10/18/2001	11:10
LCS	BLK_SPK	I1984.D	10/18/2001	11:44
SB-40B_5-5.5	7151-002	I1985.D	10/18/2001	12:18
SB-44B_9.5-10	7151-007	I1986.D	10/18/2001	12:52
SB-44C_13.75-14.27	7151-008	I1987.D	10/18/2001	13:26
SB-44D_18-18.5	7151-009	I1988.D	10/18/2001	14:00
5-F1	7041-001	I1989.D	10/18/2001	14:34
5A-3-V_7-7.5'	6996-002	I1990.D	10/18/2001	15:08
5A-4-V_13-13.5'	6996-012	I1991.D	10/18/2001	15:41
5R-8-V_10-10.5'	6996-015	I1992.D	10/18/2001	16:15
IJ_WIPE	7061-001	I1993.D	10/18/2001	16:49
SB-1A	6924-004	I1996.D	10/18/2001	18:29
SB-1C	6924-006	I1997.D	10/18/2001	19:03
SB-43E_21.7-22.2	7085-008	I1998.D	10/18/2001	19:37
SB-43D_15.5-16	7085-007	I1999.D	10/18/2001	20:10
SB-43C_13.5-14	7085-006	I2000.D	10/18/2001	20:43

VOLATILE METHOD BLANK SUMMARY

Lab File ID: I1948.D

Instrument ID: MSD I

Date Analyzed: 10/17/2001

Time Analyzed: 04:02

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
LCS	BLK_SPK	10/17/2001	4:35
TB	7040-007	10/17/2001	5:09
TRIP_BLANK	6996-010	10/17/2001	5:42
31-E-FILL_MAT_0-9	6996-017	10/17/2001	6:15
6-S2	7040-005	10/17/2001	6:48
26D-1-B/2H-C-2-B	7090-003	10/17/2001	7:55
5R-3_0-0.5'	7090-005	10/17/2001	8:28
23A-3_0-0.5'	7090-008	10/17/2001	9:01
23A-4_0-0.5'	7090-009	10/17/2001	9:35
7B-1-V_1.5-2'	7090-010	10/17/2001	10:09
22G-3_0-0.5'	7090-012	10/17/2001	10:43
MS	7090-012MS	10/17/2001	11:17
MSD	7090-012MSD	10/17/2001	11:50
SB-43B_5-5.5	7085-005	10/17/2001	12:23
SB-43C_13.5-14	7085-006	10/17/2001	12:57

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/17/2001

Data file: I1948.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

Compound	Concentration	Q	MDL
Chloromethane	ND		250
Vinyl Chloride	ND		250
Bromomethane	ND		250
Chloroethane	ND		250
Trichlorofluoromethane	ND		250
Acrolein	ND		500
1,1-Dichloroethene	ND		250
Methylene Chloride	ND		250
Acrylonitrile	ND		500
trans-1,2-Dichloroethene	ND		250
1,1-Dichloroethane	ND		250
Chloroform	ND		250
1,1,1-Trichloroethane	ND		250
Carbon Tetrachloride	ND		250
1,2-Dichloroethane(EDC)	ND		250
Benzene	ND		250
Trichloroethene	ND		250
1,2-Dichloropropane	ND		250
Bromodichloromethane	ND		250
2-Chloroethylvinyl Ether	ND		250
cis-1,3-Dichloropropene	ND		250
Toluene	ND		250
trans-1,3-Dichloropropene	ND		250
1,1,2-Trichloroethane	ND		250
Tetrachloroethene	ND		250
Dibromochloromethane	ND		250
Chlorobenzene	ND		250
Ethylbenzene	ND		250
Total Xylenes	ND		250
Styrene	ND		250
Bromoform	ND		250
1,1,2,2-Tetrachloroethane	ND		250
1,3-Dichlorobenzene	ND		250
1,4-Dichlorobenzene	ND		250
1,2-Dichlorobenzene	ND		250
Naphthalene	ND		250

Total Target Compounds: 0

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS
Tentatively Identified Compounds

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/17/2001

Data file: I1948.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

CAS #	Compound	Estimated Concentration	Retention Time
-------	----------	----------------------------	-------------------

No peaks detected

Total TICs = 0

VOLATILE METHOD BLANK SUMMARY

Lab File ID: I1983.D

Instrument ID: MSD I

Date Analyzed: 10/18/2001

Time Analyzed: 11:10

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
LCS	BLK_SPK	10/18/2001	11:44
SB-40B_5-5.5	7151-002	10/18/2001	12:18
SB-44B_9.5-10	7151-007	10/18/2001	12:52
SB-44C_13.75-14.2	7151-008	10/18/2001	13:26
SB-44D_18-18.5	7151-009	10/18/2001	14:00
5-F1	7041-001	10/18/2001	14:34
5A-3-V_7-7.5'	6996-002	10/18/2001	15:08
5A-4-V_13-13.5'	6996-012	10/18/2001	15:41
5R-8-V_10-10.5'	6996-015	10/18/2001	16:15
IJ_WIPE	7061-001	10/18/2001	16:49
SB-1A	6924-004	10/18/2001	18:29
SB-1C	6924-006	10/18/2001	19:03
SB-43E_21.7-22.2	7085-008	10/18/2001	19:37
SB-43D_15.5-16	7085-007	10/18/2001	20:10
SB-43C_13.5-14	7085-006	10/18/2001	20:43

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/18/2001

Data file: I1983.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

Compound	Concentration	Q	MDL
Chloromethane	ND		250
Vinyl Chloride	ND		250
Bromomethane	ND		250
Chloroethane	ND		250
Trichlorofluoromethane	ND		250
Acrolein	ND		500
1,1-Dichloroethene	ND		250
Methylene Chloride	ND		250
Acrylonitrile	ND		500
trans-1,2-Dichloroethene	ND		250
1,1-Dichloroethane	ND		250
Chloroform	ND		250
1,1,1-Trichloroethane	ND		250
Carbon Tetrachloride	ND		250
1,2-Dichloroethane(EDC)	ND		250
Benzene	ND		250
Trichloroethene	ND		250
1,2-Dichloropropane	ND		250
Bromodichloromethane	ND		250
2-Chloroethylvinyl Ether	ND		250
cis-1,3-Dichloropropene	ND		250
Toluene	ND		250
trans-1,3-Dichloropropene	ND		250
1,1,2-Trichloroethane	ND		250
Tetrachloroethene	ND		250
Dibromochloromethane	ND		250
Chlorobenzene	ND		250
Ethylbenzene	ND		250
Total Xylenes	ND		250
Bromoform	ND		250
1,1,2,2-Tetrachloroethane	ND		250
1,3-Dichlorobenzene	ND		250
1,4-Dichlorobenzene	ND		250
1,2-Dichlorobenzene	ND		250

Total Target Compounds: 0

INTEGRATED ANALYTICAL LABORATORIES

VOLATILE ORGANICS Tentatively Identified Compounds

Client/Project: N/A

Lab ID: METHOD_BLK

Client ID: N/A

Date Received: N/A

Date Analyzed: 10/18/2001

Data file: I1983.D

GC/MS Column: DB-624

Sample wt/vol: 0.1g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 50

% Moisture: 0.0

CAS #	Compound	Estimated Concentration	Retention Time
-------	----------	-------------------------	----------------

No peaks detected

Total TICs = 0

Response Factor Report 5970 - In

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Last Update : Wed Oct 10 12:41:09 2001
 Response via : Initial Calibration

Calibration Files

20 =I1751.D 50 =I1750.D 100 =I1747.D
 150 =I1749.D 200 =I1748.D

Compound		20	50	100	150	200	Avg	%RSD
1) I	Pentafluorobenzene	-----ISTD-----						
2) T	Dichlorodifluorometha	0.315	0.311	0.294	0.284	0.290	0.299	4.58
3) P	Chloromethane	0.229	0.244	0.248	0.228	0.231	0.236	3.91
4) C	Vinyl Chloride	0.209	0.216	0.219	0.208	0.209	0.212	2.36
5) T	Bromomethane	0.289	0.265	0.233	0.219	0.223	0.246	12.24
6) T	Chloroethane	0.163	0.164	0.168	0.159	0.161	0.163	1.99
7) T	Trichlorofluoromethan	0.866	0.828	0.778	0.784	0.769	0.805	5.11
8) T	Acrolein	0.009	0.009	0.007	0.007	0.009	0.008	14.00
9) MC	1,1-Dichloroethene	0.274	0.278	0.279	0.260	0.263	0.271	3.26
10) T	Acetone	0.065	0.087	0.093	0.070	0.065	0.076	17.35
11) T	Carbon Disulfide	0.789	0.756	0.783	0.770	0.754	0.770	2.04
12) T	Vinyl Acetate	0.997	1.052	0.915	0.935	0.877	0.955	7.24
13) T	Methylene Chloride	0.310	0.316	0.301	0.300	0.284	0.302	3.95
14) T	Acrylonitrile	0.032	0.035	0.034	0.032	0.036	0.034	5.40
15) T	t-Butyl Alcohol (TBA)	0.018	0.016	0.019	0.020	0.016	0.018	9.22
16) T	trans-1,2-Dichloroeth	0.332	0.340	0.346	0.326	0.315	0.332	3.65
17) T	Methyl-t-Butyl Ether(	0.833	0.872	0.737	0.799	0.717	0.792	8.17
18) P	1,1-Dichloroethane	0.765	0.790	0.758	0.738	0.719	0.754	3.57
19) T	Diisopropyl Ether(DIP	1.467	1.485	1.351	1.382	1.319	1.401	5.18
20) T	cis-1,2-Dichloroethen	0.360	0.358	0.363	0.349	0.340	0.354	2.62
21) T	2,2-Dichloropropane	0.646	0.646	0.643	0.629	0.614	0.636	2.23
22) T	2-Butanone (MEK)	0.108	0.106	0.073	0.083	0.073	0.089	19.37
23) T	Bromochloromethane	0.229	0.236	0.220	0.221	0.208	0.223	4.72
25) C	Chloroform	1.040	1.014	1.000	0.975	0.959	0.998	3.21
26) T	1,1,1-Trichloroethane	0.950	0.950	0.911	0.920	0.885	0.923	3.02
27) T	Carbon Tetrachloride	0.708	0.775	0.666	0.734	0.685	0.714	5.97
28) T	1,1-Dichloropropene	0.542	0.544	0.519	0.524	0.509	0.527	2.83
29) T	1,2-Dichloroethane(ED	0.717	0.742	0.665	0.692	0.634	0.690	6.15
30) S	1,2-Dichloroethane-d4	0.592	0.649	0.607	0.660	0.651	0.632	4.77
31) I	1,4-Difluorobenzene	-----ISTD-----						
32) M	Benzene	0.698	0.703	0.706	0.675	0.648	0.686	3.54
33) M	Trichloroethene	0.476	0.460	0.454	0.430	0.414	0.447	5.48
34) C	1,2-Dichloropropane	0.353	0.342	0.332	0.322	0.314	0.333	4.69
35) T	Dibromomethane	0.304	0.319	0.282	0.278	0.256	0.288	8.45
37) T	Bromodichloromethane	0.809	0.807	0.755	0.788	0.708	0.773	5.47
38) T	2-Chloroethylvinyl Et	0.145	0.160	0.139	0.149	0.136	0.146	6.60
39) T	cis-1,3-Dichloroprope	0.478	0.503	0.476	0.461	0.435	0.471	5.32
40) T	4-Methyl-2-pentanone(	0.226	0.240	0.177	0.205	0.178	0.205	13.74
41) S	Toluene-d8	0.780	0.788	0.783	0.770	0.796	0.784	1.21
42) MC	Toluene	0.476	0.489	0.462	0.444	0.436	0.461	4.77
43) T	trans-1,3-Dichloropro	0.435	0.448	0.399	0.403	0.376	0.412	7.08
44) T	1,1,2-Trichloroethane	0.212	0.223	0.189	0.205	0.185	0.203	7.75
45) T	Tetrachloroethene	0.390	0.375	0.357	0.344	0.334	0.360	6.29
46) T	1,3-Dichloropropane	0.395	0.407	0.360	0.367	0.332	0.372	7.97
47) T	2-Hexanone	0.158	0.163	0.123	0.137	0.120	0.140	14.11
48) T	Dibromochloromethane	0.492	0.521	0.435	0.456	0.423	0.465	8.78
49) T	1,2-Dibromoethane(EDB	0.350	0.335	0.297	0.308	0.289	0.316	8.22

		-----ISTD-----						
50) I	Chlorobenzene-d5							
51) MP	Chlorobenzene	0.822	0.827	0.782	0.761	0.760	0.790	4.09
52) T	1,1,1,2-Tetrachloroet	0.532	0.545	0.480	0.524	0.491	0.514	5.37
53) C	Ethylbenzene	1.286	1.301	1.200	1.217	1.170	1.235	4.56
54) T	m,p-Xylene	0.436	0.437	0.413	0.415	0.398	0.420	3.97
55) T	o-Xylene	0.461	0.440	0.397	0.405	0.398	0.420	6.94
56) T	Styrene	0.697	0.737	0.654	0.705	0.680	0.695	4.47
57) P	Bromoform	0.371	0.399	0.342	0.382	0.346	0.368	6.52
58) T	Isopropylbenzene	1.141	1.076	1.078	1.069	1.036	1.080	3.55
59) S	Bromofluorobenzene	0.895	0.905	0.847	0.881	0.883	0.882	2.51
60) P	1,1,2,2-Tetrachloroet	0.337	0.349	0.276	0.314	0.287	0.313	10.06
61) T	Bromobenzene	0.452	0.403	0.382	0.400	0.390	0.405	6.72
62) T	1,2,3-Trichloropropan	0.422	0.388	0.322	0.334	0.318	0.357	12.89
63) T	n-Propylbenzene	1.332	1.269	1.275	1.195	1.207	1.255	4.45
64) T	2-Chlorotoluene	1.039	1.006	0.931	0.991	0.942	0.982	4.57
65) T	1,3,5-Trimethylbenzen	0.991	0.938	0.880	0.926	0.897	0.926	4.59
66) T	4-Chlorotoluene	1.215	1.200	1.120	1.178	1.085	1.160	4.76
67) T	tert-Butylbenzene	0.884	0.843	0.835	0.792	0.822	0.835	4.02
68) T	1,2,4-Trimethylbenzen	0.995	0.986	0.946	0.971	0.911	0.962	3.51
69) T	sec-Butylbenzene	0.994	0.958	0.906	0.925	0.900	0.937	4.18
70) T	1,3-Dichlorobenzene	0.658	0.683	0.573	0.616	0.580	0.622	7.73
71) T	4-Isopropyltoluene	0.879	0.824	0.817	0.769	0.777	0.813	5.42
72) T	1,4-Dichlorobenzene	0.658	0.683	0.573	0.616	0.580	0.622	7.73
73) T	n-Butylbenzene	0.850	0.805	0.796	0.754	0.757	0.792	4.95
74) T	1,2-Dichlorobenzene	0.586	0.569	0.477	0.516	0.468	0.523	10.18
75) T	1,2-Dibromo-3-chlorop	0.109	0.104	0.063	0.080	0.069	0.085	24.49
76) T	1,2,4-Trichlorobenzen	0.194	0.188	0.131	0.229	0.173	0.183	19.49
77) T	Hexachlorobutadiene	0.176	0.127	0.135	0.112	0.117	0.133	19.12
78) T	Naphthalene	0.249	0.315	0.194	0.361	0.241	0.272	24.19
79) T	1,2,3-Trichlorobenzen	0.167	0.196	0.094	0.171	0.115	0.149	28.61
80) T	1,1,2-Trichloro-1,2,2'	0.489	0.460	0.415	0.402	0.409	0.435	8.68
81) T	Methyl Acetate	0.173	0.175	0.134	0.159	0.155	0.159	10.39
82) T	Cyclohexane	0.323	0.295	0.266	0.265	0.264	0.283	9.19
83) T	Methylcyclohexane	0.212	0.208	0.180	0.178	0.184	0.192	8.39

(#) = Out of Range

IME1010.M

Wed Oct 10 12:41:37 2001

MSD_I

Instrument ID: MSD_I
Method ID: SOIL1003.M
Date: 10/03/2001

Average %RSD = 10.22

Refer to SW846 Method 8000B Section 7.5.1.

Evaluate Continuing Calibration Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1946.D

Acq On : 17 Oct 2001 2:56

Sample : 100,100_PPb_STD,W,5mL,100

Misc : N/A,N/A,N/A

MS Integration Params: rteint.p

Vial: 29

Operator: DP

Inst : 5970 - In

Multiplr: 1.00

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Multiple Level Calibration

Min. RRF : 0.000 Min. Rel. Area : 50% Max. R.T. Dev 0.50min

Max. RRF Dev : 30% Max. Rel. Area : 200%

	Compound	AvgRF	CCRF	%Dev	Area%	Dev(min)
1 I	Pentafluorobenzene	1.000	1.000	0.0	167	-0.03
2 T	Dichlorodifluoromethane	0.299	0.337	-12.7	192	-0.02
3 P	Chloromethane	0.236	0.187	20.8	127	-0.01
4 C	Vinyl Chloride	0.212	0.181	14.6	139	-0.03
5 T	Bromomethane	0.246	0.249	-1.2	179	-0.02
6 T	Chloroethane	0.163	0.152	6.7	152	-0.02
7 T	Trichlorofluoromethane	0.805	0.616	23.5	132	-0.04
8 T	Acrolein	0.008	0.007	12.5	183	-0.03
9 MC	1,1-Dichloroethene	0.271	0.266	1.8	160	-0.03
10 T	Acetone	0.076	0.071	6.6	128	-0.05
11 T	Carbon Disulfide	0.770	0.720	6.5	154	-0.02
12 T	Vinyl Acetate	0.955	0.956	-0.1	175	-0.04
13 T	Methylene Chloride	0.302	0.304	-0.7	169	-0.03
14 T	Acrylonitrile	0.034	0.031	8.8	154	-0.05
15 T	t-Butyl Alcohol (TBA)	0.018	0.014	22.2	122	-0.05
16 T	trans-1,2-Dichloroethene	0.332	0.340	-2.4	164	-0.05
17 T	Methyl-t-Butyl Ether (MTBE)	0.792	0.734	7.3	167	-0.04
18 P	1,1-Dichloroethane	0.754	0.713	5.4	158	-0.03
19 T	Diisopropyl Ether (DIPE)	1.401	1.309	6.6	162	-0.03
20 T	cis-1,2-Dichloroethene	0.354	0.361	-2.0	166	-0.04
21 T	2,2-Dichloropropane	0.636	0.519	18.4	135	-0.04
22 T	2-Butanone (MEK)	0.089	0.077	13.5	176	-0.04
23 T	Bromochloromethane	0.223	0.220	1.3	167	-0.03
25 C	Chloroform	0.998	0.902	9.6	151	-0.04
26 T	1,1,1-Trichloroethane	0.923	0.748	19.0	137	-0.03
27 T	Carbon Tetrachloride	0.714	0.577	19.2	145	-0.02
28 T	1,1-Dichloropropene	0.527	0.498	5.5	161	-0.03
29 T	1,2-Dichloroethane (EDC)	0.690	0.594	13.9	149	-0.04
30 S	1,2-Dichloroethane-d4	0.632	0.536	15.2	148	-0.02
31 I	1,4-Difluorobenzene	1.000	1.000	0.0	177	-0.04
32 M	Benzene	0.686	0.698	-1.7	175	-0.04
33 M	Trichloroethene	0.447	0.400	10.5	156	-0.03
34 C	1,2-Dichloropropane	0.333	0.305	8.4	163	-0.03
35 T	Dibromomethane	0.288	0.235	18.4	148	-0.03
37 T	Bromodichloromethane	0.773	0.647	16.3	152	-0.04
38 T	2-Chloroethylvinyl Ether	0.146	0.121	17.1	154	-0.03
39 T	cis-1,3-Dichloropropene	0.471	0.423	10.2	157	-0.03
40 T	4-Methyl-2-pentanone (MIBK)	0.205	0.170	17.1	170	-0.02
41 S	Toluene-d8	0.784	0.795	-1.4	180	-0.03
42 MC	Toluene	0.461	0.462	-0.2	177	-0.03
43 T	trans-1,3-Dichloropropene	0.412	0.348	15.5	155	-0.03
44 T	1,1,2-Trichloroethane	0.203	0.192	5.4	180	-0.03
45 T	Tetrachloroethene	0.360	0.318	11.7	158	-0.02
46 T	1,3-Dichloropropane	0.372	0.328	11.8	161	-0.02

47	T	2-Hexanone	0.140	0.126	10.0	182	-0.02
48	T	Dibromochloromethane	0.465	0.381	18.1	155	-0.03
49	T	1,2-Dibromoethane (EDB)	0.316	0.271	14.2	162	-0.02
50	I	Chlorobenzene-d5	1.000	1.000	0.0	183	-0.03
51	MP	Chlorobenzene	0.790	0.722	8.6	169	-0.02
52	T	1,1,1,2-Tetrachloroethane	0.514	0.417	18.9	159	-0.03
53	C	Ethylbenzene	1.235	1.099	11.0	167	-0.03
54	T	m,p-Xylene	0.420	0.394	6.2	174	-0.02
55	T	o-Xylene	0.420	0.380	9.5	175	-0.02
56	T	Styrene	0.695	0.637	8.3	178	-0.02
57	P	Bromoform	0.368	0.277	24.7	148	-0.03
58	T	Isopropylbenzene	1.080	0.919	14.9	156	0.00
59	S	Bromofluorobenzene	0.882	0.809	8.3	175	-0.01
60	P	1,1,2,2-Tetrachloroethane	0.313	0.257	17.9	171	-0.02
61	T	Bromobenzene	0.405	0.346	14.6	165	-0.02
62	T	1,2,3-Trichloropropane	0.357	0.253	29.1	144	0.00
63	T	n-Propylbenzene	1.255	1.022	18.6	147	-0.03
64	T	2-Chlorotoluene	0.982	0.840	14.5	165	-0.03
65	T	1,3,5-Trimethylbenzene	0.926	0.783	15.4	163	0.00
66	T	4-Chlorotoluene	1.160	0.947	18.4	155	-0.02
67	T	tert-Butylbenzene	0.835	0.686	17.8	150	-0.03
68	T	1,2,4-Trimethylbenzene	0.962	0.799	16.9	155	-0.01
69	T	sec-Butylbenzene	0.937	0.809	13.7	163	0.00
70	T	1,3-Dichlorobenzene	0.622	0.503	19.1	161	-0.01
71	T	4-Isopropyltoluene	0.813	0.643	20.9	144	-0.01
72	T	1,4-Dichlorobenzene	0.622	0.503	19.1	161	-0.01
73	T	n-Butylbenzene	0.792	0.617	22.1	142	-0.01
74	T	1,2-Dichlorobenzene	0.523	0.402	23.1	154	-0.03
75	T	1,2-Dibromo-3-chloropropane	0.085	0.065	23.5	190	0.00
76	T	1,2,4-Trichlorobenzene	0.183	0.146	20.2	205	-0.02
77	T	Hexachlorobutadiene	0.133	0.077	42.1	105	0.00
78	T	Naphthalene	0.272	0.210	22.8	198	0.00
79	T	1,2,3-Trichlorobenzene	0.149	0.098	34.2	191	-0.02
80	T	1,1,2-Trichloro-1,2,2-trifl	0.435	0.430	1.1	190	-0.03
81	T	Methyl Acetate	0.159	0.132	17.0	180	-0.05
82	T	Cyclohexane	0.283	0.237	16.3	163	-0.03
83	T	Methylcyclohexane	0.192	0.154	19.8	156	-0.03

(#) = Out of Range

I1747.D IME1010.M

SPCC's out = 0 CCC's out = 0

Thu Oct 18 13:35:20 2001 MSD_I

Evaluate Continuing Calibration Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1981.D

Acq On : 18 Oct 2001 10:04

Sample : 100,100_PPb_STD,W,5mL,100

Misc : N/A,N/A,N/A

MS Integration Params: rteint.p

Vial: 1

Operator: DP

Inst : 5970 - In

Multiplr: 1.00

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Multiple Level Calibration

Min. RRF : 0.000 Min. Rel. Area : 50% Max. R.T. Dev 0.50min

Max. RRF Dev : 30% Max. Rel. Area : 200%

	Compound	AvgRF	CCRF	%Dev	Area%	Dev(min)
1 I	Pentafluorobenzene	1.000	1.000	0.0	165	-0.02
2 T	Dichlorodifluoromethane	0.299	0.238	20.4	133	-0.01
3 P	Chloromethane	0.236	0.251	-6.4	167	-0.01
4 C	Vinyl Chloride	0.212	0.226	-6.6	170	-0.01
5 T	Bromomethane	0.246	0.242	1.6	171	-0.03
6 T	Chloroethane	0.163	0.149	8.6	146	-0.09
7 T	Trichlorofluoromethane	0.805	0.675	16.1	143	-0.08
8 T	Acrolein	0.008	0.006	25.0	139	-0.03
9 MC	1,1-Dichloroethene	0.271	0.293	-8.1	173	-0.04
10 T	Acetone	0.076	0.056	26.3	100	-0.02
11 T	Carbon Disulfide	0.770	0.847	-10.0	178	-0.03
12 T	Vinyl Acetate	0.955	0.803	15.9	144	-0.01
13 T	Methylene Chloride	0.302	0.311	-3.0	170	0.00
14 T	Acrylonitrile	0.034	0.025	26.5	120	0.00
15 T	t-Butyl Alcohol (TBA)	0.018	0.016	11.1	144	0.04
16 T	trans-1,2-Dichloroethene	0.332	0.375	-13.0	178	-0.04
17 T	Methyl-t-Butyl Ether (MTBE)	0.792	0.663	16.3	148	-0.04
18 P	1,1-Dichloroethane	0.754	0.760	-0.8	165	-0.02
19 T	Diisopropyl Ether (DIPE)	1.401	1.287	8.1	157	-0.01
20 T	cis-1,2-Dichloroethene	0.354	0.365	-3.1	166	-0.02
21 T	2,2-Dichloropropane	0.636	0.535	15.9	137	0.00
22 T	2-Butanone (MEK)	0.089	0.073	18.0	164	0.00
23 T	Bromochloromethane	0.223	0.199	10.8	149	0.00
25 C	Chloroform	0.998	0.893	10.5	147	-0.02
26 T	1,1,1-Trichloroethane	0.923	0.784	15.1	142	-0.02
27 T	Carbon Tetrachloride	0.714	0.606	15.1	150	0.00
28 T	1,1-Dichloropropene	0.527	0.512	2.8	162	-0.01
29 T	1,2-Dichloroethane (EDC)	0.690	0.518	24.9	128	-0.03
30 S	1,2-Dichloroethane-d4	0.632	0.577	8.7	157	-0.01
31 I	1,4-Difluorobenzene	1.000	1.000	0.0	178	-0.02
32 M	Benzene	0.686	0.681	0.7	172	-0.03
33 M	Trichloroethene	0.447	0.413	7.6	162	0.00
34 C	1,2-Dichloropropane	0.333	0.284	14.7	152	-0.02
35 T	Dibromomethane	0.288	0.209	27.4	132	-0.02
37 T	Bromodichloromethane	0.773	0.613	20.7	144	0.00
38 T	2-Chloroethylvinyl Ether	0.146	0.114	21.9	146	-0.01
39 T	cis-1,3-Dichloropropene	0.471	0.384	18.5	144	-0.02
40 T	4-Methyl-2-pentanone (MIBK)	0.205	0.167	18.5	169	0.00
41 S	Toluene-d8	0.784	0.816	-4.1	185	-0.01
42 MC	Toluene	0.461	0.463	-0.4	178	0.00
43 T	trans-1,3-Dichloropropene	0.412	0.303	26.5	135	0.00
44 T	1,1,2-Trichloroethane	0.203	0.160	21.2	150	0.00
45 T	Tetrachloroethene	0.360	0.326	9.4	162	0.00
46 T	1,3-Dichloropropane	0.372	0.285	23.4	141	0.00

47	T	2-Hexanone	0.140	0.113	19.3	163	0.01
48	T	Dibromochloromethane	0.465	0.343	26.2	140	0.00
49	T	1,2-Dibromoethane (EDB)	0.316	0.230	27.2	138	0.01
50	I	Chlorobenzene-d5	1.000	1.000	0.0	180	0.00
51	MP	Chlorobenzene	0.790	0.712	9.9	164	0.00
52	T	1,1,1,2-Tetrachloroethane	0.514	0.413	19.6	155	0.00
53	C	Ethylbenzene	1.235	1.112	10.0	167	0.00
54	T	m,p-Xylene	0.420	0.385	8.3	168	0.00
55	T	o-Xylene	0.420	0.388	7.6	176	0.02
56	T	Styrene	0.695	0.612	11.9	169	0.02
57	P	Bromoform	0.368	0.269	26.9	142	0.00
58	T	Isopropylbenzene	1.080	0.962	10.9	161	0.01
59	S	Bromofluorobenzene	0.882	0.957	-8.5	204	0.01
60	P	1,1,2,2-Tetrachloroethane	0.313	0.259	17.3	169	0.01
61	T	Bromobenzene	0.405	0.322	20.5	152	0.01
62	T	1,2,3-Trichloropropane	0.357	0.290	18.8	162	0.01
63	T	n-Propylbenzene	1.255	1.069	14.8	151	0.00
64	T	2-Chlorotoluene	0.982	0.845	14.0	164	0.00
65	T	1,3,5-Trimethylbenzene	0.926	0.802	13.4	164	0.00
66	T	4-Chlorotoluene	1.160	0.987	14.9	159	0.00
67	T	tert-Butylbenzene	0.835	0.730	12.6	158	0.02
68	T	1,2,4-Trimethylbenzene	0.962	0.828	13.9	158	0.00
69	T	sec-Butylbenzene	0.937	0.868	7.4	173	0.01
70	T	1,3-Dichlorobenzene	0.622	0.509	18.2	160	0.00
71	T	4-Isopropyltoluene	0.813	0.716	11.9	158	0.01
72	T	1,4-Dichlorobenzene	0.622	0.509	18.2	160	0.00
73	T	n-Butylbenzene	0.792	0.676	14.6	153	0.00
74	T	1,2-Dichlorobenzene	0.523	0.418	20.1	158	0.00
75	T	1,2-Dibromo-3-chloropropane	0.085	0.066	22.4	187	0.01
76	T	1,2,4-Trichlorobenzene	0.183	0.134	26.8	185	0.00
77	T	Hexachlorobutadiene	0.133	0.097	27.1	129	0.02
78	T	Naphthalene	0.272	0.199	26.8	185	0.02
79	T	1,2,3-Trichlorobenzene	0.149	0.103	30.9	199	0.00
80	T	1,1,2-Trichloro-1,2,2-trifl	0.435	0.328	24.6	142	-0.05
81	T	Methyl Acetate	0.159	0.144	9.4	193	-0.02
82	T	Cyclohexane	0.283	0.234	17.3	159	-0.03
83	T	Methylcyclohexane	0.192	0.177	7.8	177	0.00

(#) = Out of Range

I1747.D IME1010.M

SPCC's out = 0 CCC's out = 0
Thu Oct 18 12:06:18 2001 MSD_I

VOLATILE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/17/2001

Lab Sample ID	Matrix	File ID	SMC1 #	SMC2 #	SMC3 #
METHOD_BLK	SOIL	I1948.D	81	100	91
BLK_SPK	SOIL	I1949.D	76	100	87
7040-007	MEOH	I1950.D	85	97	77
6996-010	MEOH	I1951.D	83	91	76
6996-017	MEOH	I1952.D	81	77	72
7040-005	MEOH	I1953.D	84	98	74
7090-003	MEOH	I1955.D	71	73	61
7090-005	MEOH	I1956.D	74	80	65
7090-008	MEOH	I1957.D	79	84	66
7090-009	MEOH	I1958.D	77	88	66
7090-010	MEOH	I1959.D	78	77	63
7090-012	MEOH	I1960.D	79	82	66
7090-012MS	MEOH	I1961.D	86	88	75
7090-012MSD	MEOH	I1962.D	82	86	69
7085-005	MEOH	I1963.D	100	100	100
7085-006	MEOH	I1964.D	80	90	70

	Concentration	Aqueous/Meoh	Soil
SMC1 = 1,2-Dichloroethane-d4	50 ppb	48-125	51-118
SMC2 = Toluene-d8	50 ppb	54-123	89-109
SMC3 = Bromofluorobenzene	50 ppb	46-118	80-111

Column to be used to flag recovery values

VOLATILE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/18/2001

Lab Sample ID	Matrix	File ID	SMC1 #	SMC2 #	SMC3 #
METHOD_BLK	SOIL	I1983.D	81	97	93
BLK_SPK	SOIL	I1984.D	84	103	93
7151-002	MEOH	I1985.D	64	70	58
7151-007	MEOH	I1986.D	74	85	74
7151-008	MEOH	I1987.D	68	72	62
7151-009	MEOH	I1988.D	90	85	80
7041-001	MEOH	I1989.D	76	75	71
6996-002	MEOH	I1990.D	86	88	86
6996-012	MEOH	I1991.D	N/A	N/A	N/A
6996-015	MEOH	I1992.D	100	100	100
7061-001	MEOH	I1993.D	77	80	73
6924-004	MEOH	I1996.D	80	90	80
6924-006	MEOH	I1997.D	80	100	90
7085-008	MEOH	I1998.D	66	76	63
7085-007	MEOH	I1999.D	59	71	57
7085-006	MEOH	I2000.D	56	67	51

	Concentration	Aqueous/Meoh	Soil
SMC1 = 1,2-Dichloroethane-d4	50 ppb	48-125	51-118
SMC2 = Toluene-d8	50 ppb	54-123	89-109
SMC3 = Bromofluorobenzene	50 ppb	46-118	80-111

Column to be used to flag recovery values

MEOH VOLATILE MATRIX SPIKE/SPIKE DUPLICATE RECOVERY

Matrix spike Lab sample ID: 7090-012

Batch No.: IM0233

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
1,1-Dichloroethene	50.0	0.0	48.6	97	48 - 149
Benzene	50.0	0.0	47.4	95	59 - 149
Trichloroethene	50.0	0.0	39.9	80	55 - 143
Toluene	50.0	0.0	45.5	91	54 - 150
Chlorobenzene	50.0	0.0	47.3	95	55 - 144

Compound	SAMPLE CONC. (ug/Kg)	MSD CONC. (ug/Kg)	MSD % # REC	% RPD #	QC LIMITS	
					RPD	REC.
1,1-Dichloroethene	0.0	49.8	100	3	17	48 - 149
Benzene	0.0	46.3	93	2	15	59 - 149
Trichloroethene	0.0	38.5	77	4	15	55 - 143
Toluene	0.0	45.6	91	0	16	54 - 150
Chlorobenzene	0.0	48.4	97	2	15	55 - 144

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

RPD: 0 out of 5 outside limits

Spike Recovery: 0 out of 10 outside limits

SOIL VOLATILE LABORATORY CONTROL SAMPLE RECOVERY

Matrix spike Lab sample ID: BLK SPK

Batch No.: IM0233

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
1,1-Dichloroethene	50.0	0.0	55.8	112	70 - 130
Benzene	50.0	0.0	51.5	103	70 - 130
Trichloroethene	50.0	0.0	43.7	87	70 - 130
Toluene	50.0	0.0	52.0	104	70 - 130
Chlorobenzene	50.0	0.0	47.3	95	70 - 130

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

Spike Recovery: 0 out of 5 outside limits

SOIL VOLATILE LABORATORY CONTROL SAMPLE RECOVERY

Matrix spike Lab sample ID: BLK SPK

Batch No.: IM0235

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
1,1-Dichloroethene	50.0	0.0	59.9	120	70 - 130
Benzene	50.0	0.0	54.9	110	70 - 130
Trichloroethene	50.0	0.0	45.5	91	70 - 130
Toluene	50.0	0.0	53.1	106	70 - 130
Chlorobenzene	50.0	0.0	46.3	93	70 - 130

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

Spike Recovery: 0 out of 5 outside limits

VOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): I1747.D

Date Analyzed: 10/10/2001

Instrument ID: MSD_I

Time Analyzed: 9:34

	50UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
	12 HOUR STD	653387	9.73	784077	10.95	634368	15.90
	UPPER LIMIT	1306774	10.23	1568154	11.45	1268736	16.40
	LOWER LIMIT	326693.5	9.23	392038.5	10.45	317184	15.40
	LAB SAMPLE ID						
01	200_PPb_STD	647856	9.70	798050	10.90	615497	15.84
02	150_PPb_STD	656854	9.65	814005	10.87	640682	15.82
03	50_PPb_STD	675342	9.65	821122	10.84	635952	15.80
04	20_PPb_STD	674167	9.65	799529	10.87	623610	15.82
05	METHOD_BLK	648350	9.64	792486	10.86	668256	15.81
06	BLK_SPK	656491	9.64	789960	10.86	643953	15.82
07	6805-017	694488	9.63	832579	10.85	592975	15.82
08	6805-002	700505	9.65	828125	10.87	586715	15.83
09	6805-003	708272	9.65	845648	10.88	603174	15.83
10	6805-005	770218	9.65	860282	10.85	626482	15.82
11	6805-006	758939	9.63	860569	10.85	628899	15.82
12	6805-007	779707	9.64	875607	10.86	633294	15.82
13	6805-009	734189	9.65	892746	10.87	624207	15.82
14	6805-010	775090	9.66	914234	10.86	684844	15.81
15	6805-012	787746	9.63	907557	10.85	657037	15.80
16	6805-013	788950	9.63	913993	10.85	642244	15.80
17	6805-015	795434	9.62	941670	10.85	663946	15.81
18	6805-016	803676	9.63	945405	10.86	671447	15.81
19	6805-016MS	813939	9.65	941658	10.86	677527	15.81
20	6805-016MSD	795946	9.66	950448	10.87	668293	15.83
21							
22							

IS1 = PENTAFLUOROBENZENE

IS2 = 1,4-DIFLUOROBENZENE

IS3 = CHLOROBENZENE-D5

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

VOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): I1946.D

Date Analyzed: 10/17/2001

Instrument ID: MSD_I

Time Analyzed: 2:56

	50UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
	12 HOUR STD	1093433	9.70	1389694	10.92	1160240	15.86
	UPPER LIMIT	2186866	10.20	2779388	11.42	2320480	16.36
	LOWER LIMIT	546716.5	9.20	694847	10.42	580120	15.36
	LAB SAMPLE ID						
01	METHOD_BLK	1047735	9.71	1387504	10.92	1182634	15.87
02	BLK_SPK	1069109	9.71	1381684	10.92	1165754	15.88
03	7040-007	1063650	9.70	1430560	10.92	1160991	15.87
04	6996-010	1092890	9.71	1407551	10.92	1169632	15.88
05	6996-017	1174540	9.71	1498070	10.91	1071222	15.88
06	7040-005	1186341	9.71	1443196	10.93	1087576	15.88
07	7090-003	1216331	9.72	1542878	10.93	1105934	15.88
08	7090-005	1235988	9.71	1497297	10.91	1101262	15.88
09	7090-008	1188178	9.68	1480973	10.90	1116937	15.85
10	7090-009	1208938	9.67	1461196	10.90	1146334	15.84
11	7090-010	1185468	9.67	1434970	10.87	1111598	15.83
12	7090-012	1211880	9.64	1493263	10.86	1110437	15.80
13	7090-012MS	1176326	9.61	1476249	10.83	1089950	15.78
14	7090-012MSD	1161925	9.61	1477952	10.82	1100599	15.78
15	7085-005	1168648	9.63	1391800	10.85	1070052	15.82
16	7085-006	1155584	9.62	1387096	10.84	1071641	15.79
17							
18							
19							
20							
21							
22							

IS1 = PENTAFLUOROBENZENE

IS2 = 1,4-DIFLUOROBENZENE

IS3 = CHLOROBENZENE-D5

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

VOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): I1981.D

Date Analyzed: 10/18/2001

Instrument ID: MSD_I

Time Analyzed: 10:04

50UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
12 HOUR STD	1076012	9.72	1395275	10.93	1144071	15.89
UPPER LIMIT	2152024	10.22	2790550	11.43	2288142	16.39
LOWER LIMIT	538006	9.22	697637.5	10.43	572035.5	15.39
LAB SAMPLE ID						
01 METHOD_BLK	979871	9.70	1328900	10.91	1166256	15.86
02 BLK_SPK	978347	9.69	1295224	10.89	1141282	15.86
03 7151-002	1259618	9.66	1525394	10.88	1048419	15.84
04 7151-007	1265139	9.67	1505730	10.87	1051577	15.83
05 7151-008	1232713	9.65	1530886	10.86	1061118	15.82
06 7151-009	1223793	9.63	1518847	10.85	1053581	15.80
07 7041-001	1240241	9.61	1508275	10.82	1042060	15.79
08 6996-002	1219108	9.60	1498709	10.83	1031945	15.78
09 6996-012	1199375	9.63	1478935	10.83	1042233	15.80
10 6996-015	1206901	9.63	1448399	10.85	1032766	15.81
11 7061-001	1222390	9.60	1499117	10.83	1042421	15.79
12 6924-004	1215307	9.65	1456996	10.87	995608	15.83
13 6924-006	1197069	9.68	1406644	10.90	988620	15.87
14 7085-008	1185234	9.69	1453316	10.92	1000206	15.88
15 7085-007	1196154	9.72	1417971	10.95	961446	15.90
16 7085-006	1173725	9.76	1434631	10.96	955522	15.93
17						
18						
19						
20						
21						
22						

IS1 = PENTAFLUOROBENZENE

IS2 = 1,4-DIFLUOROBENZENE

IS3 = CHLOROBENZENE-D5

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

VOLATILE METHANOL PRESERVATION CONTAINER SUMMARY

Sample ID	Client ID	Bottle ID	Date Prepared	Initial Wgt.(g)	Final Wgt.(g)	Sample Wgt.(g)	MeOH Lot #	Surrogate Lot #
7041-001	5-F1	111769	10/9/01	107.3	117.2	9.9	cc226	ls224

Limits : Sample Weight = 8 to 12 grams, * = Outside limits

Comments :

Sample weight is reported on a wet weight basis

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D Vial: 9
Acq On : 18 Oct 2001 14:34 Operator: DP
Sample : 5-F1,7041-001,M,0.040g Inst : 5970 - In
Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01 Multiplr: 1.00
MS Integration Params: rteint.p
Quant Time: Oct 19 12:20 2001 Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Last Update : Thu Oct 11 09:23:20 2001
Response via : Initial Calibration
DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) Pentafluorobenzene	9.61	168	1240241	50.00	UG	-0.12
31) 1,4-Difluorobenzene	10.82	114	1508275	50.00	UG	-0.13
50) Chlorobenzene-d5	15.79	117	1042060	50.00	UG	-0.11

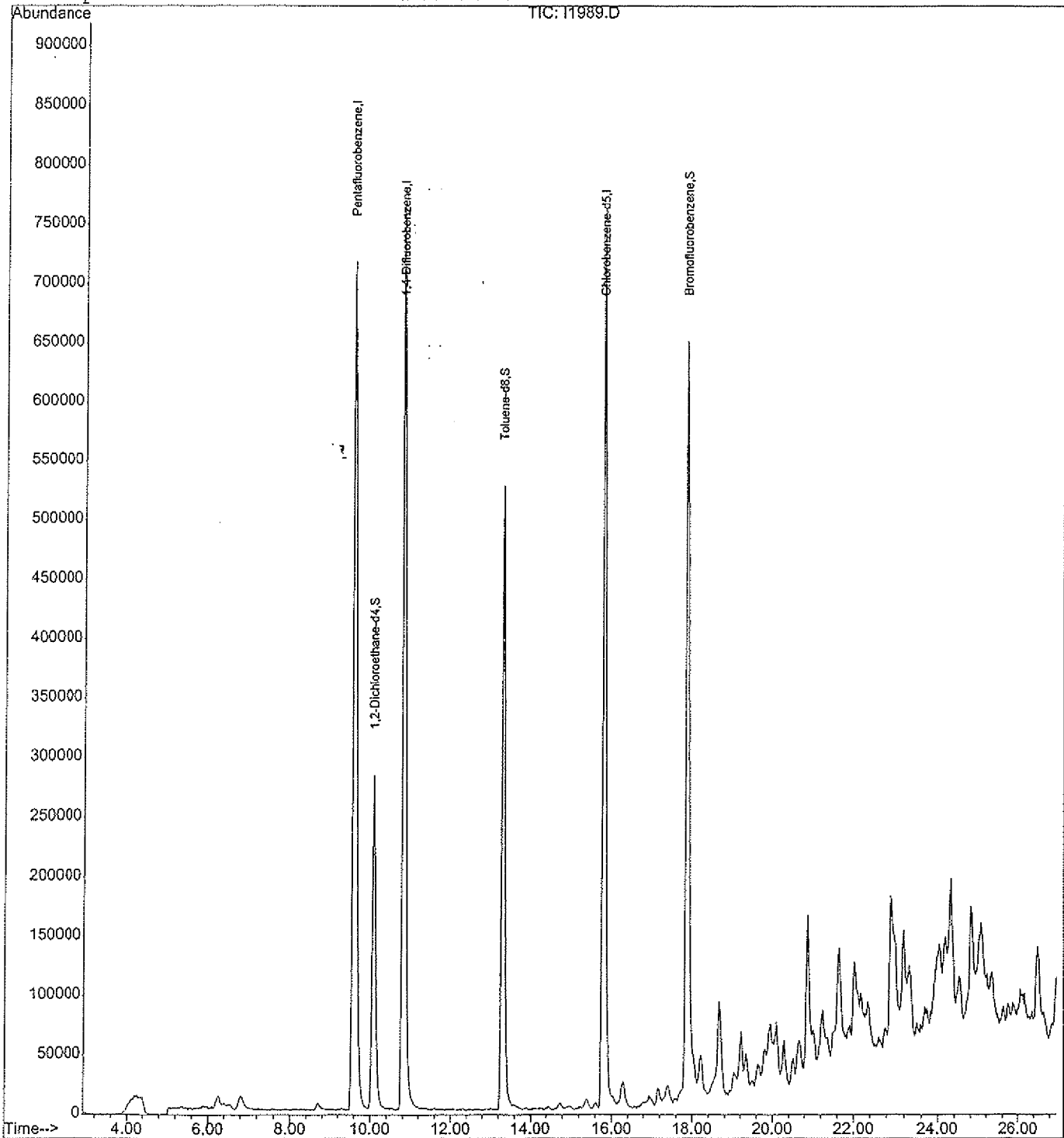
System Monitoring Compounds		R.T.	QIon	Response	Conc	Units	Dev(Min)
30) 1,2-Dichloroethane-d4		10.09	65	598259	38.17	UG	-0.12
Spiked Amount	50.000	Range	36 - 115	Recovery	=	76.34%	
41) Toluene-d8		13.30	98	888900	37.60	UG	-0.11
Spiked Amount	50.000	Range	35 - 117	Recovery	=	75.20%	
59) Bromofluorobenzene		17.87	95	651897	35.45	UG	-0.08
Spiked Amount	50.000	Range	47 - 115	Recovery	=	70.90%	

Target Compounds Qvalue

Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D Vial: 9
Acq On : 18 Oct 2001 14:34 Operator: DP
Sample : 5-F1,7041-001,M,0.040g Inst : 5970 - In
Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01 Multiplr: 1.00
MS Integration Params: rteint.p
Quant Time: Oct 19 12:20 2001 Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Last Update : Thu Oct 11 09:23:20 2001
Response via : Initial Calibration



LSC Area Percent Report

Data File : C:\HPCHEM\1\DATA\10-18-01\11989.D Vial: 9
 Acq On : 18 Oct 2001 14:34 Operator: DP
 Sample : 5-F1,7041-001,M,0.040g Inst : 5970 - In
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01 Multiplr: 1.00
 MS Integration Params: LSCINT.P

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Smoothing : ON Filtering: 5
 Sampling : 1 Min Area: 3 % of largest Peak
 Start Thrs: 0.2 Max Peaks: 100
 Stop Thrs : 0.15 Peak Location: TOP

If leading or trailing edge < 100 prefer < Baseline drop else tangent >
 Peak separation: 5

Signal : TIC

peak #	R.T. min	first scan	max scan	last scan	PK TY	peak height	corr. area	corr. % max.	% of total
1	4.395	99	101	110	rVB5	14602	52586	1.14%	0.189%
2	6.248	215	223	229	rBV2	10871	75991	1.65%	0.273%
3	6.835	251	261	269	rBV4	11758	99639	2.16%	0.357%
4	9.609	432	441	458	rBV	715254	4610355	100.00%	16.534%
5	10.086	464	472	486	rBV	279407	1554866	33.73%	5.576%
6	10.824	512	520	541	rBV	761634	4462644	96.80%	16.004%
7	13.304	673	681	697	rBV2	526009	2943790	63.85%	10.557%
8	15.385	810	816	823	rVB6	7798	49333	1.07%	0.177%
9	15.786	835	842	857	rBV	737600	4015038	87.09%	14.399%
10	16.282	867	874	885	rVB6	21474	156060	3.38%	0.560%
11	17.163	925	931	937	rBV3	14494	85703	1.86%	0.307%
12	17.409	942	947	955	rVB6	15823	103586	2.25%	0.371%
13	17.873	969	977	993	rVV	637064	3725518	80.81%	13.360%
14	18.214	993	999	1010	rVB8	33603	237954	5.16%	0.853%
15	18.676	1024	1029	1039	rVB2	79121	506847	10.99%	1.818%
16	19.034	1047	1052	1058	rBV8	16204	113202	2.46%	0.406%
17	19.220	1058	1064	1068	rVV2	47170	261050	5.66%	0.936%
18	19.345	1069	1072	1078	rVB6	27043	135610	2.94%	0.486%
19	19.639	1086	1091	1096	rBV7	17793	107793	2.34%	0.387%
20	19.810	1097	1102	1105	rVV4	24948	129804	2.82%	0.466%
21	19.949	1106	1111	1115	rVV3	41565	259304	5.62%	0.930%
22	20.103	1119	1121	1127	rVB3	44376	165398	3.59%	0.593%
23	20.288	1129	1133	1141	rVB3	37897	195685	4.24%	0.702%
24	20.521	1142	1148	1151	rBV6	21820	113130	2.45%	0.406%
25	20.659	1151	1157	1164	rBV6	29157	197546	4.28%	0.708%
26	20.874	1165	1171	1177	rBV2	125398	652590	14.15%	2.340%
27	21.246	1187	1195	1200	rBV5	39719	259875	5.64%	0.932%
28	21.645	1215	1221	1230	rVB	78464	497236	10.79%	1.783%
29	22.010	1241	1245	1254	rBV9	62042	463657	10.06%	1.663%
30	22.775	1291	1295	1298	rBV6	17137	87117	1.89%	0.312%
31	22.896	1298	1303	1306	rBV2	117988	542024	11.76%	1.944%

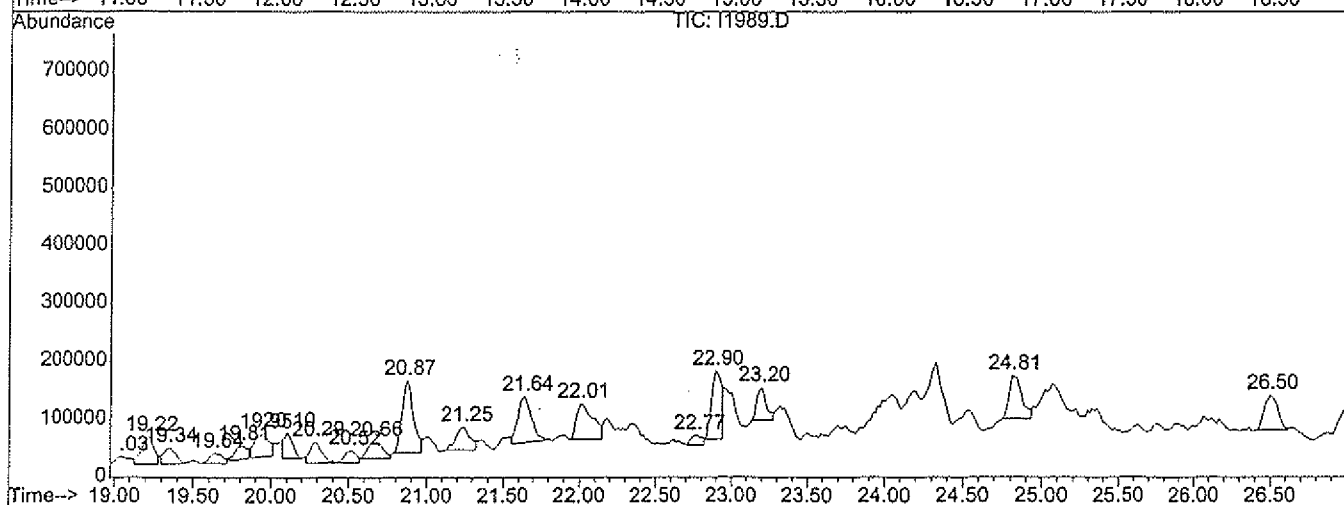
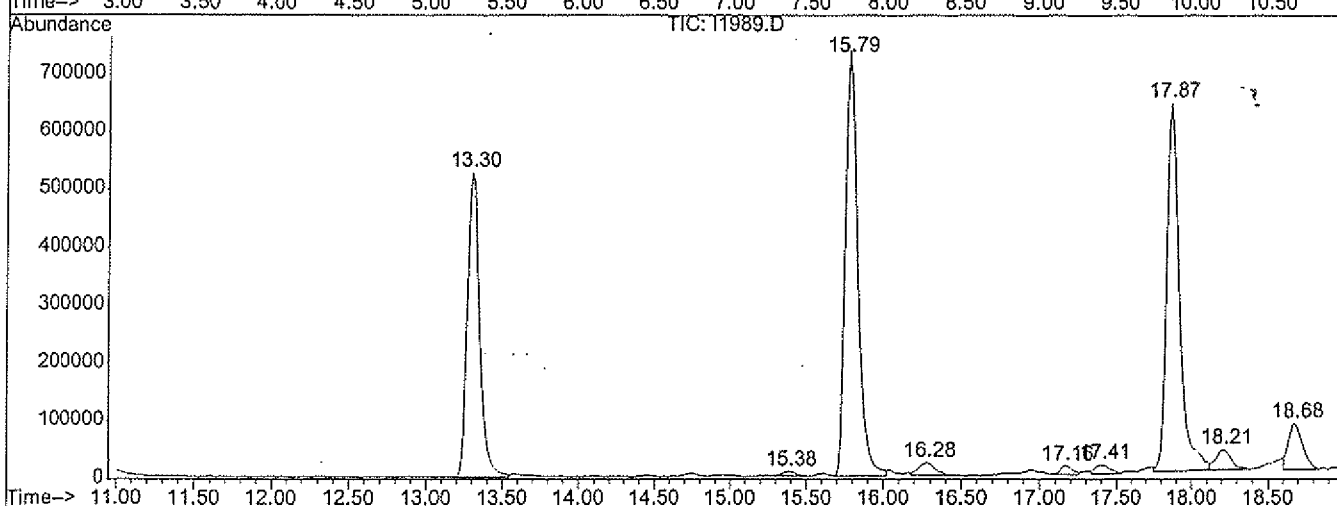
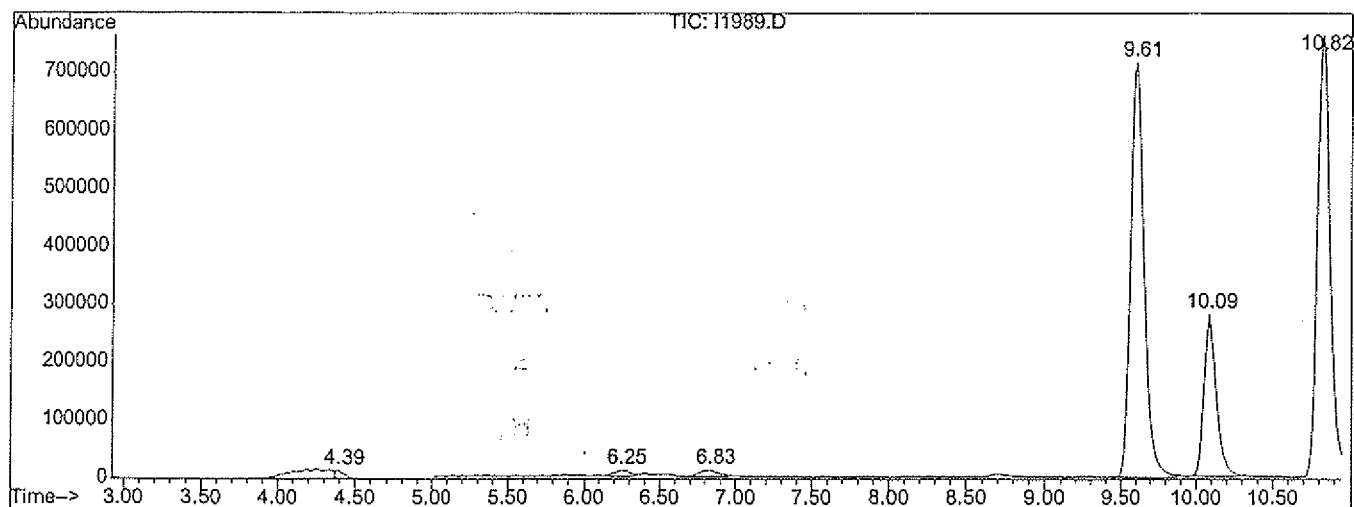
32	23.200	1319	1323	1328	rBV2	55505	233348	5.06%	0.837%
33	24.815	1426	1430	1438	rBV2	74096	420986	9.13%	1.510%
34	26.500	1537	1542	1550	rBV5	60094	369523	8.02%	1.325%

Sum of corrected areas: 27884788

I1989.D IME1010.M Fri Oct 19 12:31:24 2001 MSD_I

LSC Report - Integrated Chromatogram

File : C:\HPCHEM\1\DATA\10-18-01\I1989.D
 Operator : DP
 Acquired : 18 Oct 2001 14:34 using AcqMethod VOA1
 Instrument : 5970 - In
 Sample Name: 5-F1,7041-001,M,0.040g
 Misc Info : BRENNAN/99704_FINETEX,10/11/01,10/11/01
 Vial Number: 9
 Quant File : IME1010.RES (RTE Integrator)



I1989.D IME1010.M Fri Oct 19 12:31:24 2001 MSD_I

0001

Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D

Vial: 9

Acq On : 18 Oct 2001 14:34

Operator: DP

Sample : 5-F1,7041-001,M,0.040g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01

Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

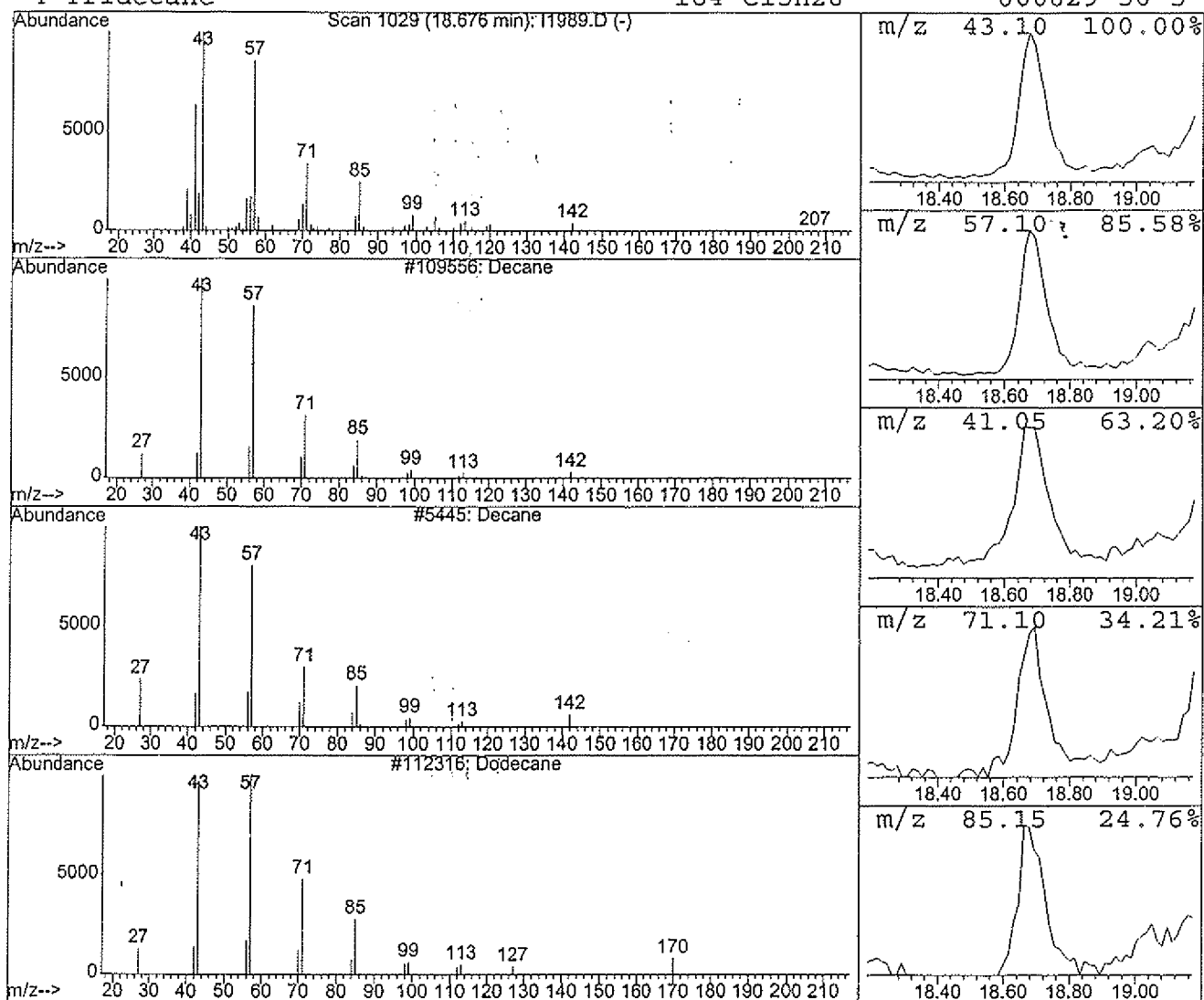
Library : C:\DATABASE\NIST98.L

Peak Number 1 Unknown alkane

Concentration Rank 3

R.T.	EstConc	Area	Relative to ISTD	R.T.
18.68	6.31 UG	506847	Chlorobenzene-d5	15.79

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1	Decane			142	C10H22	000124-18-5	87
2	Decane			142	C10H22	000124-18-5	72
3	Dodecane			170	C12H26	000112-40-3	64
4	Tridecane			184	C13H28	000629-50-5	64



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D

Vial: 9

Acq On : 18 Oct 2001 14:34

Operator: DP

Sample : 5-F1,7041-001,M,0.040g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01

Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

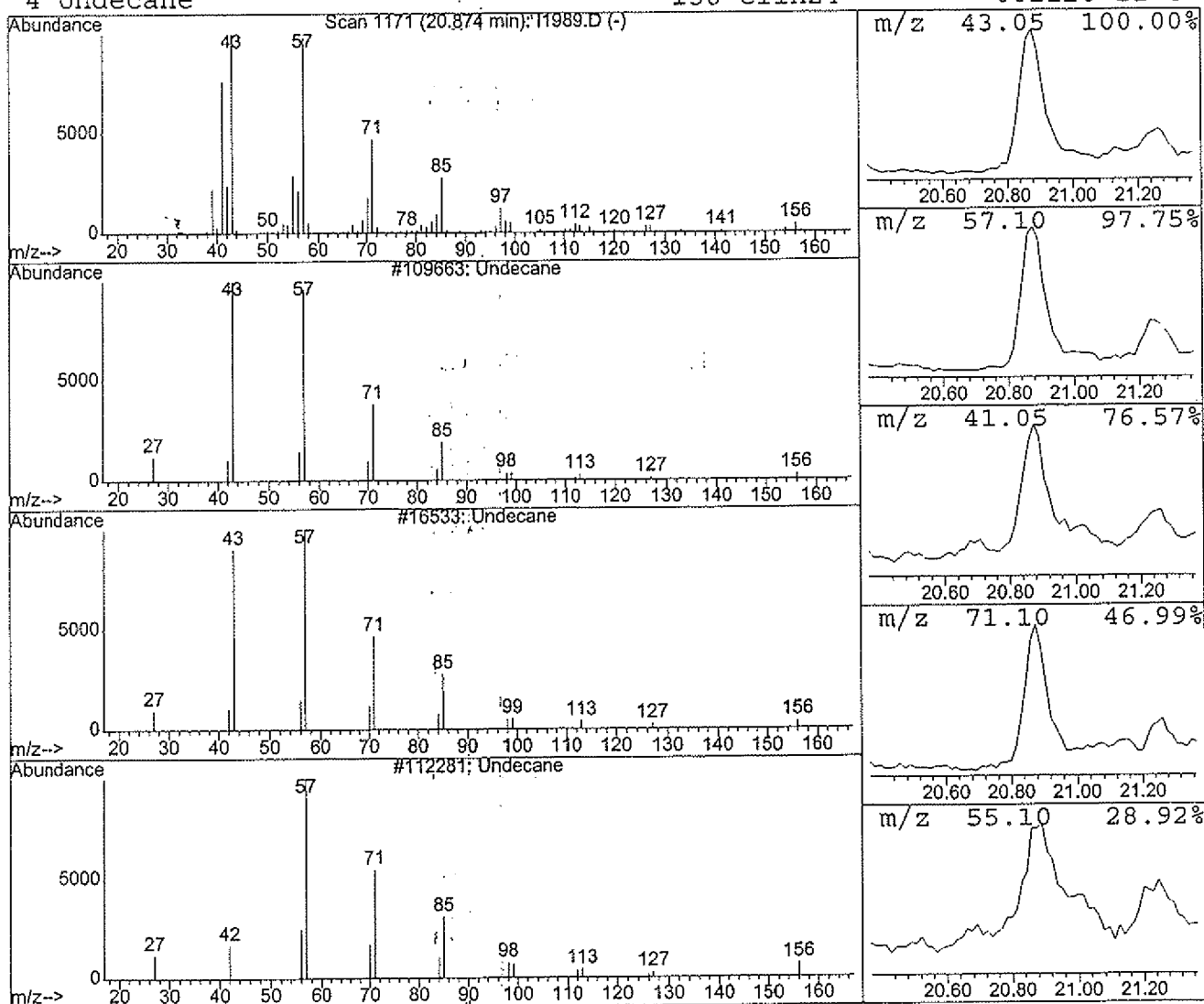
Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 2 Undecane Concentration Rank 1

R.T.	EstConc	Area	Relative to ISTD	R.T.
20.87	8.13 UG	652590	Chlorobenzene-d5	15.79

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1	Undecane			156	C11H24	001120-21-4	87
2	Undecane			156	C11H24	001120-21-4	87
3	Undecane			156	C11H24	001120-21-4	87
4	Undecane			156	C11H24	001120-21-4	81



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D

Vial: 9

Acq On : 18 Oct 2001 14:34

Operator: DP

Sample : 5-F1,7041-001,M,0.040g

Inst : 5970 - In

Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01

Multiplr: 1.00

MS Integration Params: LSCINT.P

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

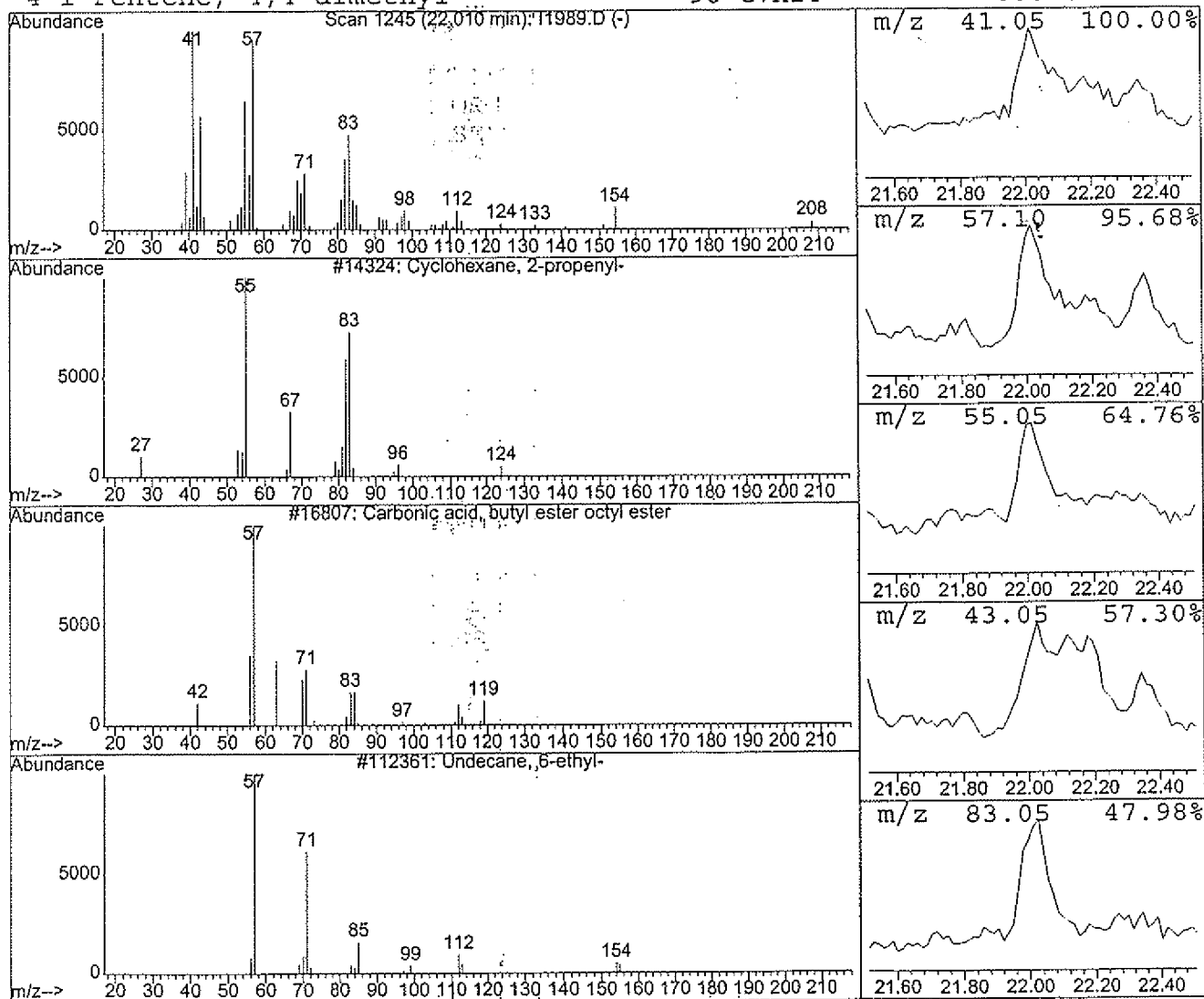
Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Library : C:\DATABASE\NIST98.L

Peak Number 3 Unknown alkane Concentration Rank 5

R.T.	EstConc	Area	Relative to ISTD	R.T.
22.01	5.77 UG	463657	Chlorobenzene-d5	15.79

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1			Cyclohexane, 2-propenyl-	124	C9H16	002114-42-3	38
2			Carbonic acid, butyl ester octyl es	230	C13H26O3	1000190-11-5	38
3			Undecane, 6-ethyl-	184	C13H28	017312-60-6	35
4			1-Pentene, 4,4-dimethyl-	98	C7H14	000762-62-9	35



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D
Acq On : 18 Oct 2001 14:34
Sample : 5-F1,7041-001,M,0.040g
Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01
MS Integration Params: LSCINT.P

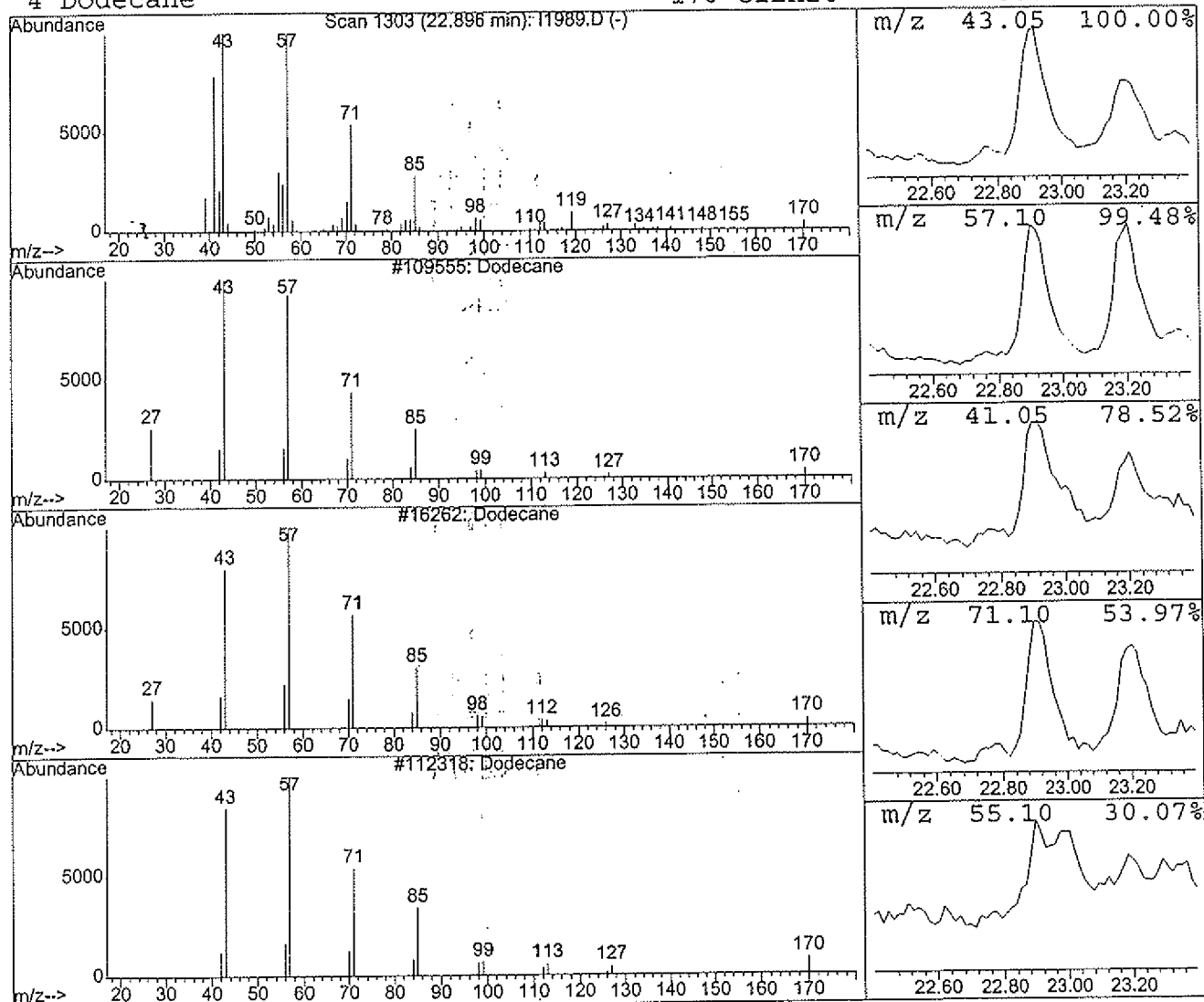
Vial: 9
Operator: DP
Inst : 5970 - In
Multiplr: 1.00

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Library : C:\DATABASE\NIST98.L

Peak Number 4 Dodecane Concentration Rank 2

R.T.	EstConc	Area	Relative to ISTD	R.T.
22.90	6.75 UG	542024	Chlorobenzene-d5	15.79

Hit#	of	5	Tentative ID	MW	MolForm	CAS#	Qual
1	Dodecane			170	C12H26	000112-40-3	94
2	Dodecane			170	C12H26	000112-40-3	93
3	Dodecane			170	C12H26	000112-40-3	81
4	Dodecane			170	C12H26	000112-40-3	81



Library Search Compound Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1989.D
 Acq On : 18 Oct 2001 14:34
 Sample : 5-F1,7041-001,M,0.040g
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01
 MS Integration Params: LSCINT.P

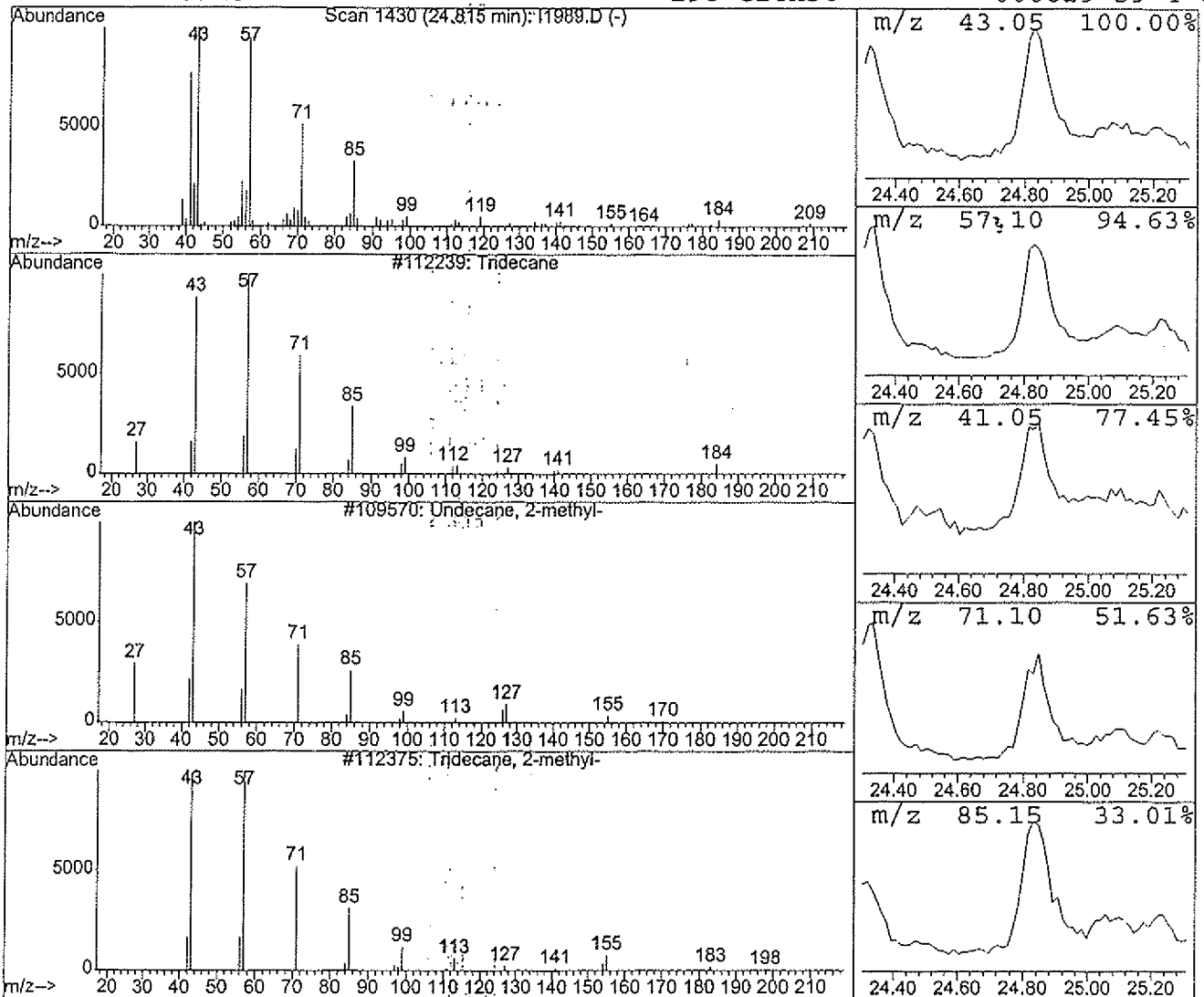
Vial: 9
 Operator: DP
 Inst : 5970 - In
 Multiplr: 1.00

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Library : C:\DATABASE\NIST98.L

 Peak Number 5 Unknown alkane Concentration Rank 6

R.T.	EstConc	Area	Relative to ISTD	R.T.
24.81	5.24 UG	420986	Chlorobenzene-d5	15.79

Hit# of 5	Tentative ID	MW	MolForm	CAS#	Qual
1	Tridecane	184	C13H28	000629-50-5	80
2	Undecane, 2-methyl-	170	C12H26	007045-71-8	72
3	Tridecane, 2-methyl-	198	C14H30	001560-96-9	64
4	Tetradecane	198	C14H30	000629-59-4	64



Quantitation Report

(QT Reviewed)

Data File : C:\HPCHEM\1\DATA\10-18-01\I1983.D

Vial: 3

Acq On : 18 Oct 2001 11:10

Operator: DP

Sample : N/A, METHOD BLK, S, 0.1g, 0.0

Inst : 5970 - In

Misc : N/A, N/A, N/A

Multiplr: 1.00

MS Integration Params: rteint.p

Quant Time: Oct 18 12:13 2001

Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Initial Calibration

DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) Pentafluorobenzene	9.70	168	979871	50.00	UG	-0.03
31) 1,4-Difluorobenzene	10.91	114	1328900	50.00	UG	-0.04
50) Chlorobenzene-d5	15.86	117	1166256	50.00	UG	-0.03

System Monitoring Compounds

30) 1,2-Dichloroethane-d4	10.17	65	502721	40.60	UG	-0.04
Spiked Amount	50.000	Range	36 - 115	Recovery	=	81.20%
41) Toluene-d8	13.38	98	1014573	48.71	UG	-0.04
Spiked Amount	50.000	Range	35 - 117	Recovery	=	97.42%
59) Bromofluorobenzene	17.93	95	958544	46.58	UG	-0.02
Spiked Amount	50.000	Range	47 - 115	Recovery	=	93.16%

Target Compounds

Qvalue

(#) = qualifier out of range (m) = manual integration

I1983.D IME1010.M

Thu Oct 18 12:13:38 2001

MSD_I

Page 1

TIERRA-B-010405

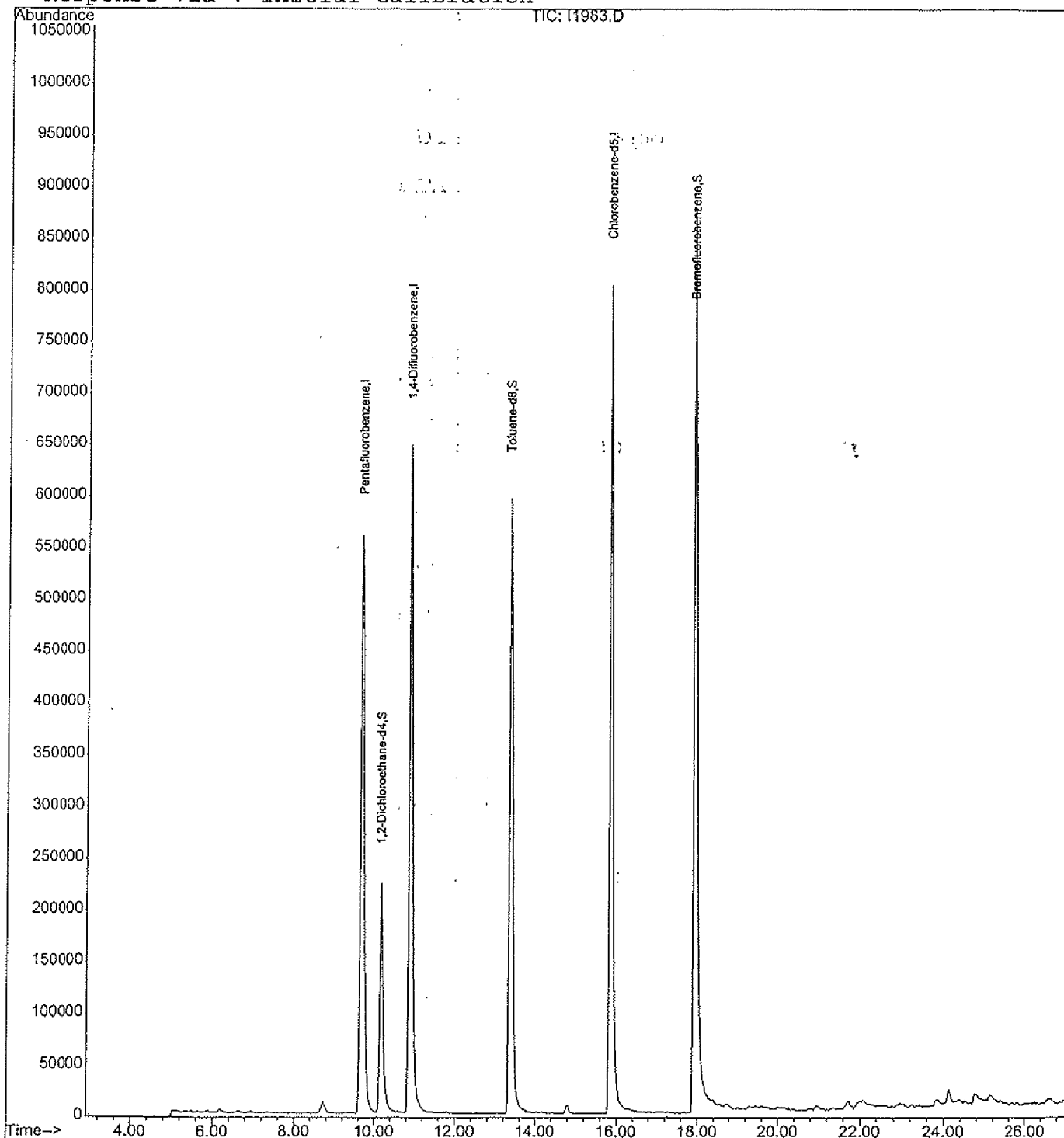
Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1983.D
 Acq On : 18 Oct 2001 11:10
 Sample : N/A, METHOD BLK, S, 0.1g, 0.0
 Misc : N/A, N/A, N/A
 MS Integration Params: rteint.p
 Quant Time: Oct 18 12:13 2001

Vial: 3
 Operator: DP
 Inst : 5970 - In
 Multiplr: 1.00

Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
 Title : VOLATILE ORGANICS BY EPA METHOD 8260B
 Last Update : Thu Oct 11 09:23:20 2001
 Response via : Initial Calibration



LSC Area Percent Report

Data File : C:\HPCHEM\1\DATA\10-18-01\I1983.D

Vial: 3

Acq On : 18 Oct 2001 11:10

Operator: DP

Sample : N/A,METHOD_BLK,S,0.1g,0.0

Inst : 5970 - In

Misc : N/A,N/A,N/A

Multiplr: 1.00

MS Integration Params: LSCINT.P

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Smoothing : OFF

Filtering: 5

Sampling : 1

Min Area: 1 % of largest Peak

Start Thrs: 0.2

Max Peaks: 100

Stop Thrs : 0

Peak Location: TOP

If leading or trailing edge < 100 prefer < Baseline drop else tangent >

Peak separation: 5

Signal : TIC

peak #	R.T. min	first scan	max scan	last scan	PK TY	peak height	corr. area	corr. % max.	% of total
1	8.730	376	383	392	rVB2	11148	73237	1.39%	0.329%
2	9.688	436	445	467	rBV	559178	3623461	68.94%	16.274%
3	10.182	469	477	499	rVB	222125	1345861	25.61%	6.045%
4	10.909	516	524	549	rBV	646623	3903418	74.27%	17.532%
5	13.378	677	684	704	rBV	594937	3362054	63.97%	15.100%
6	15.863	838	845	869	rBV	803087	4554965	86.66%	20.458%
7	17.934	973	979	1001	rBV	875690	5255861	100.00%	23.606%
8	24.136	1375	1380	1386	rBV8	13319	64079	1.22%	0.288%
9	24.754	1415	1420	1426	rBV9	11723	81786	1.56%	0.367%

Sum of corrected areas: 22264722

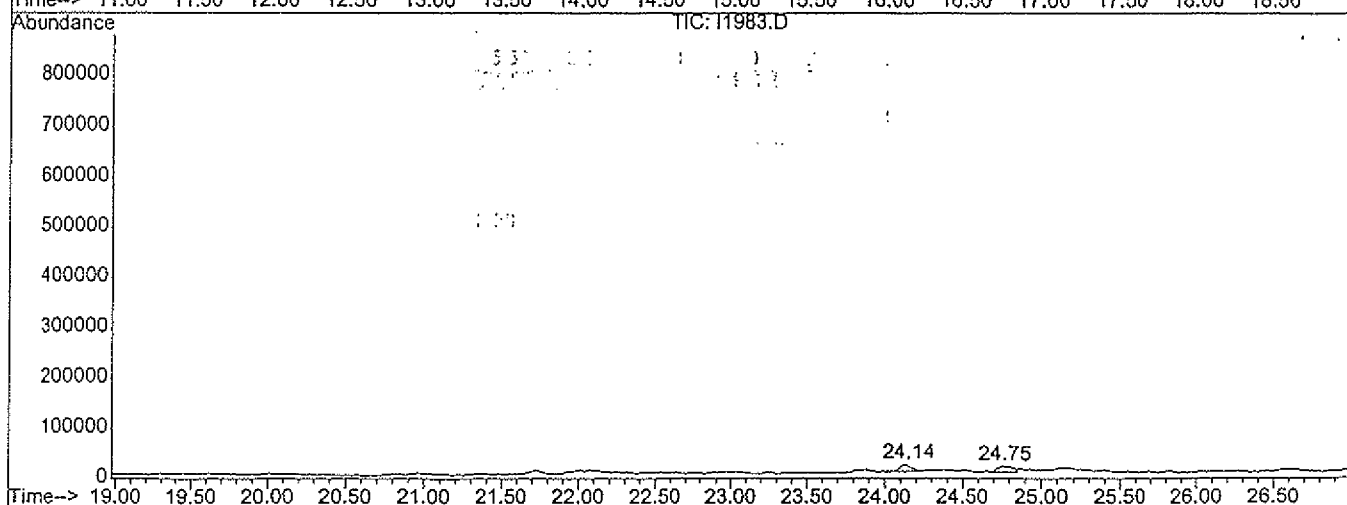
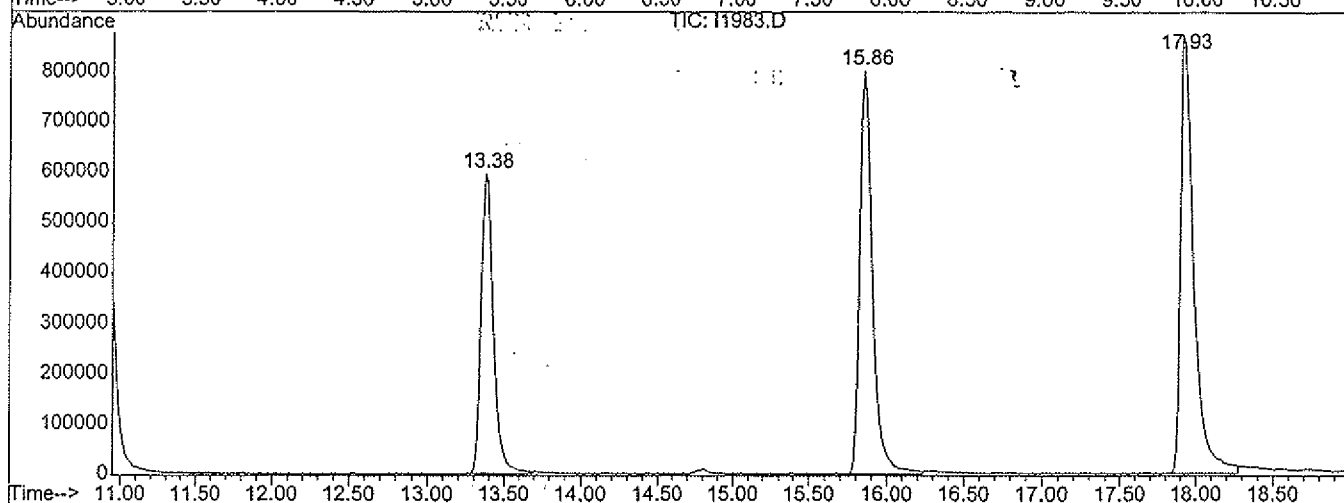
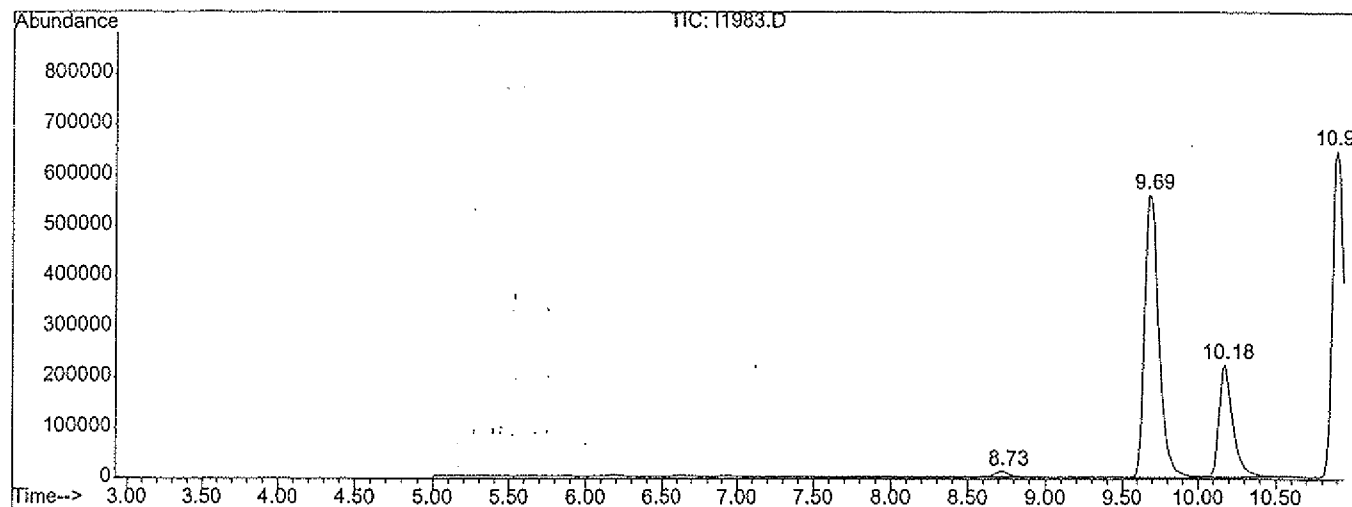
I1983.D IME1010.M

Thu Oct 18 12:13:45 2001

MSD_I

LSC Report - Integrated Chromatogram

File : C:\HPCHEM\1\DATA\10-18-01\I1983.D
 Operator : DP
 Acquired : 18 Oct 2001 11:10 using AcqMethod VOA1
 Instrument : 5970 - In
 Sample Name: N/A, METHOD BLK, S, 0.1g, 0.0
 Misc Info : N/A, N/A, N/A
 Vial Number: 3
 Quant File : IME1010.RES (RTE Integrator)



I1983.D IME1010.M

Thu Oct 18 12:13:45 2001

MSD_I

Quantitation Report (QT Reviewed)

Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D

Vial: 31

Acq On : 17 Oct 2001 4:02

Operator: DP

Sample : N/A, METHOD_BLK, S, 0.1g, 0.0

Inst : 5970 - In

Misc : N/A, N/A, N/A

Multiplr: 1.00

MS Integration Params: rteint.p

Quant Time: Oct 18 13:36 2001

Quant Results File: IME1010.RES

Quant Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)

Title : VOLATILE ORGANICS BY EPA METHOD 8260B

Last Update : Thu Oct 11 09:23:20 2001

Response via : Initial Calibration

DataAcq Meth : VOAI

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) Pentafluorobenzene	9.71	168	1047735	50.00	UG	-0.02
31) 1,4-Difluorobenzene	10.92	114	1387504	50.00	UG	-0.03
50) Chlorobenzene-d5	15.87	117	1182634	50.00	UG	-0.02

System Monitoring Compounds

30) 1,2-Dichloroethane-d4	10.20	65	533676	40.31	UG	0.00
Spiked Amount	50.000	Range	36 - 115	Recovery	=	80.62%
41) Toluene-d8	13.40	98	1090226	50.14	UG	-0.02
Spiked Amount	50.000	Range	35 - 117	Recovery	=	100.28%
59) Bromofluorobenzene	17.96	95	944331	45.25	UG	0.00
Spiked Amount	50.000	Range	47 - 115	Recovery	=	90.50%

Target Compounds

Qvalue

(#) = qualifier out of range (m) = manual integration

I1948.D IME1010.M

Thu Oct 18 13:36:49 2001

MSD_I

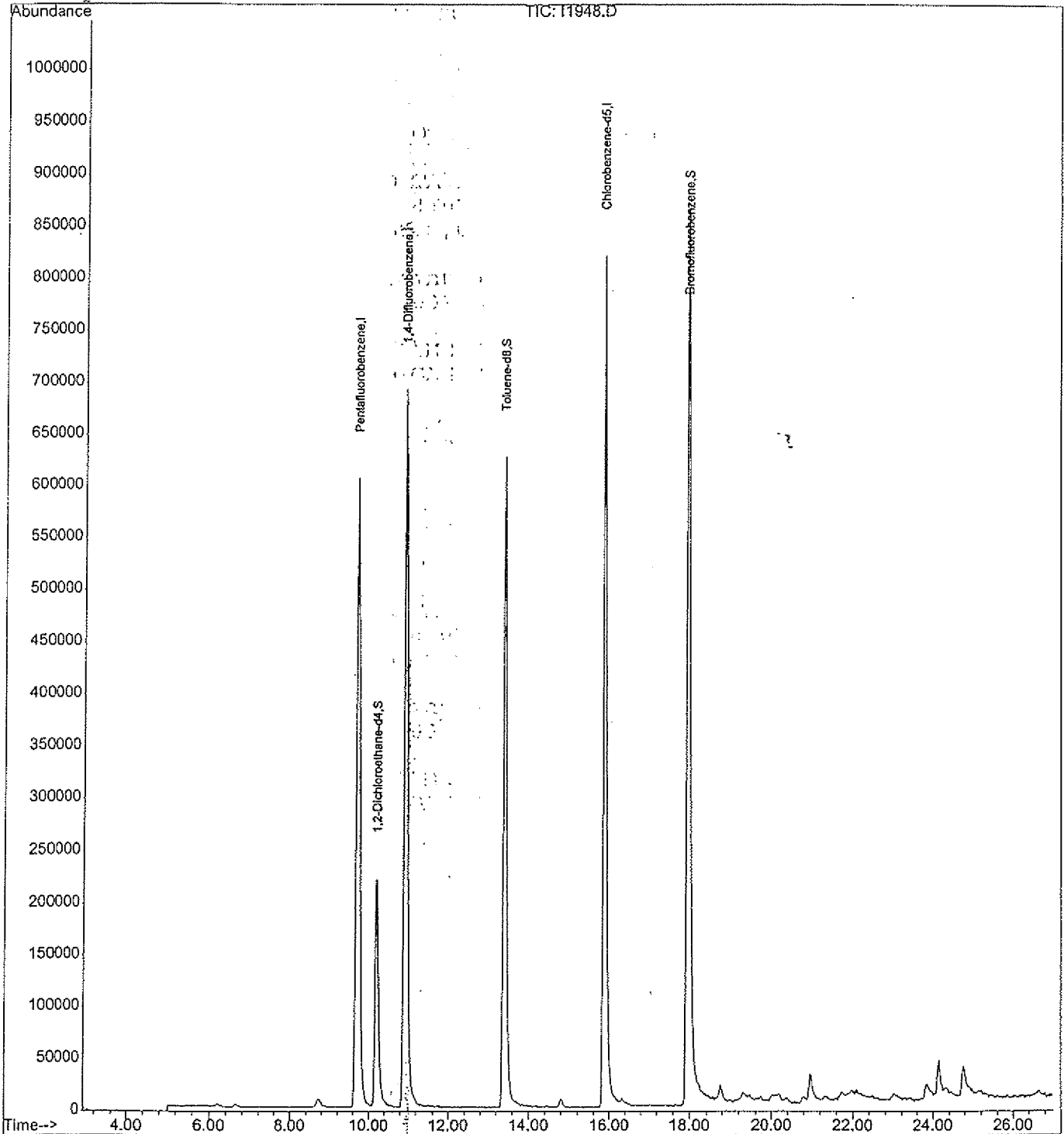
Page 1

Quantitation Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
Acq On : 17 Oct 2001 4:02
Sample : N/A,METHOD BLK,S,0.1g,0.0
Misc : N/A,N/A,N/A
MS Integration Params: rteint.p
Quant Time: Oct 18 13:36 2001

Vial: 31
Operator: DP
Inst : 5970 - In
Multiplr: 1.00
Quant Results File: IME1010.RES

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Last Update : Thu Oct 11 09:23:20 2001
Response via : Initial Calibration



LSC Area Percent Report

Data File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
Acq On : 17 Oct 2001 4:02
Sample : N/A,METHOD_BLK,S,0.1g,0.0
Misc : N/A,N/A,N/A
MS Integration Params: LSCINT.P

Vial: 31
Operator: DP
Inst : 5970 - In
Multiplr: 1.00

Method : C:\HPCHEM\1\METHODS\IME1010.M (RTE Integrator)
Title : VOLATILE ORGANICS BY EPA METHOD 8260B
Smoothing : OFF Filtering: 5
Sampling : 1 Min Area: 1 % of largest Peak
Start Thrs: 0.2 Max Peaks: 100
Stop Thrs : 0 Peak Location: TOP

If leading or trailing edge < 100 prefer < Baseline drop else tangent >
Peak separation: 5

Signal : TIC

peak #	R.T. min	first scan	max scan	last scan	PK TY	peak height	corr. area	corr. % max.	% of total
1	9.711	438	447	462	rBV	604591	3830995	75.35%	16.546%
2	10.201	471	479	491	rBV	218025	1363833	26.82%	5.890%
3	10.923	518	526	550	rBV	689956	4065116	79.95%	17.557%
4	13.399	679	687	700	rBV	625464	3560689	70.03%	15.379%
5	15.871	841	848	865	rBV	818788	4556628	89.62%	19.680%
6	17.958	977	984	1000	rBV	866079	5084454	100.00%	21.960%
7	18.751	1032	1036	1041	rVB5	12835	53258	1.05%	0.230%
8	20.969	1174	1180	1189	rVV3	25691	165419	3.25%	0.714%
9	23.851	1361	1367	1373	rBV8	14062	101750	2.00%	0.439%
10	24.143	1380	1386	1393	rBV10	32926	173906	3.42%	0.751%
11	24.769	1420	1427	1434	rBV4	28846	197383	3.88%	0.853%

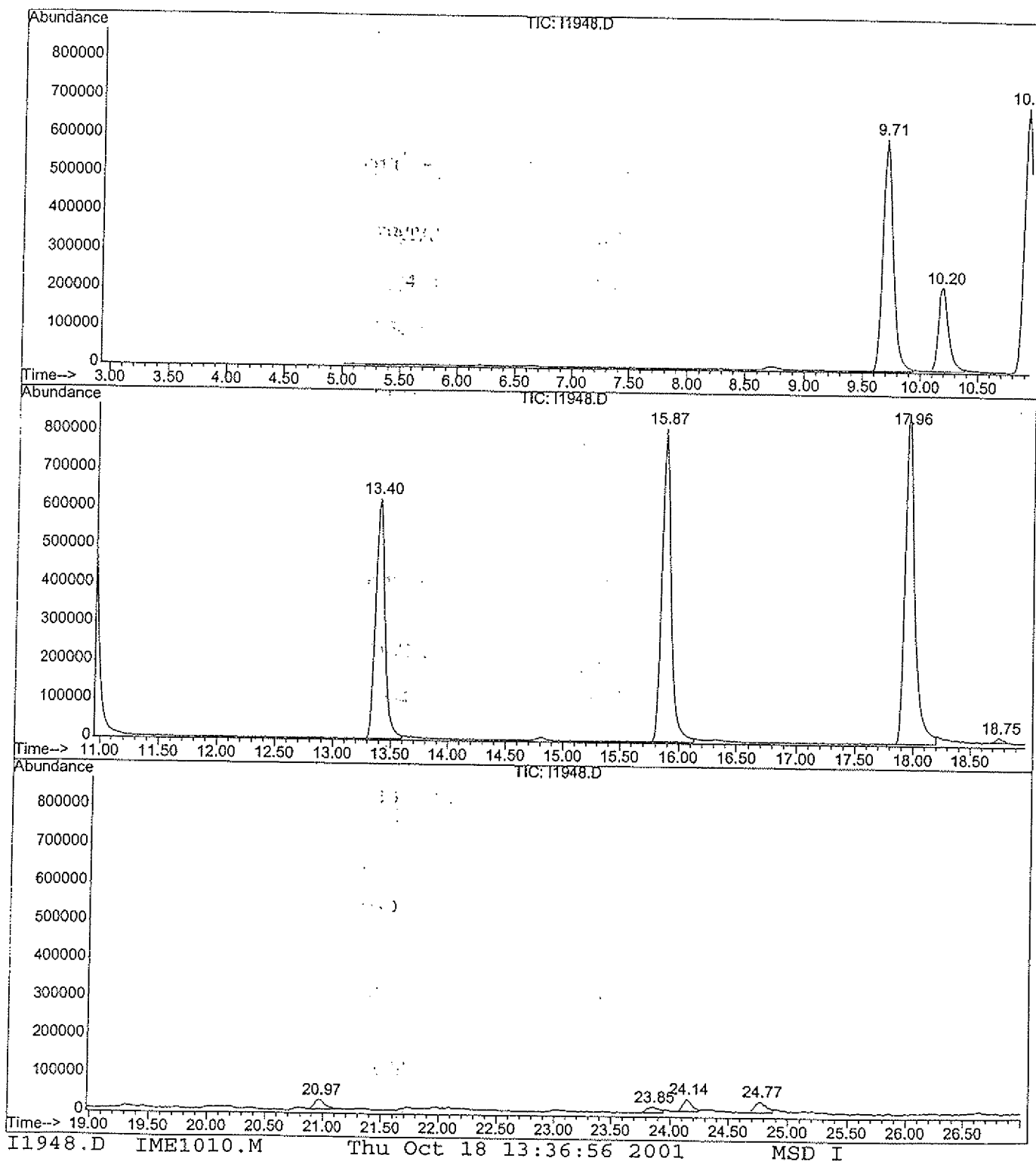
Sum of corrected areas: 23153431

I1948.D IME1010.M

Thu Oct 18 13:36:56 2001 MSD_I

LSC Report - Integrated Chromatogram

File : C:\HPCHEM\1\DATA\10-16-01\I1948.D
 Operator : DP
 Acquired : 17 Oct 2001 4:02 using AcqMethod VOAI
 Instrument : 5970 - In
 Sample Name: N/A,METHOD BLK,S,0.1g,0.0
 Misc Info : N/A,N/A,N/A
 Vial Number: 31
 Quant File : IME1010.RES (RTE Integrator)



SEMIVOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: B6618.D

DFTPP Injection Date : 10/08/2001

Inst ID: MSDB

DFTPP Injection Time: 07:29

m/z	Ion Abundance Criteria	%Relative Abundance
51	30.0 - 60.0% of mass 198	44.1
68	Less than 2.0% of mass 69	0.0 (0.0)1
69	Mass 69 relative abundance	43.3
70	Less than 2.0% of mass 69	0.0 (0.0)1
127	40.0 - 60.0% of mass 198	53.0
197	Less than 1.0% of mass 198	0.0
198	Base peak, 100% relative abundance	100.0
199	5.0 - 9.0% of mass 198	5.9
275	10.0 - 30.0% of mass 198	28.3
365	Greater than 1.0% of mass 198	4.8
441	Present, but less than mass 443	7.33 (49.4)3
442	40.0 - 100.0% of mass 198	70.0
443	17.0 - 23.0% of mass 442	14.8 (21.2)2

1-Value is % mass 69 2-Value is % mass 442 3-Value is % mass 443

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
.	80-bna-STD-INJ-F	B6619.D	10/08/2001	07:54
.	20-bna-STD-INJ-F	B6620.D	10/08/2001	08:26
.	50-bna-STD-INJ-F	B6621.D	10/08/2001	08:41
.	120-bna-STD-INJ-	B6622.D	10/08/2001	09:03
.	160-bna-STD-INJ-	B6623.D	10/08/2001	09:19
.	80-OLMO4-STD-INB	B6624.D	10/08/2001	09:34
.	20-OLMO4-STD-INB	B6625.D	10/08/2001	09:54
.	50-OLMO4-STD-INB	B6626.D	10/08/2001	10:09
.	120-OLMO4-STD-	B6627.D	10/08/2001	10:24
.	160-OLMO4-STD-	B6628.D	10/08/2001	10:39
.	80-OLMO4-STD-INB	B6629.D	10/08/2001	10:54
4A-1	6683-003	B6630.D	10/08/2001	11:09
T3B_4-4.5	6677-003	B6631.D	10/08/2001	11:25
T4B_4-4.5	6677-004	B6632.D	10/08/2001	11:40
.	METHOD-BLK	B6633.D	10/08/2001	11:56
T4B_4-4.5	6677-004MS	B6634.D	10/08/2001	12:19
T4B_4-4.5	6677-004MSD	B6635.D	10/08/2001	12:34
.	LCS	B6636.D	10/08/2001	12:49
.	METHOD-BLK	B6637.D	10/08/2001	13:05
18-1_0-0.5	6764-002	B6638.D	10/08/2001	13:20
30N-1_0-0.5	6764-004	B6639.D	10/08/2001	13:35
5S-1_0-0.5	6764-005	B6640.D	10/08/2001	13:50
9F-3_0-0.5	6764-006	B6641.D	10/08/2001	14:06

SEMIVOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: B6618.D

DFTPP Injection Date : 10/08/2001

Inst ID: MSDB

DFTPP Injection Time: 07:29

m/z	Ion Abundance Criteria	%Relative Abundance
51	30.0 - 60.0% of mass 198	44.1
68	Less than 2.0% of mass 69	0.0 (0.0)1
69	Mass 69 relative abundance	43.3
70	Less than 2.0% of mass 69	0.0 (0.0)1
127	40.0 - 60.0% of mass 198	53.0
197	Less than 1.0% of mass 198	0.0
198	Base peak, 100% relative abundance	100.0
199	5.0 - 9.0% of mass 198	5.9
275	10.0 - 30.0% of mass 198	28.3
365	Greater than 1.0% of mass 198	4.8
441	Present, but less than mass 443	7.3 (49.4)3
442	40.0 - 100.0% of mass 198	70.0
443	17.0 - 23.0% of mass 442	14.8 (21.2)2

1-Value is % mass 69 2-Value is % mass 442 3-Value is % mass 443

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
9F-1_2-2.5	6764-007	B6642.D	10/08/2001	14:21
1G-3_0-0.5	6764-008	B6643.D	10/08/2001	14:36
OS-SED-1	6442-001	B6644.D	10/08/2001	14:51
B-1	6793-001	B6645.D	10/08/2001	15:06
B-2A	6793-002	B6646.D	10/08/2001	15:22
B-2B	6793-003	B6647.D	10/08/2001	15:37
B-4	6793-005	B6648.D	10/08/2001	15:52
B-6	6793-007	B6649.D	10/08/2001	16:07
B-7	6793-008	B6650.D	10/08/2001	16:22
B-7	6793-008MS	B6651.D	10/08/2001	16:38
B-7	6793-008MSD	B6652.D	10/08/2001	16:53
.	LCS	B6653.D	10/08/2001	17:08
SW1D1_0-0.5	6666-021	B6654.D	10/08/2001	17:23
SW1D2_0-0.5	6666-022	B6655.D	10/08/2001	17:39
SW1D3_0-0.5	6666-023	B6656.D	10/08/2001	17:54

SEMIVOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: B6834.D

DFTPP Injection Date : 10/15/2001

Inst ID: MSDB

DFTPP Injection Time: 08:23

m/z	Ion Abundance Criteria	%Relative Abundance		
51	30.0 - 60.0% of mass 198	38.5		
68	Less than 2.0% of mass 69	0.0	(0.0)	1
69	Mass 69 relative abundance	45.8		
70	Less than 2.0% of mass 69	0.0	(0.0)	1
127	40.0 - 60.0% of mass 198	52.1		
197	Less than 1.0% of mass 198	0.0		
198	Base peak, 100% relative abundance	100.0		
199	5.0 - 9.0% of mass 198	6.5		
275	10.0 - 30.0% of mass 198	24.8		
365	Greater than 1.0% of mass 198	4.3		
441	Present, but less than mass 443	14.61	(72.0)	3
442	40.0 - 100.0% of mass 198	93.5		
443	17.0 - 23.0% of mass 442	20.3	(21.7)	2

1-Value is % mass 69 2-Value is % mass 442 3-Value is % mass 443

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
.	80-bna-STD-INJ-F	B6835.D	10/15/2001	08:56
.	80-OLMO4-STD-INB	B6836.D	10/15/2001	09:11
S-9B	6844-003	B6837.D	10/15/2001	09:26
S-9_2-2.5	6844-001	B6838.D	10/15/2001	09:42
S-10_2-2.5	6844-005	B6839.D	10/15/2001	09:57
S-9C	6844-004	B6840.D	10/15/2001	10:13
S-10C	6844-008	B6841.D	10/15/2001	10:34
.	METHOD-BLK	B6842.D	10/15/2001	11:07
PX-1	7058-001	B6843.D	10/15/2001	11:22
PX-2	7058-002	B6844.D	10/15/2001	11:38
PX-3	7058-003	B6845.D	10/15/2001	11:53
PX-4	7058-004	B6846.D	10/15/2001	12:08
PX-5	7058-005	B6847.D	10/15/2001	12:24
PX-6	7058-006	B6848.D	10/15/2001	12:39
PX-7	7058-007	B6849.D	10/15/2001	12:54
PX-8	7058-008	B6850.D	10/15/2001	13:10
PX-9	7058-009	B6851.D	10/15/2001	13:25
PX-10	7058-010	B6852.D	10/15/2001	13:41
PX-11	7058-011	B6853.D	10/15/2001	13:56
PX-12	7058-012	B6854.D	10/15/2001	14:11
SB-43A_3.5-4	7085-004	B6855.D	10/15/2001	14:27
SB-43B_5-5.5	7085-005	B6856.D	10/15/2001	14:58
SB-43C_13.5-14	7085-006	B6857.D	10/15/2001	15:14

SEMIVOLATILE ORGANIC INSTRUMENT PERFORMANCE CHECK

Lab File ID: B6834.D

DFTPP Injection Date : 10/15/2001

Inst ID: MSDB

DFTPP Injection Time: 08:23

m/z	Ion Abundance Criteria	%Relative Abundance
51	30.0 - 60.0% of mass 198	38.5
68	Less than 2.0% of mass 69	0.0 (0.0)1
69	Mass 69 relative abundance	45.8
70	Less than 2.0% of mass 69	0.0 (0.0)1
127	40.0 - 60.0% of mass 198	52.1
197	Less than 1.0% of mass 198	0.0
198	Base peak, 100% relative abundance	100.0
199	5.0 - 9.0% of mass 198	6.5
275	10.0 - 30.0% of mass 198	24.8
365	Greater than 1.0% of mass 198	4.3
441	Present, but less than mass 443	14.6 (72.0)3
442	40.0 - 100.0% of mass 198	93.5
443	17.0 - 23.0% of mass 442	20.3 (21.7)2

1-Value is % mass 69 2-Value is % mass 442 3-Value is % mass 443

This check applies to the following SAMPLES, MS, MSD, BLANKS and STANDARDS:

Client ID	Lab Sample ID	File ID	Date Analyzed	Time Analyzed
SB-43D_15.5-16	7085-007	B6858.D	10/15/2001	15:29
SB-43E_21.7-22.2	7085-008	B6859.D	10/15/2001	15:44
ABSEGAMI	7035-001	B6860.D	10/15/2001	15:59
5-F1	7041-001	B6861.D	10/15/2001	16:15
IJ-WIPE	7061-001	B6862.D	10/15/2001	16:30
PX-1	7058-001MS	B6863.D	10/15/2001	16:48
PX-1	7058-001MSD	B6864.D	10/15/2001	17:03
	LCS	B6865.D	10/15/2001	17:20

SEMIVOLATILE METHOD BLANK SUMMARY

Lab File ID: B6842.D

Instrument ID: MSDB

Date Extracted: 10/15/01

Matrix: SOIL

Date Analyzed: 10/15/2001

Time Analyzed: 11:07

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
PX-1	7058-001	10/15/2001	11:22
PX-2	7058-002	10/15/2001	11:38
PX-3	7058-003	10/15/2001	11:53
PX-4	7058-004	10/15/2001	12:08
PX-5	7058-005	10/15/2001	12:24
PX-6	7058-006	10/15/2001	12:39
PX-7	7058-007	10/15/2001	12:54
PX-8	7058-008	10/15/2001	13:10
PX-9	7058-009	10/15/2001	13:25
PX-10	7058-010	10/15/2001	13:41
PX-11	7058-011	10/15/2001	13:56
PX-12	7058-012	10/15/2001	14:11
SB-43A_3.5-4	7085-004	10/15/2001	14:27
SB-43B_5-5.5	7085-005	10/15/2001	14:58
SB-43C_13.5-14	7085-006	10/15/2001	15:14
SB-43D_15.5-16	7085-007	10/15/2001	15:29
SB-43E_21.7-22.	7085-008	10/15/2001	15:44
ABSEGAMI	7035-001	10/15/2001	15:59
5-F1	7041-001	10/15/2001	16:15
IJ-WIPE	7061-001	10/15/2001	16:30
PX-1	7058-001MS	10/15/2001	16:48
PX-1	7058-001MSD	10/15/2001	17:03
	LCS	10/15/2001	17:20

INTEGRATED ANALYTICAL LABORATORIES
SEMIVOLATILE ORGANICS

Client/Project: N/A

Lab ID: METHOD-BLK
Client ID: .
Date Received: N/A
Date Extracted: 10/15/2001
Date Analyzed: 10/15/2001
Data file: B6842.D

GC/MS Column: DB-5
Sample wt/vol: 30.0g
Matrix-Units: Soil- μ g/Kg (ppb)
Dilution Factor: 1
% Moisture: 0.0

Compound	Concentration	Q	MDL
N-Nitrosodimethylamine	ND		33.3
Benzaldehyde	ND		33.3
Phenol	ND		33.3
Aniline	ND		33.3
bis(2-Chloroethyl)ether	ND		33.3
2-Chlorophenol	ND		33.3
1,3-Dichlorobenzene	ND		33.3
1,4-Dichlorobenzene	ND		33.3
Benzyl alcohol	ND		33.3
1,2-Dichlorobenzene	ND		33.3
2-Methylphenol	ND		33.3
bis(2-chloroisopropyl)ether	ND		33.3
4-Methylphenol	ND		33.3
N-Nitroso-di-n-propylamine	ND		33.3
Acetophenone	ND		33.3
Hexachloroethane	ND		33.3
Nitrobenzene	ND		33.3
Isophorone	ND		33.3
2-Nitrophenol	ND		33.3
2,4-Dimethylphenol	ND		33.3
bis(2-Chloroethoxy)methane	ND		33.3
Benzoic acid	ND		33.3
2,4-Dichlorophenol	ND		33.3
1,2,4-Trichlorobenzene	ND		33.3
Naphthalene	ND		33.3
4-Chloroaniline	ND		33.3
Hexachlorobutadiene	ND		33.3
Caprolactam	ND		33.3
4-Chloro-3-methylphenol	ND		33.3
2-Methylnaphthalene	ND		33.3
Hexachlorocyclopentadiene	ND		33.3
2,4,6-Trichlorophenol	ND		33.3
2,4,5-Trichlorophenol	ND		33.3
1-1'-Biphenyl	ND		33.3
2-Chloronaphthalene	ND		33.3
2-Nitroaniline	ND		33.3
Dimethylphthalate	ND		33.3
2,6-Dinitrotoluene	ND		33.3
Acenaphthylene	ND		33.3

INTEGRATED ANALYTICAL LABORATORIES

SEMIVOLATILE ORGANICS

Client/Project: N/A

Lab ID: METHOD-BLK

Client ID: .

Date Received: N/A

Date Extracted: 10/15/2001

Date Analyzed: 10/15/2001

Data file: B6842.D

GC/MS Column: DB-5

Sample wt/vol: 30.0g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 0.0

Compound	Concentration	Q	MDL
3-Nitroaniline	ND		33.3
Acenaphthene	ND		33.3
2,4-Dinitrophenol	ND		33.3
4-Nitrophenol	ND		33.3
2,4-Dinitrotoluene	ND		33.3
Dibenzofuran	ND		33.3
Diethylphthalate	ND		33.3
Fluorene	ND		33.3
4-Chlorophenyl-phenylether	ND		33.3
4-Nitroaniline	ND		33.3
4,6-Dinitro-2-methylphenol	ND		33.3
N-Nitrosodiphenylamine	ND		33.3
1,2-Diphenylhydrazine/Azobenzene	ND		33.3
4-Bromophenyl-phenylether	ND		33.3
Hexachlorobenzene	ND		33.3
Atrazine	ND		33.3
Pentachlorophenol	ND		33.3
Phenanthrene	ND		33.3
Anthracene	ND		33.3
Carbazole	ND		33.3
Di-n-butylphthalate	ND		33.3
Fluoranthene	ND		33.3
Benzidine	ND		33.3
Pyrene	ND		33.3
3,3'-Dimethylbenzidine	ND		33.3
Butylbenzylphthalate	ND		33.3
3,3'-Dichlorobenzidine	ND		33.3
Benzo[a]anthracene	ND		33.3
Chrysene	ND		33.3
bis(2-Ethylhexyl)phthalate	ND		33.3
Di-n-octylphthalate	ND		33.3
Benzo[b]fluoranthene	ND		33.3
Benzo[k]fluoranthene	ND		33.3
Benzo[a]pyrene	ND		33.3
Indeno[1,2,3-cd]pyrene	ND		33.3
Dibenz[a,h]anthracene	ND		33.3
Benzo[g,h,i]perylene	ND		33.3

Total Target Compounds: 0

INTEGRATED ANALYTICAL LABORATORIES

SEMIVOLATILE ORGANICS
Tentatively Identified Compounds

Client/Project: N/A

Lab ID: METHOD-BLK

Client ID: .

Date Received: N/A

Date Extracted: 10/15/2001

Date Analyzed: 10/15/2001

Data file: B6842.D

GC/MS Column: DB-5

Sample wt/vol: 30.0g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 1

% Moisture: 0.0

CAS #	Compound	Estimated Concentration	Retention Time
-------	----------	----------------------------	-------------------

No peaks detected

Total TICs = 0

Response Factor Report MSD_B

Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
 Title : BNA CALIBRATION METHOD
 Last Update : Mon Oct 08 11:06:23 2001
 Response via : Initial Calibration

Calibration Files

160 =B6623.D 80 =B6619.D 50 =B6621.D
 20 =B6620.D 120 =B6622.D

Compound		160	80	50	20	120	Avg	%RSD
1) I	1,4-Dichlorobenzene-d	-----ISTD-----						
2) T	N-Nitrosodimethylam	0.729	0.683	0.626	0.581	0.633	0.650	8.78
3) T	Pyridine	0.670	0.691	0.553	0.648	0.656	0.643	8.28
4) S	2-Fluorophenol	0.979	1.051	1.005	1.157	1.044	1.047	6.49
5) T	Benzaldehyde	2.378	1.884	2.198	2.206	2.694	2.272	13.01
6) S	Phenol-d5	1.404	1.324	1.353	1.440	1.375	1.379	3.26
7) MC	Phenol	1.332	1.271	1.208	1.289	1.232	1.267	3.84
8) T	Aniline	0.497	0.494	0.472	0.530	0.529	0.505	4.92
9) T	bis(2-Chloroethyl)e	0.689	0.643	0.640	0.687	0.683	0.669	3.65
10) M	2-Chlorophenol	1.138	0.984	0.944	1.005	1.102	1.035	7.90
11) T	1,3-Dichlorobenzene	1.562	1.354	1.196	1.214	1.381	1.341	11.05
12) MC	1,4-Dichlorobenzene	1.639	1.425	1.277	1.354	1.511	1.441	9.75
13) T	Benzyl alcohol	0.616	0.538	0.544	0.551	0.604	0.571	6.38
14) T	1,2-Dichlorobenzene	1.416	1.224	1.083	1.140	1.338	1.240	11.09
15) T	2-Methylphenol	1.025	0.876	0.848	0.914	0.948	0.922	7.47
16) T	bis(2-chloroisoprop	1.132	1.135	1.156	1.265	1.154	1.168	4.69
17) T	4-Methylphenol	1.069	0.859	0.817	0.834	1.032	0.922	12.88
18) MP	N-Nitroso-di-n-prop	0.664	0.650	0.610	0.642	0.680	0.649	4.06
19) T	Acetophenone	1.632	1.500	1.375	1.450	1.598	1.511	6.98
20) T	2-Aminotoluene +4-A	1.872	1.818	1.638	1.707	1.882	1.784	5.99
21) T	Hexachloroethane	0.521	0.456	0.406	0.446	0.491	0.464	9.51
22) I	Naphthalene-d8	-----ISTD-----						
23) S	Nitrobenzene-d5	0.438	0.430	0.469	0.452	0.436	0.445	3.52
24) T	Nitrobenzene	0.281	0.255	0.266	0.255	0.270	0.265	4.16
25) T	Isophorone	0.478	0.460	0.468	0.447	0.456	0.462	2.50
26) TC	2-Nitrophenol	0.169	0.138	0.134	0.104	0.165	0.142	18.61
27) T	2,4-Dimethylphenol	0.292	0.277	0.242	0.208	0.272	0.258	12.87
28) T	bis(2-Chloroethoxy)	0.335	0.310	0.306	0.283	0.325	0.312	6.38
29) T	Benzoic acid	0.067	0.041	0.049	0.036	0.054	0.049	24.33
30) T	2,4-Dimethylaniline	0.346	0.324	0.280	0.263	0.333	0.309	11.55
31) TC	2,4-Dichlorophenol	0.289	0.231	0.203	0.187	0.242	0.231	17.06
32) M	1,2,4-Trichlorobenz	0.347	0.305	0.266	0.234	0.322	0.295	15.36
33) T	Naphthalene	0.997	1.021	0.872	0.828	1.014	0.946	9.49
34) T	4-Chloroaniline	0.503	0.460	0.304	0.290	0.480	0.407	25.03
35) TC	Hexachlorobutadiene	0.229	0.190	0.163	0.139	0.209	0.186	19.35
36) T	Caprolactam	0.087	0.080	0.081	0.073	0.084	0.081	6.66
37) MC	4-Chloro-3-methylph	0.205	0.173	0.171	0.154	0.196	0.180	11.46
38) T	2-Methylnaphthalene	0.689	0.681	0.567	0.514	0.622	0.615	12.21
39) I	Acenaphthene-d10	-----ISTD-----						
40) TP	Hexachlorocyclopent	0.168	0.072	0.105	0.053	0.121	0.104	43.18
41) TC	2,4,6-Trichlorophen	0.332	0.251	0.239	0.201	0.317	0.268	20.55
42) T	2,4,5-Trichlorophen	0.394	0.310	0.332	0.278	0.366	0.336	13.60
43) S	2-Fluorobiphenyl	2.228	1.999	1.941	1.821	2.164	2.031	8.15
44) T	1-1'-Biphenyl	1.737	1.521	1.489	1.194	1.817	1.551	15.71
45) T	2-Chloronaphthalene	1.184	1.101	0.938	0.808	1.103	1.027	14.76
46) T	2-Nitroaniline	0.248	0.185	0.182	0.171	0.212	0.200	

48)	T	2,6-Dinitrotoluene	0.251	0.199	0.206	0.175	0.254	0.217	15.92
49)	T	Acenaphthylene	1.984	1.707	1.458	1.254	1.714	1.623	17.13
50)	T	3-Nitroaniline	0.365	0.281	0.274	0.246	0.334	0.300	16.14
51)	MC	Acenaphthene	1.321	1.029	0.973	0.790	1.206	1.064	19.41
52)	T	2,4-Dinitrophenol	0.068	0.038	0.036	0.026	0.055	0.045	36.88
53)	MP	4-Nitrophenol	0.069	0.075	0.095	0.067	0.057	0.073	18.96
54)	M	2,4-Dinitrotoluene	0.384	0.314	0.300	0.219	0.400	0.323	22.41
55)	T	Dibenzofuran	1.930	1.530	1.439	1.155	1.843	1.580	19.88
56)	T	Diethylphthalate	1.264	0.971	0.903	0.787	1.061	0.997	17.99
57)	T	Fluorene	1.372	1.112	1.098	0.900	1.340	1.164	16.67
58)	T	4-Chlorophenyl-phen	0.651	0.592	0.530	0.418	0.629	0.564	16.58
59)	T	4-Nitroaniline	0.257	0.212	0.207	0.163	0.230	0.214	15.99
60)	I	Phenanthrene-d10	-----ISTD-----						
61)	T	4,6-Dinitro-2-methy	0.104	0.054	0.048	0.035	0.075	0.063	42.46
62)	TC	N-Nitrosodiphenylam	0.541	0.443	0.391	0.374	0.474	0.445	15.04
63)	T	1,2-Diphenylhydraz	0.689	0.606	0.580	0.529	0.686	0.618	11.20
64)	S	2,4,6-Tribromopheno	0.111	0.098	0.117	0.100	0.107	0.107	7.55
65)	T	4-Bromophenyl-pheny	0.260	0.235	0.180	0.157	0.235	0.213	20.15
66)	T	Hexachlorobenzene	0.326	0.283	0.219	0.197	0.297	0.264	20.56
67)	T	Atrazine	0.199	0.146	0.133	0.133	0.164	0.155	17.83
68)	MC	Pentachlorophenol	0.063	0.058	0.060	0.039	0.056	0.055	17.03
69)	T	Phenanthrene	1.166	0.962	0.834	0.784	1.054	0.960	16.29
70)	T	Anthracene	1.231	1.001	0.815	0.730	1.053	0.966	20.54
71)	T	Carbazole	0.829	0.699	0.666	0.615	0.750	0.712	11.51
72)	T	Di-n-butylphthalate	1.015	0.829	0.779	0.662	0.923	0.842	16.05
73)	TC	Fluoranthene	0.846	0.789	0.716	0.624	0.834	0.762	12.15
74)	T	Benzidine	0.107	0.067	0.079	0.062	0.096	0.082	23.01
75)	I	Chrysene-d12	-----ISTD-----						
76)	M	Pyrene	1.260	1.171	1.123	1.009	1.197	1.152	8.15
77)	S	Terphenyl-d14	0.937	1.031	1.085	1.110	0.932	1.019	8.06
78)	T	3,3'-Dimethylbenzid	0.101	0.067	0.100	0.130	0.094	0.098	22.62
79)	T	Butylbenzylphthalat	0.486	0.357	0.371	0.306	0.431	0.390	17.81
80)	T	3,3'-Dichlorobenzid	0.262	0.214	0.243	0.215	0.250	0.237	9.05
81)	T	Benzo[a]anthracene	0.933	0.858	0.811	0.762	0.886	0.850	7.79
82)	T	Chrysene	0.873	0.827	0.780	0.786	0.852	0.823	4.92
83)	T	bis(2-Ethylhexyl)ph	0.628	0.471	0.498	0.417	0.581	0.519	16.32
84)	I	Perylene-d12	-----ISTD-----						
85)	TC	Di-n-octylphthalate	0.914	0.670	0.749	0.653	0.828	0.763	14.35
86)	T	Benzo[b]fluoranthene	0.959	0.714	0.778	0.648	1.038	0.827	19.97
87)	T	Benzo[k]fluoranthene	1.142	1.161	1.086	1.063	1.192	1.129	4.74
88)	TC	Benzo[a]pyrene	0.957	0.841	0.817	0.792	0.908	0.863	7.89
89)	T	Indeno[1,2,3-cd]pyr	1.228	1.051	0.974	0.867	1.178	1.059	13.93
90)	T	Dibenz[a,h]anthracene	1.069	0.883	0.847	0.722	1.018	0.908	15.27
91)	T	Benzo[g,h,i]perylene	0.985	0.894	0.807	0.695	0.935	0.863	13.25

(#) = Out of Range

BS2301.M

Mon Oct 08 11:06:33 2001 MSD_B

Instrument ID: MSD_B
Method ID: BS2301.M
Date: 10/08/2001

Average %RSD = 14.37

Refer to SW846 Method 8000B Section 7.5.1.

Evaluate Continuing Calibration Report

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6835.D

Acq On : 15 Oct 2001 8:56

Sample : .,80-bna-STD-INJ-FOR-10/12/01

Misc :

MS Integration Params: rteint.p

Vial: 2

Operator: JC

Inst : MSD_B

Multiplr: 1.00

Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)

Title : BNA CALIBRATION METHOD

Last Update : Mon Oct 08 11:06:23 2001

Response via : Multiple Level Calibration

	Compound	AvgRF	CCRF	%Dev	Area%	Dev(min)
1 I	1,4-Dichlorobenzene-d4	1.000	1.000	0.0	151	0.00
2 T	N-Nitrosodimethylamine	0.650	0.745	-14.6	165	0.02
3 T	Pyridine	0.643	0.740	-15.1	162	0.02
4 S	2-Fluorophenol	1.047	1.187	-13.4	171	0.00
5 T	Benzaldehyde	2.272	2.299	-1.2	130	0.00
6 S	Phenol-d5	1.379	1.318	4.4	151	0.00
7 MC	Phenol	1.267	1.442	-13.8	172	0.00
8 T	Aniline	0.505	0.474	6.1	145	0.00
9 T	bis(2-Chloroethyl)ether	0.669	0.695	-3.9	163	0.00
10 M	2-Chlorophenol	1.035	1.054	-1.8	162	0.00
11 T	1,3-Dichlorobenzene	1.341	1.362	-1.6	152	0.00
12 MC	1,4-Dichlorobenzene	1.441	1.325	8.0	141	0.00
13 T	Benzyl alcohol	0.571	0.578	-1.2	163	0.00
14 T	1,2-Dichlorobenzene	1.240	1.172	5.5	145	0.00
15 T	2-Methylphenol	0.922	0.889	3.6	154	0.00
16 T	bis(2-chloroisopropyl) ether	1.168	1.228	-5.1	164	0.00
17 T	4-Methylphenol	0.922	0.971	-5.3	171	0.00
18 MP	N-Nitroso-di-n-propylamine	0.649	0.594	8.5	138	0.00
19 T	Acetophenone	1.511	1.359	10.1	137	0.00
20 T	2-Aminotoluene +4-Aminotolu	1.784	1.530	14.2	127	0.00
21 T	Hexachloroethane	0.464	0.485	-4.5	161	0.00
22 I	Naphthalene-d8	1.000	1.000	0.0	141	0.00
23 S	Nitrobenzene-d5	0.445	0.520	-16.9	171	0.00
24 T	Nitrobenzene	0.265	0.279	-5.3	155	0.00
25 T	Isophorone	0.462	0.469	-1.5	144	0.00
26 TC	2-Nitrophenol	0.142	0.161	-13.4	166	0.00
27 T	2,4-Dimethylphenol	0.258	0.269	-4.3	137	0.00
28 T	bis(2-Chloroethoxy)methane	0.312	0.307	1.6	140	0.00
29 T	Benzoic acid	0.049	0.045	8.2	154	0.02
30 T	2,4-Dimethylaniline	0.309	0.283	8.4	123	0.00
31 TC	2,4-Dichlorophenol	0.231	0.218	5.6	133	0.00
32 M	1,2,4-Trichlorobenzene	0.295	0.273	7.5	127	0.00
33 T	Naphthalene	0.946	0.983	-3.9	136	0.00
34 T	4-Chloroaniline	0.407	0.284	30.2	88	0.00
35 TC	Hexachlorobutadiene	0.186	0.185	0.5	138	0.00
36 T	Caprolactam	0.081	0.091	-12.3	161	0.02
37 MC	4-Chloro-3-methylphenol	0.180	0.180	0.0	147	0.00
38 T	2-Methylnaphthalene	0.615	0.584	5.0	121	0.00
39 I	Acenaphthene-d10	1.000	1.000	0.0	112	0.00
40 TP	Hexachlorocyclopentadiene	0.104	0.085	18.3	132	0.00
41 TC	2,4,6-Trichlorophenol	0.268	0.262	2.2	117	0.00
42 T	2,4,5-Trichlorophenol	0.336	0.376	-11.9	136	0.00
43 S	2-Fluorobiphenyl	2.031	2.043	-0.6	115	0.00
44 T	1-1'-Biphenyl	1.551	1.810	-16.7	133	0.00
45 T	2-Chloronaphthalene	1.027	1.177	-14.6	120	0.00

46 T	2-Nitroaniline	0.204	0.241	-18.1	139	0.01
47 T	Dimethylphthalate	1.020	1.092	-7.1	129	0.00
48 T	2,6-Dinitrotoluene	0.217	0.225	-3.7	127	0.00
49 T	Acenaphthylene	1.623	1.647	-1.5	108	0.00
50 T	3-Nitroaniline	0.300	0.334	-11.3	133	0.00
51 MC	Acenaphthene	1.064	1.157	-8.7	126	0.00
52 T	2,4-Dinitrophenol	0.045	0.049	-8.9	146	0.00
53 MP	4-Nitrophenol	0.073	0.065	11.0	97	0.01
54 M	2,4-Dinitrotoluene	0.323	0.322	0.3	115	0.00
55 T	Dibenzofuran	1.580	1.634	-3.4	120	0.00
56 T	Diethylphthalate	0.997	0.973	2.4	112	0.00
57 T	Fluorene	1.164	1.180	-1.4	119	0.00
58 T	4-Chlorophenyl-phenylether	0.564	0.551	2.3	104	0.00
59 T	4-Nitroaniline	0.214	0.198	7.5	104	0.01
60 I	Phenanthrene-d10	1.000	1.000	0.0	109	0.00
61 T	4,6-Dinitro-2-methylphenol	0.063	0.061	3.2	124	0.01
62 TC	N-Nitrosodiphenylamine	0.445	0.468	-5.2	115	0.01
63 T	1,2-Diphenylhydrazine/Azobe	0.618	0.709	-14.7	127	0.00
64 S	2,4,6-Tribromophenol	0.107	0.125	-16.8	139	0.00
65 T	4-Bromophenyl-phenylether	0.213	0.205	3.8	95	0.00
66 T	Hexachlorobenzene	0.264	0.277	-4.9	106	0.00
67 T	Atrazine	0.155	0.134	13.5	100	0.01
68 MC	Pentachlorophenol	0.055	0.061	-10.9	115	0.01
69 T	Phenanthrene	0.960	0.911	5.1	103	0.00
70 T	Anthracene	0.966	0.959	0.7	104	0.01
71 T	Carbazole	0.712	0.675	5.2	105	0.00
72 T	Di-n-butylphthalate	0.842	0.901	-7.0	118	0.00
73 TC	Fluoranthene	0.762	0.733	3.8	101	0.01
74 T	Benzidine	0.082	0.067	18.3	108	0.00
75 I	Chrysene-d12	1.000	1.000	0.0	92	0.02
76 M	Pyrene	1.152	1.218	-5.7	96	0.01
77 S	Terphenyl-d14	1.019	1.057	-3.7	94	0.01
78 T	3,3'-Dimethylbenzidine	0.098	0.096	2.0	132	0.00
79 T	Butylbenzylphthalate	0.390	0.373	4.4	96	0.01
80 T	3,3'-Dichlorobenzidine	0.237	0.266	-12.2	114	0.01
81 T	Benzo[a]anthracene	0.850	0.899	-5.8	96	0.01
82 T	Chrysene	0.823	0.848	-3.0	94	0.02
83 T	bis(2-Ethylhexyl)phthalate	0.519	0.477	8.1	93	0.01
84 I	Perylene-d12	1.000	1.000	0.0	103	0.02
85 TC	Di-n-octylphthalate	0.763	0.805	-5.5	124	0.01
86 T	Benzo[b]fluoranthene	0.827	0.886	-7.1	128	0.02
87 T	Benzo[k]fluoranthene	1.129	1.051	6.9	93	0.03
88 TC	Benzo[a]pyrene	0.863	0.881	-2.1	108	0.02
89 T	Indeno[1,2,3-cd]pyrene	1.059	1.164	-9.9	114	0.07
90 T	Dibenz[a,h]anthracene	0.908	1.028	-13.2	120	0.06
91 T	Benzo[g,h,i]perylene	0.863	0.907	-5.1	105	0.08

(#) = Out of Range

B6619.D BS2301.M

SPCC's out = 0 CCC's out = 0
Mon Oct 15 11:24:17 2001 MSD_B

SEMIVOLATILE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/15/2001

Lab Sample ID	Matrix	File ID	S1	S2	S3	S4	S5	S6
6844-003	SOIL	B6837.D	N/A	N/A	55	75	N/A	77
6844-001	SOIL	B6838.D	N/A	N/A	96	84	N/A	82
6844-005	SOIL	B6839.D	N/A	N/A	48	71	N/A	69
6844-004	SOIL	B6840.D	N/A	N/A	98	77	N/A	77
6844-008	SOIL	B6841.D	N/A	N/A	93	75	N/A	71
METHOD-BLK	SOIL	B6842.D	77	85	84	65	101	91
7058-001	SOIL	B6843.D	N/A	N/A	66	47	N/A	78
7058-002	SOIL	B6844.D	N/A	N/A	63	45	N/A	77
7058-003	SOIL	B6845.D	N/A	N/A	67	45	N/A	76
7058-004	SOIL	B6846.D	N/A	N/A	66	46	N/A	71
7058-005	SOIL	B6847.D	N/A	N/A	64	45	N/A	77
7058-006	SOIL	B6848.D	N/A	N/A	60	41	N/A	67
7058-007	SOIL	B6849.D	N/A	N/A	63	44	N/A	70
7058-008	SOIL	B6850.D ¹	N/A	N/A	64	45	N/A	71
7058-009	SOIL	B6851.D	N/A	N/A	69	48	N/A	77
7058-010	SOIL	B6852.D	N/A	N/A	69	49	N/A	79
7058-011	SOIL	B6853.D	N/A	N/A	75	51	N/A	80
7058-012	SOIL	B6854.D	N/A	N/A	72	51	N/A	80
7085-004	SOIL	B6855.D	N/A	N/A	62	44	N/A	80
7085-005	SOIL	B6856.D	N/A	N/A	58	47	N/A	61
7085-006	SOIL	B6857.D	N/A	N/A	71	49	N/A	73
7085-007	SOIL	B6858.D	N/A	N/A	72	50	N/A	74
7085-008	SOIL	B6859.D	N/A	N/A	62	44	N/A	71

	<u>Soil</u>	<u>Aqueous</u>
S1 (2FP) = 2-Fluorophenol	28-100	47-93
S2 (PHL) = Phenol-d5	28-102	46-110
S3 (NBZ) = Nitrobenzene-d5	35-101	42-120
S4 (FBP) = 2-Fluorobiphenyl	37-103	55-109
S5 (TBP) = 2,4,6-Tribromophenol	44-121	46-119
S6 (TPH) = Terphenyl-d14	40-120	51-118

* Column to be used to flag recovery values

SEMIVOLATILE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/15/2001

Lab Sample ID	Matrix	File ID	S1	S2	S3	S4	S5	S6
7035-001	SOIL	B6860.D	65	74	70	49	88	66
7041-001	SOIL	B6861.D	70	72	65	46	94	70
7061-001		B6862.D	76	77	75	53	98	68
7058-001MS	SOIL	B6863.D	82	86	71	56	107	83
7058-001MSD	SOIL	B6864.D	80	89	77	59	112	85
LCS	SOIL	B6865.D	72	79	71	51	96	78

	<u>Soil</u>	<u>Aqueous</u>
S1 (2FP) = 2-Fluorophenol	28-100	47-93
S2 (PHL) = Phenol-d5	28-102	46-110
S3 (NBZ) = Nitrobenzene-d5	35-101	42-120
S4 (FBP) = 2-Fluorobiphenyl	37-103	55-109
S5 (TBP) = 2,4,6-Tribromophenol	44-121	46-119
S6 (TPH) = Terphenyl-d14	40-120	51-118

* Column to be used to flag recovery values

SOIL SEMIVOLATILE MATRIX SPIKE/SPIKE DUPLICATE RECOVERY

Matrix spike Lab sample ID:

7058-001

Batch No.:

B101501S1

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
Phenol	100.0	0.0	80.9	81	26 - 90
2-Chlorophenol	100.0	0.0	80.6	81	25 - 102
1,4-Dichlorobenzene	50.0	0.0	29.6	59	28 - 104
N-Nitroso-di-n-propylamine	50.0	0.0	36.4	73	41 - 126
1,2,4-Trichlorobenzene	50.0	0.0	28.6	57	38 - 107
4-Chloro-3-methylphenol	100.0	0.0	82.1	82	26 - 103
Acenaphthene	50.0	0.0	33.7	67	31 - 137
4-Nitrophenol	100.0	0.0	100.6	101	11 - 114
2,4-Dinitrotoluene	50.0	0.0	30.1	60	28 - 89
Pentachlorophenol	100.0	0.0	108.4	108	17 - 109
Pyrene	50.0	0.0	37.7	75	35 - 142

Compound	SAMPLE CONC. (ug/Kg)	MSD CONC. (ug/Kg)	MSD % # REC	% RPD #	QC LIMITS	
					RPD	REC.
Phenol	0.0	89.1	89	9	42	26 - 90
2-Chlorophenol	0.0	84.3	84	4	40	25 - 102
1,4-Dichlorobenzene	0.0	30.1	60	2	28	28 - 104
N-Nitroso-di-n-propylamine	0.0	36.0	72	1	38	41 - 126
1,2,4-Trichlorobenzene	0.0	29.5	59	3	28	38 - 107
4-Chloro-3-methylphenol	0.0	91.0	91	10	42	26 - 103
Acenaphthene	0.0	33.2	66	2	31	31 - 137
4-Nitrophenol	0.0	111.4	111	9	50	11 - 114
2,4-Dinitrotoluene	0.0	29.6	59	2	38	28 - 89
Pentachlorophenol	0.0	107.7	108	0	50	17 - 109
Pyrene	0.0	36.4	73	3	31	35 - 142

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

RPD: 0 out of 11 outside limits

Spike Recovery: 0 out of 22 outside limits

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6619.D

Date Analyzed: 10/08/2001

Instrument ID: MSDB

Time Analyzed: 07:54

40UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
12 HOUR STD	112820	2.42	381902	3.06	209421	3.95
UPPER LIMIT	225640	2.92	763804	3.56	418842	4.45
LOWER LIMIT	56410	1.92	190951	2.56	104711	3.45
LAB SAMPLE ID						
01 20-bna-STD-INJ-FOR	113341	2.42	446092	3.05	249915	3.95
02 50-bna-STD-INJ-FOR	116736	2.42	390650	3.05	209832	3.95
03 120-bna-STD-INJ-FOR	105206	2.42	358996	3.05	182330	3.95
04 160-bna-STD-INJ-FOR	102526	2.42	345683	3.05	171216	3.95
05 80-OLMO4-STD-INJ-F	100195	2.42	379090	3.05	208299	3.94
06 20-OLMO4-STD-INJ-F	88491	2.42	338641	3.05	178258	3.95
07 50-OLMO4-STD-INJ-F	91447	2.42	348173	3.05	178202	3.94
08 120-OLMO4-STD-INJ	95175	2.42	358986	3.05	181169	3.94
09 160-OLMO4-STD-INJ	92636	2.42	332914	3.05	169316	3.94
10 80-OLMO4-STD-INJ-F	109023	2.42	414066	3.05	206190	3.94
11 6683-003	158020	2.42	603319	3.05	337180	3.95
12 6677-003	98728	2.42	371543	3.05	195218	3.94
13 6677-004	98027	2.42	379471	3.05	195318	3.94
14 METHOD-BLK	96903	2.42	361718	3.05	195443	3.94
15 6677-004MS	103725	2.42	389058	3.05	205814	3.94
16 6677-004MSD	108853	2.42	401605	3.05	220417	3.94
17 LCS	115042	2.42	425439	3.05	247915	3.94
18 METHOD-BLK	106903	2.42	403747	3.05	220494	3.94
19 6764-002	115275	2.42	435434	3.05	228705	3.94
20 6764-004	108919	2.42	419440	3.05	220136	3.94
21 6764-005	119730	2.42	459951	3.05	227995	3.94
22 6764-006	117056	2.42	447436	3.05	224717	3.94

IS1 = 1,4-Dichlorobenzene-d4

IS2 = Naphthalene-d8

IS3 = Acenaphthene-d10

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6619.D

Date Analyzed: 10/08/2001

Instrument ID: MSDB

Time Analyzed: 07:54

40UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
12 HOUR STD	112820	2.42	381902	3.06	209421	3.95
UPPER LIMIT	225640	2.92	763804	3.56	418842	4.45
LOWER LIMIT	56410	1.92	190951	2.56	104711	3.45
LAB SAMPLE ID						
01 6764-007	111284	2.42	414019	3.05	214932	3.94
02 6764-008	111661	2.42	404600	3.05	196976	3.94
03 6442-001	102797	2.42	394159	3.05	200548	3.94
04 6793-001	99211	2.42	383927	3.05	197655	3.94
05 6793-002	99239	2.42	386399	3.05	191389	3.94
06 6793-003	99633	2.42	369219	3.05	186975	3.94
07 6793-005	101382	2.42	385854	3.05	195345	3.94
08 6793-007	105049	2.42	396574	3.05	201700	3.94
09 6793-008	100979	2.42	382501	3.05	191904	3.94
10 6793-008MS	113088	2.42	415449	3.05	210670	3.94
11 6793-008MSD	115444	2.42	421207	3.05	212728	3.94
12 LCS	111972	2.42	406208	3.05	208789	3.94
13 6666-021	99362	2.42	368508	3.05	184248	3.94
14 6666-022	88843	2.42	325473	3.05	156859	3.94
15 6666-023	82212	2.43	311258	3.05	154538	3.94
16						
17						
18						
19						
20						
21						
22						

IS1 = 1,4-Dichlorobenzene-d4

IS2 = Naphthalene-d8

IS3 = Acenaphthene-d10

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6619.D

Date Analyzed: 10/08/2001

Instrument ID: MSDB

Time Analyzed: 07:54

40UG/L	IS4 AREA #	RT #	IS5 AREA #	RT #	IS6 AREA #	RT #
12 HOUR STD	309992	4.74	191236	6.56	165723	7.77
UPPER LIMIT	619984	5.24	382472	7.06	331446	8.27
LOWER LIMIT	154996	4.24	95618	6.06	82862	7.27
LAB SAMPLE ID						
01 20-bna-STD-INJ-FOR	391589	4.74	216719	6.56	171810	7.78
02 50-bna-STD-INJ-FOR	338588	4.73	204527	6.55	165251	7.79
03 120-bna-STD-INJ-FOR	288896	4.74	187424	6.56	167415	7.77
04 160-bna-STD-INJ-FOR	277815	4.74	186632	6.56	169910	7.77
05 80-OLMO4-STD-INJ-F	347153	4.73	220214	6.55	142782	7.80
06 20-OLMO4-STD-INJ-F	308909	4.73	194066	6.57	148446	7.78
07 50-OLMO4-STD-INJ-F	300793	4.73	189688	6.56	141414	7.78
08 120-OLMO4-STD-INJ	318154	4.73	197817	6.56	142552	7.78
09 160-OLMO4-STD-INJ	287156	4.73	185241	6.56	140109	7.78
10 80-OLMO4-STD-INJ-F	347744	4.73	212022	6.55	158546	7.80
11 6683-003	562886	4.73	327593	6.55	206466	7.79
12 6677-003	302705	4.73	181059	6.56	94899	7.77
13 6677-004	319218	4.73	176311	6.55	112730	7.79
14 METHOD-BLK	318797	4.73	202850	6.55	163970	7.80
15 6677-004MS	319909	4.73	176514	6.55	114176	7.80
16 6677-004MSD	343266	4.73	183593	6.55	124466	7.79
17 LCS	376093	4.73	225564	6.55	181484	7.79
18 METHOD-BLK	367222	4.73	224553	6.55	170792	7.80
19 6764-002	360652	4.73	211351	6.55	181629	7.77
20 6764-004	357856	4.73	216974	6.55	174569	7.79
21 6764-005	357891	4.73	208526	6.55	181476	7.79
22 6764-006	349740	4.73	208506	6.55	174924	7.79

IS4 = Phenanthrene-d10

IS5 = Chrysene-d12

IS6 = Perylene-d12

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6619.D

Date Analyzed: 10/08/2001

Instrument ID: MSDB

Time Analyzed: 07:54

50UG/L	IS4 AREA #	RT #	IS5 AREA #	RT #	IS6 AREA #	RT #
12 HOUR STD	309992	4.74	191236	6.56	165723	7.77
UPPER LIMIT	619984	5.24	382472	7.06	331446	8.27
LOWER LIMIT	154996	4.24	95618	6.06	82862	7.27
LAB SAMPLE ID						
01 6764-007	328899	4.73	200880	6.55	165209	7.79
02 6764-008	298250	4.73	183299	6.55	174245	7.79
03 6442-001	301303	4.73	173962	6.55	169315	7.79
04 6793-001	307010	4.73	191723	6.56	183181	7.77
05 6793-002	304025	4.73	194087	6.56	182430	7.77
06 6793-003	290374	4.73	191967	6.55	171146	7.79
07 6793-005	303483	4.73	195428	6.55	170108	7.79
08 6793-007	306226	4.73	198108	6.55	174939	7.79
09 6793-008	285087	4.73	193103	6.55	169219	7.79
10 6793-008MS	320419	4.73	194205	6.55	170526	7.79
11 6793-008MSD	312650	4.73	191462	6.55	170581	7.79
12 LCS	306186	4.73	191734	6.55	163934	7.79
13 6666-021	275462	4.73	214376	6.56	206910	7.77
14 6666-022	237928	4.73	199717	6.55	200010	7.79
15 6666-023	244462	4.73	203495	6.55	204690	7.79
16						
17						
18						
19						
20						
21						
22						

IS4 = Phenanthrene-d10

IS5 = Chrysene-d12

IS6 = Perylene-d12

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6835.D

Date Analyzed: 10/15/2001

Instrument ID: MSDB

Time Analyzed: 08:56

40UG/L	IS1 AREA #	RT #	IS2 AREA #	RT #	IS3 AREA #	RT #
12 HOUR STD	170820	2.42	540031	3.05	234737	3.95
UPPER LIMIT	341640	2.92	1080062	3.55	469474	4.45
LOWER LIMIT	85410	1.92	270016	2.55	117369	3.45
LAB SAMPLE ID						
01 80-OLMO4-STD-INJ-F	145164	2.42	532928	3.05	258341	3.95
02 6844-003	137556	2.42	527685	3.05	243743	3.95
03 6844-001	133443	2.42	514084	3.05	236748	3.94
04 6844-005	114740	2.42	426478	3.05	199276	3.94
05 6844-004	137096	2.42	492491	3.05	218545	3.94
06 6844-008	113518	2.42	436050	3.05	201707	3.95
07 METHOD-BLK	159276	2.42	593009	3.05	278510	3.95
08 7058-001	124056	2.42	479147	3.05	216681	3.94
09 7058-002	111637	2.42	434508	3.05	210160	3.95
10 7058-003	122758	2.42	462740	3.05	215323	3.94
11 7058-004	110101	2.42	405709	3.05	191557	3.94
12 7058-005	132913	2.42	483639	3.05	225774	3.94
13 7058-006	115183	2.42	452274	3.05	217492	3.94
14 7058-007	120454	2.42	452145	3.05	207275	3.94
15 7058-008	122651	2.42	461511	3.05	219453	3.94
16 7058-009	124654	2.42	458096	3.05	214928	3.94
17 7058-010	139805	2.42	510113	3.05	232073	3.94
18 7058-011	127418	2.42	479372	3.05	228707	3.94
19 7058-012	131950	2.42	478363	3.05	223040	3.94
20 7085-004	130010	2.42	481717	3.05	221081	3.94
21 7085-005	130997	2.42	514187	3.05	191035	3.95
22 7085-006	119084	2.42	445148	3.05	207631	3.94

IS1 = 1,4-Dichlorobenzene-d4

IS2 = Naphthalene-d8

IS3 = Acenaphthene-d10

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6835.D

Date Analyzed: 10/15/2001

Instrument ID: MSDB

Time Analyzed: 08:56

40UG/L		IS1		IS2		IS3	
		AREA	#	RT	#	AREA	#
12 HOUR STD		170820		2.42		540031	3.05
UPPER LIMIT		341640		2.92		1080062	3.55
LOWER LIMIT		85410		1.92		270016	2.55
LAB SAMPLE ID							
01	7085-007	131139		2.42		482651	3.05
02	7085-008	133387		2.42		483836	3.05
03	7035-001	116692		2.42		406851	3.05
04	7041-001	141047		2.42		483709	3.05
05	7061-001	113456		2.42		382353	3.05
06	7058-001MS	126837		2.42		452948	3.05
07	7058-001MSD	110206		2.42		382844	3.05
08	LCS	118493		2.42		399297	3.05
09							
10							
11							
12							
13							
14							
15							
16							
17							
18							
19							
20							
21							
22							

IS1 = 1,4-Dichlorobenzene-d4

IS2 = Naphthalene-d8

IS3 = Acenaphthene-d10

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6835.D

Date Analyzed: 10/15/2001

Instrument ID: MSDB

Time Analyzed: 08:56

40UG/L	IS4 AREA #	RT #	IS5 AREA #	RT #	IS6 AREA #	RT #
12 HOUR STD	336494	4.73	175811	6.56	171148	7.81
UPPER LIMIT	672988	5.23	351622	7.06	342296	8.31
LOWER LIMIT	168247	4.23	87906	6.06	85574	7.31
LAB SAMPLE ID						
01 80-OLMO4-STD-INJ-F	401005	4.73	205295	6.56	148277	7.80
02 6844-003	344905	4.73	181240	6.55	182307	7.80
03 6844-001	334918	4.73	193744	6.55	208479	7.80
04 6844-005	289056	4.73	171299	6.56	180849	7.78
05 6844-004	303447	4.73	181800	6.55	188776	7.80
06 6844-008	280742	4.73	186128	6.56	194065	7.80
07 METHOD-BLK	376072	4.73	230179	6.55	247210	7.80
08 7058-001	285812	4.73	175390	6.55	196024	7.80
09 7058-002	292416	4.73	179353	6.55	203681	7.79
10 7058-003	279112	4.73	164036	6.55	185281	7.79
11 7058-004	263960	4.74	166920	6.56	186463	7.78
12 7058-005	293387	4.73	166775	6.55	191878	7.79
13 7058-006	294042	4.73	172471	6.55	191038	7.79
14 7058-007	276055	4.73	166802	6.55	188063	7.80
15 7058-008	295095	4.73	169909	6.55	188028	7.80
16 7058-009	288212	4.73	175426	6.55	196673	7.79
17 7058-010	294349	4.73	171853	6.55	191451	7.80
18 7058-011	294573	4.73	169421	6.55	191961	7.79
19 7058-012	285287	4.73	167931	6.55	186777	7.80
20 7085-004	280616	4.73	164406	6.55	189039	7.80
21 7085-005	243476	4.74	151591	6.57	179228	7.78
22 7085-006	282828	4.73	173844	6.56	190625	7.78

IS4 = Phenanthrene-d10

IS5 = Chrysene-d12

IS6 = Perylene-d12

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

SEMIVOLATILE INTERNAL STANDARD AREA AND RT SUMMARY

Lab File ID (Standard): B6835.D

Date Analyzed: 10/15/2001

Instrument ID: MSDB

Time Analyzed: 08:56

50UG/L	IS4 AREA #	RT #	IS5 AREA #	RT #	IS6 AREA #	RT #
12 HOUR STD	336494	4.73	175811	6.56	171148	7.81
UPPER LIMIT	672988	5.23	351622	7.06	342296	8.31
LOWER LIMIT	168247	4.23	87906	6.06	85574	7.31
LAB SAMPLE ID						
01 7085-007	275767	4.73	171175	6.55	189403	7.8
02 7085-008	267387	4.74	176454	6.56	197769	7.77
03 7035-001	253757	4.73	169856	6.55	196953	7.8
04 7041-001	270562	4.73	175124	6.56	197006	7.78
05 7061-001	211834	4.73	133934	6.55	154696	7.8
06 7058-001MS	248461	4.73	151555	6.56	175142	7.81
07 7058-001MSD	226523	4.73	147096	6.56	170042	7.8
08 LCS	233247	4.73	148313	6.56	174372	7.8
09						
10						
11						
12						
13						
14						
15						
16						
17						
18						
19						
20						
21						
22						

IS4 = Phenanthrene-d10

IS5 = Chrysene-d12

IS6 = Perylene-d12

AREA UPPER LIMIT = +100% of internal standard area

AREA LOWER LIMIT = -50% of internal standard area

RT UPPER LIMIT = +0.50 minutes of internal standard RT

RT LOWER LIMIT = -0.50 minutes of internal standard RT

Column used to flag values outside QC limits with an asterisk

* Values outside of QC limits.

Quantitation Report (QT Reviewed)

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6861.D Vial: 28
Acq On : 15 Oct 2001 16:15 Operator: JC
Sample : 5-F1,7041-001,S,10.39g,9.40,10/15/01 Inst : MSD_B
Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00
MS Integration Params: rteint.p
Quant Time: Oct 15 16:25:30 2001 Quant Results File: BS2301.RES

Quant Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
Title : BNA CALIBRATION METHOD
Last Update : Mon Oct 08 11:06:23 2001
Response via : Initial Calibration
DataAcq Meth : BS2301

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) 1,4-Dichlorobenzene-d4	2.42	152	141047	40.00	UG	0.00
22) Naphthalene-d8	3.05	136	483709	40.00	UG	0.00
39) Acenaphthene-d10	3.94	164	212144	40.00	UG	0.00
60) Phenanthrene-d10	4.73	188	270562	40.00	UG	0.00
75) Chrysene-d12	6.56	240	175124	40.00	UG	0.02
84) Perylene-d12	7.78	264	197006	40.00	UG	-0.02

System Monitoring Compounds

4) 2-Fluorophenol	1.85	112	256734	69.52	UG	0.00
Spiked Amount	100.000	Range	28 - 100	Recovery	=	69.52%
6) Phenol-d5	2.24	99	350311	72.03	UG	0.00
Spiked Amount	100.000	Range	28 - 102	Recovery	=	72.03%
23) Nitrobenzene-d5	2.70	82	175840	32.68	UG	0.00
Spiked Amount	50.000	Range	35 - 101	Recovery	=	65.36%
43) 2-Fluorobiphenyl	3.59	172	248670	23.09	UG	0.00
Spiked Amount	50.000	Range	37 - 103	Recovery	=	46.18%
64) 2,4,6-Tribromophenol	4.36	330	67445	93.54	UG	0.00
Spiked Amount	100.000	Range	44 - 121	Recovery	=	93.54%
77) Terphenyl-d14	5.70	244	156517	35.09	UG	0.02
Spiked Amount	50.000	Range	40 - 120	Recovery	=	70.18%

Target Compounds

Qvalue

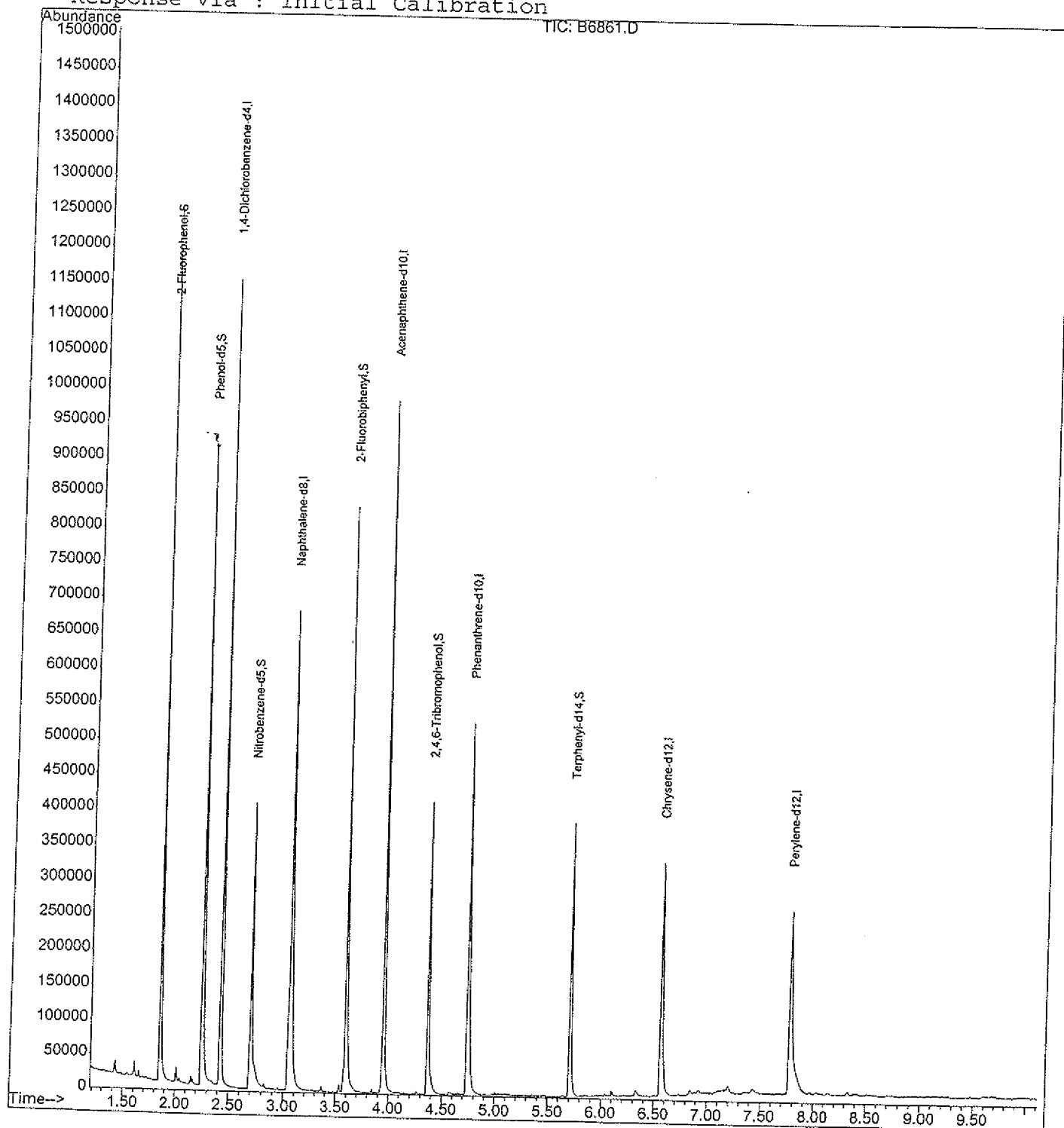
(#) = qualifier out of range (m) = manual integration

Quantitation Report (QT Reviewed)

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6861.D Vial: 28
 Acq On : 15 Oct 2001 16:15 Operator: JC
 Sample : 5-F1,7041-001,S,10.39g,9.40,10/15/01 Inst : MSD_B
 Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00
 MS Integration Params: rteint.p
 Quant Time: Oct 16 8:18 2001

Quant Results File: BS2301.RES

Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
 Title : BNA CALIBRATION METHOD
 Last Update : Mon Oct 08 11:06:23 2001
 Response via : Initial Calibration



Library Search Compound Report

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6861.D Vial: 28
Acq On : 15 Oct 2001 16:15 Operator: JC
Sample : 5-F1,7041-001,S,10.39g,9.40,10/15/01 Inst : MSD_B
Misc : BRENNAN/99704 FINETEX,10/11/01,10/11/01, Multiplr: 1.00
MS Integration Params: LSCINT.P
Quant Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
Title : BNA CALIBRATION METHOD
Library : C:\DATABASE\NIST98.L

No Library Search Compounds Detected

B6861.D BS2301.M Tue Oct 16 09:26:36 2001 MSD_B

Quantitation Report (QT Reviewed)

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6842.D
 Acq On : 15 Oct 2001 11:07
 Sample : .,METHOD-BLK,S,30.0g,0.0,10/15/01
 Misc : N/A,N/A,N/A,1
 MS Integration Params: rteint.p
 Quant Time: Oct 15 11:17:36 2001

Vial: 9
 Operator: JC
 Inst : MSD_B
 Multiplr: 1.00

Quant Results File: BS2301.RES

Quant Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
 Title : BNA CALIBRATION METHOD
 Last Update : Mon Oct 08 11:06:23 2001
 Response via : Initial Calibration
 DataAcq Meth : BS2301

Internal Standards	R.T.	QIon	Response	Conc	Units	Dev(Min)
1) 1,4-Dichlorobenzene-d4	2.42	152	159276	40.00	UG	0.00
22) Naphthalene-d8	3.05	136	593009	40.00	UG	0.00
39) Acenaphthene-d10	3.95	164	278510	40.00	UG	0.00
60) Phenanthrene-d10	4.73	188	376072	40.00	UG	0.00
75) Chrysene-d12	6.55	240	230179	40.00	UG	0.00
84) Perylene-d12	7.80	264	247210	40.00	UG	0.00

System Monitoring Compounds

4) 2-Fluorophenol	1.84	112	323163	77.49	UG	0.00
Spiked Amount 100.000	Range 28 - 100		Recovery =	77.49%		
6) Phenol-d5	2.24	99	465410	84.74	UG	0.00
Spiked Amount 100.000	Range 28 - 102		Recovery =	84.74%		
23) Nitrobenzene-d5	2.70	82	278635	42.24	UG	0.00
Spiked Amount 50.000	Range 35 - 101		Recovery =	84.48%		
43) 2-Fluorobiphenyl	3.58	172	460403	32.56	UG	0.00
Spiked Amount 50.000	Range 37 - 103		Recovery =	65.12%		
64) 2,4,6-Tribromophenol	4.36	330	101440	101.22	UG	0.00
Spiked Amount 100.000	Range 44 - 121		Recovery =	101.22%		
77) Terphenyl-d14	5.69	244	265516	45.28	UG	0.00
Spiked Amount 50.000	Range 40 - 120		Recovery =	90.56%		

Target Compounds

Qvalue

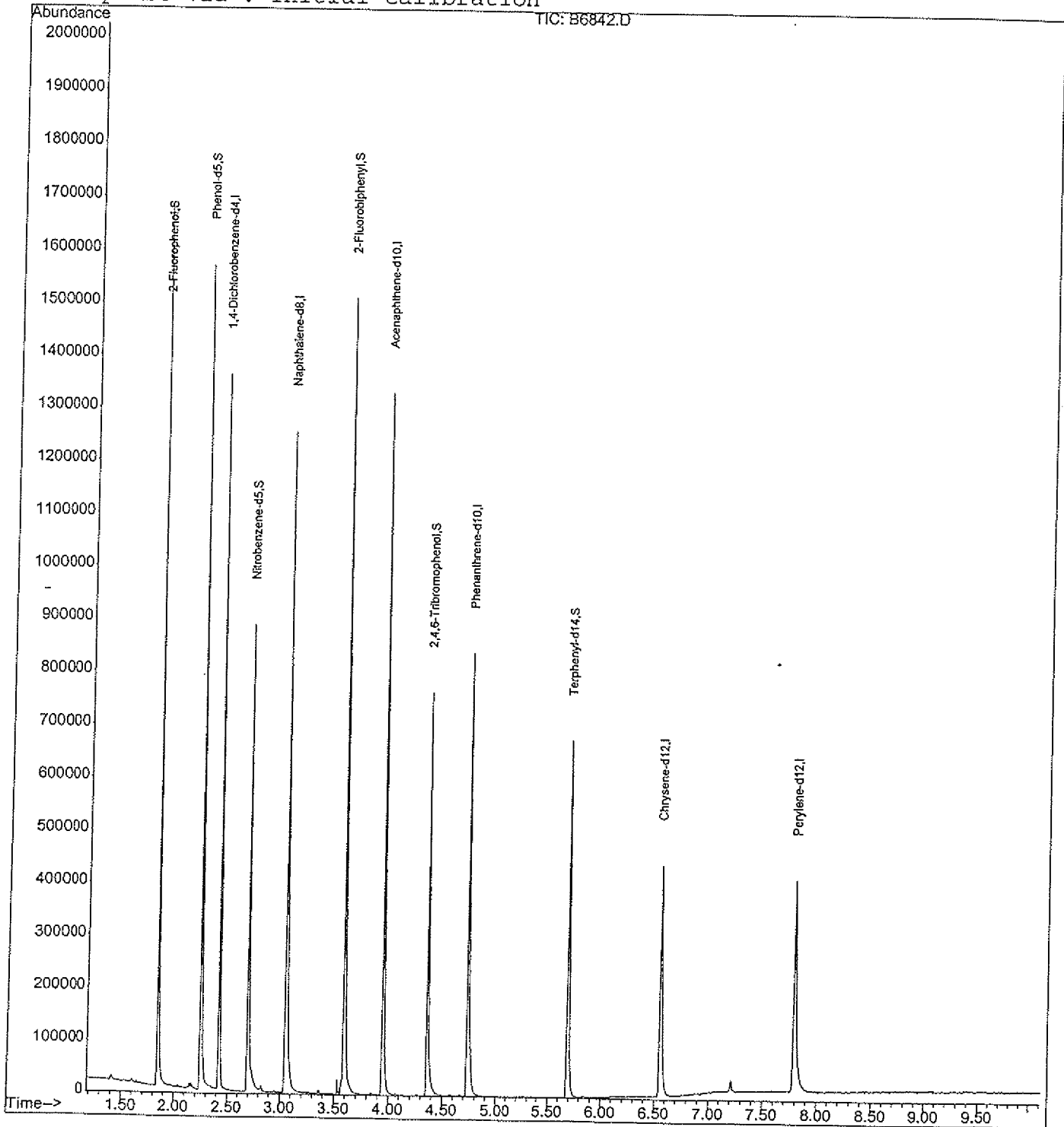
Quantitation Report (QT Reviewed)

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6842.D
 Acq On : 15 Oct 2001 11:07
 Sample : .,METHOD-BLK,S,30.0g,0.0,10/15/01
 Misc : N/A,N/A,N/A,1
 MS Integration Params: rteint.p
 Quant Time: Oct 16 10:00 2001

Vial: 9
 Operator: JC
 Inst : MSD_B
 Multiplr: 1.00

Quant Results File: BS2301.RES

Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
 Title : BNA CALIBRATION METHOD
 Last Update : Mon Oct 08 11:06:23 2001
 Response via : Initial Calibration



Library Search Compound Report

Data File : C:\MSDCHEM\1\DATA\10-15-01\B6842.D Vial: 9
Acq On : 15 Oct 2001 11:07 Operator: JC
Sample : .,METHOD-BLK,S,30.0g,0.0,10/15/01 Inst : MSD_B
Misc : N/A,N/A,N/A,1 Multiplr: 1.00
MS Integration Params: LSCINT.P
Quant Method : C:\MSDCHEM\1\METHODS\BS2301.M (RTE Integrator)
Title : BNA CALIBRATION METHOD
Library : C:\DATABASE\NIST98.L

No Library Search Compounds Detected

B6842.D BS2301.M Tue Oct 16 09:24:22 2001 MSD_B

PCB METHOD BLANK SUMMARY

Lab File ID: V6528.D

Instrument ID: GC-V

Date Extracted: 10/15/01.4

Matrix: SOIL

Date Analyzed: 10/16/2001

Time Analyzed: 13:06

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
ABSEGAMI	7035-001	10/16/2001	13:38
5-F1	7041-001	10/16/2001	14:10
UG-1_10-10.5	7023-001	10/16/2001	14:42
UG-2_9-9.5	7023-002	10/16/2001	15:14
FO-1_6-6.5	7023-004	10/16/2001	15:46
DF-1_10-10.5	7023-005	10/16/2001	16:18
DF-2_6-6.5	7023-006	10/16/2001	16:50
SANDBLAST_GR	7055-001	10/16/2001	17:23
PMK101001	7070-001	10/16/2001	17:55
DF-3_10-10.5	7023-007	10/16/2001	18:27
DF-4_8.5-9	7023-008	10/16/2001	18:59
DW-1_5-5.5	7023-010	10/16/2001	19:31
DW-2_8-8.5	7023-011	10/16/2001	20:03
PCB	7041-001-MS1	10/16/2001	20:35
PCB	7041-001-MSD1	10/16/2001	21:07
PCB	PBS1015-MS1	10/16/2001	21:39

INTEGRATED ANALYTICAL LABORATORIES

PCB's

Client/Project: NA

Lab ID: 1015-BLK1

Client ID: (7023..7055)

Date Received: NA

Date Extracted: 10/15/2001

Date Analyzed: 10/16/2001

Data file: V6528.D

GC Column: DB-5/DB1701P

Sample wt/vol: 5.93g

Matrix-Units: Soil-µg/Kg (ppb)

Dilution Factor: 1

% Moisture: 0

Compound	Concentration	Q	MDL
Aroclor-1016	ND		135
Aroclor-1221	ND		135
Aroclor-1232	ND		135
Aroclor-1242	ND		135
Aroclor-1248	ND		135
Aroclor-1254	ND		135
Aroclor-1260	ND		135

PCB INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/15/2001

Instrument ID: Varian 3400

GC Column (1st): DB-5

Data File: V6519.D V6520.D V6521.D V6522.D V6523.D

Compound	RT OF STANDARDS					MEAN RT	RT WI NDOW	
	200	500	1000	2000	4000		FROM	TO
Aroclor-1016	5.75	5.76	5.75	5.75	5.75	5.75	5.68	5.82
Aroclor-1016 {2}	7.28	7.29	7.28	7.28	7.28	7.28	7.21	7.35
Aroclor-1016 {3}	8.30	8.31	8.30	8.30	8.30	8.30	8.23	8.37
Aroclor-1016 {4}	9.15	9.16	9.15	9.15	9.15	9.15	9.08	9.22
Aroclor-1016 {5}	9.63	9.64	9.63	9.62	9.63	9.63	9.56	9.70
Aroclor-1221			3.78				3.71	3.85
Aroclor-1221 {2}			5.39				5.32	5.46
Aroclor-1221 {3}			5.62				5.55	5.69
Aroclor-1221 {4}			5.75				5.68	5.82
Aroclor-1221 {5}			6.85				6.78	6.92
Aroclor-1232			5.75				5.68	5.82
Aroclor-1232 {2}			6.84				6.77	6.91
Aroclor-1232 {3}			8.30				8.23	8.37
Aroclor-1232 {4}			9.15				9.08	9.22
Aroclor-1232 {5}			9.99				9.92	10.06
Aroclor-1242			7.28				7.21	7.35
Aroclor-1242 {2}			9.15				9.08	9.22
Aroclor-1242 {3}			9.73				9.66	9.80
Aroclor-1242 {4}			10.94				10.87	11.01
Aroclor-1242 {5}			11.46				11.39	11.53
Aroclor-1248			8.03				7.96	8.10
Aroclor-1248 {2}			9.15				9.08	9.22
Aroclor-1248 {3}			9.73				9.66	9.80
Aroclor-1248 {4}			10.94				10.87	11.01
Aroclor-1248 {5}			11.46				11.39	11.53
Aroclor-1254			11.69				11.62	11.76
Aroclor-1254 {2}			12.52				12.45	12.59
Aroclor-1254 {3}			13.67				13.60	13.74
Aroclor-1254 {4}			14.50				14.43	14.57
Aroclor-1254 {5}			15.28				15.21	15.35
Aroclor-1260	15.27	15.29	15.28	15.27	15.27	15.28	15.21	15.35
Aroclor-1260 {2}	15.83	15.85	15.84	15.83	15.84	15.84	15.77	15.91
Aroclor-1260 {3}	17.48	17.49	17.48	17.48	17.48	17.48	17.41	17.55
Aroclor-1260 {4}	18.40	18.42	18.41	18.40	18.40	18.41	18.34	18.48
Aroclor-1260 {5}	20.45	20.46	20.45	20.45	20.45	20.45	20.38	20.52

PCB INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/15/2001

Instrument ID: Varian 3400

GC Column (1st): DB-5

Data File: V6519.D V6520.D V6521.D V6522.D V6523.D

Compound	CALIBRATION FACTORS					MEAN	%RSD
	200	500	1000	2000	4000		
Aroclor-1016	6010	5723	5347	5045	5013	5428	7.98
Aroclor-1016 {2}	8005	7573	7153	6738	6680	7230	7.79
Aroclor-1016 {3}	10608	9990	9661	9074	9076	9682	6.72
Aroclor-1016 {4}	4780	4371	4178	4004	3998	4266	7.62
Aroclor-1016 {5}	6590	6190	5904	5562	5511	5952	7.58
Aroclor-1221			1335				
Aroclor-1221 {2}			2380				
Aroclor-1221 {3}			2225				
Aroclor-1221 {4}			6180				
Aroclor-1221 {5}			752				
Aroclor-1232			3900				
Aroclor-1232 {2}			1361				
Aroclor-1232 {3}			2845				
Aroclor-1232 {4}			1281				
Aroclor-1232 {5}			2826				
Aroclor-1242			4274				
Aroclor-1242 {2}			2426				
Aroclor-1242 {3}			2938				
Aroclor-1242 {4}			5617				
Aroclor-1242 {5}			5779				
Aroclor-1248			10876				
Aroclor-1248 {2}			3633				
Aroclor-1248 {3}			3191				
Aroclor-1248 {4}			9738				
Aroclor-1248 {5}			9817				
Aroclor-1254			9207				
Aroclor-1254 {2}			7727				
Aroclor-1254 {3}			15225				
Aroclor-1254 {4}			10213				
Aroclor-1254 {5}			16486				
Aroclor-1260	29270	26701	25208	23760	24046	25797	8.76
Aroclor-1260 {2}	26811	25693	24396	23191	23499	24718	6.15
Aroclor-1260 {3}	56196	53607	51256	49892	54487	53088	4.75
Aroclor-1260 {4}	24861	24398	23301	22127	22185	23374	5.34
Aroclor-1260 {5}	13926	13585	13271	12670	12582	13207	4.39
Average %RSD							6.71

PCB INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/15/2001

Instrument ID: Varian 3400

GC Column (2nd): DB-1701P

Data File: V6519.C V6520.C V6521.C V6522.C V6523.C

Compound	RT OF STANDARDS					MEAN RT	RT WI NDOW	
	200	500	1000	2000	4000		FROM	TO
Aroclor-1016	6.24	6.25	6.24	6.24	6.24	6.24	6.17	6.31
Aroclor-1016 {2}	7.30	7.31	7.30	7.30	7.30	7.30	7.23	7.37
Aroclor-1016 {3}	8.65	8.66	8.65	8.65	8.65	8.65	8.58	8.72
Aroclor-1016 {4}	9.03	9.04	9.03	9.03	9.03	9.03	8.96	9.10
Aroclor-1016 {5}	9.35	9.36	9.35	9.35	9.35	9.35	9.28	9.42
Aroclor-1221			4.00				3.93	4.07
Aroclor-1221 {2}			5.68				5.61	5.75
Aroclor-1221 {3}			6.09				6.02	6.16
Aroclor-1221 {4}			6.25				6.18	6.32
Aroclor-1221 {5}			7.30				7.23	7.37
Aroclor-1232			6.24				6.17	6.31
Aroclor-1232 {2}			7.30				7.23	7.37
Aroclor-1232 {3}			8.65				8.58	8.72
Aroclor-1232 {4}			9.35				9.28	9.42
Aroclor-1232 {5}			10.45				10.38	10.52
Aroclor-1242			7.98				7.91	8.05
Aroclor-1242 {2}			9.35				9.28	9.42
Aroclor-1242 {3}			10.74				10.67	10.81
Aroclor-1242 {4}			11.78				11.71	11.85
Aroclor-1242 {5}			12.51				12.44	12.58
Aroclor-1248			8.66				8.59	8.73
Aroclor-1248 {2}			9.74				9.67	9.81
Aroclor-1248 {3}			10.46				10.39	10.53
Aroclor-1248 {4}			10.74				10.67	10.81
Aroclor-1248 {5}			11.40				11.33	11.47
Aroclor-1254			12.40				12.33	12.47
Aroclor-1254 {2}			13.50				13.43	13.57
Aroclor-1254 {3}			14.58				14.51	14.65
Aroclor-1254 {4}			15.02				14.95	15.09
Aroclor-1254 {5}			16.15				16.08	16.22
Aroclor-1260	15.01	15.03	15.02	15.02	15.02	15.02	14.95	15.09
Aroclor-1260 {2}	16.27	16.28	16.28	16.27	16.27	16.27	16.20	16.34
Aroclor-1260 {3}	17.28	17.29	17.28	17.28	17.28	17.28	17.21	17.35
Aroclor-1260 {4}	18.24	18.26	18.25	18.24	18.24	18.25	18.18	18.32
Aroclor-1260 {5}	20.52	20.53	20.53	20.52	20.52	20.52	20.45	20.59

PCB INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/15/2001

Instrument ID: Varian 3400

GC Column (2nd): DB-1701P

Data File: V6519.C V6520.C V6521.C V6522.C V6523.C

Compound	CALIBRATION FACTORS					MEAN	%RSD
	200	500	1000	2000	4000		
Aroclor-1016	7725	7195	6865	6524	6461	6954	7.50
Aroclor-1016 {2}	13823	13059	12161	11199	11193	12287	9.41
Aroclor-1016 {3}	33040	30365	28477	27146	28608	29527	7.70
Aroclor-1016 {4}	9511	10683	10129	9574	9543	9888	5.18
Aroclor-1016 {5}	11668	11102	10433	9889	9851	10589	7.45
Aroclor-1221			1560				
Aroclor-1221 {2}			2849				
Aroclor-1221 {3}			1905				
Aroclor-1221 {4}			6990				
Aroclor-1221 {5}			548				
Aroclor-1232			4543				
Aroclor-1232 {2}			3389				
Aroclor-1232 {3}			7986				
Aroclor-1232 {4}			2966				
Aroclor-1232 {5}			3845				
Aroclor-1242			3111				
Aroclor-1242 {2}			5904				
Aroclor-1242 {3}			6625				
Aroclor-1242 {4}			6278				
Aroclor-1242 {5}			4513				
Aroclor-1248			13189				
Aroclor-1248 {2}			8752				
Aroclor-1248 {3}			10273				
Aroclor-1248 {4}			10237				
Aroclor-1248 {5}			6192				
Aroclor-1254			4327				
Aroclor-1254 {2}			3489				
Aroclor-1254 {3}			12313				
Aroclor-1254 {4}			7254				
Aroclor-1254 {5}			15662				
Aroclor-1260	22810	21041	19598	18256	18216	19984	9.81
Aroclor-1260 {2}	19016	17933	16743	15586	15348	16925	9.20
Aroclor-1260 {3}	21335	19745	18498	17214	17052	18769	9.60
Aroclor-1260 {4}	54073	51411	48541	46628	49255	49981	5.71
Aroclor-1260 {5}	6567	6401	6295	6026	5963	6250	4.06
Average %RSD							7.56

PCB CALIBRATION VERIFICATION SUMMARY

Date/Time Analyzed: 10/16/2001

Instrument ID: Varian 3400

Data File: V6527.D

GC Column (1st): DB-5

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
Aroclor-1016	5.76	5.69	5.83	5428	5626	3.65
Aroclor-1016 {2}	7.29	7.22	7.36	7230	7499	3.72
Aroclor-1016 {3}	8.31	8.24	8.38	9682	10200	5.36
Aroclor-1016 {4}	9.16	9.09	9.23	4266	4403	3.21
Aroclor-1016 {5}	9.64	9.57	9.71	5952	6160	3.51
Aroclor-1260	15.29	15.22	15.36	25797	26314	2.00
Aroclor-1260 {2}	15.85	15.78	15.92	24718	25495	3.14
Aroclor-1260 {3}	17.49	17.42	17.56	53088	53290	0.38
Aroclor-1260 {4}	18.42	18.35	18.49	23374	24317	4.03
Aroclor-1260 {5}	20.46	20.39	20.53	13207	13808	4.55
Average %D						3.36

Data File: V6527.C

GC Column (2nd): DB-1701P

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
Aroclor-1016	6.25	6.18	6.32	6954	6449	7.25
Aroclor-1016 {2}	7.31	7.24	7.38	12287	11457	6.76
Aroclor-1016 {3}	8.66	8.59	8.73	29527	26812	9.20
Aroclor-1016 {4}	9.04	8.97	9.11	9888	9553	3.39
Aroclor-1016 {5}	9.36	9.29	9.43	10589	9867	6.82
Aroclor-1260	15.03	14.96	15.10	19984	18613	6.86
Aroclor-1260 {2}	16.28	16.21	16.35	16925	15862	6.28
Aroclor-1260 {3}	17.29	17.22	17.36	18769	17538	6.56
Aroclor-1260 {4}	18.26	18.19	18.33	49981	45545	8.88
Aroclor-1260 {5}	20.53	20.46	20.60	6250	5929	5.14
Average %D						6.71

PCB CALIBRATION VERIFICATION SUMMARY

Date/Time Analyzed: 10/16/2001

Instrument ID: Varian 3400

Data File: V6545.D

GC Column (1st): DB-5

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
Aroclor-1016	5.76	5.69	5.83	5428	5431	0.06
Aroclor-1016 {2}	7.29	7.22	7.36	7230	7323	1.29
Aroclor-1016 {3}	8.31	8.24	8.38	9682	9903	2.29
Aroclor-1016 {4}	9.16	9.09	9.23	4266	4285	0.44
Aroclor-1016 {5}	9.63	9.56	9.70	5952	5997	0.77
Aroclor-1260	15.29	15.22	15.36	25797	25764	0.13
Aroclor-1260 {2}	15.85	15.78	15.92	24718	24870	0.62
Aroclor-1260 {3}	17.49	17.42	17.56	53088	52324	1.44
Aroclor-1260 {4}	18.42	18.35	18.49	23374	24031	2.81
Aroclor-1260 {5}	20.46	20.39	20.53	13207	13717	3.86
Average %D						1.37

Data File: V6545.C

GC Column (2nd): DB-1701P

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
Aroclor-1016	6.25	6.18	6.32	6954	7409	6.54
Aroclor-1016 {2}	7.31	7.24	7.38	12287	13103	6.64
Aroclor-1016 {3}	8.66	8.59	8.73	29527	30690	3.94
Aroclor-1016 {4}	9.04	8.97	9.11	9888	10926	10.49
Aroclor-1016 {5}	9.36	9.29	9.43	10589	11279	6.52
Aroclor-1260	15.03	14.96	15.10	19984	21174	5.95
Aroclor-1260 {2}	16.28	16.21	16.35	16925	18080	6.83
Aroclor-1260 {3}	17.29	17.22	17.36	18769	20008	6.60
Aroclor-1260 {4}	18.26	18.19	18.33	49981	52658	5.35
Aroclor-1260 {5}	20.53	20.46	20.60	6250	6874	9.98
Average %D						6.88

PCB SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/16/2001

Client ID	Lab Sample ID	Matrix	TCMX 1		DCB 1		TCMX 2		DCB 2	
			% rec	#	% rec	#	% rec	#	% rec	#
(7023..7055)	1015-BLK1	SOIL	103		98		97		92	
ABSEGAMI	7035-001	SOIL	100		94		96		89	
5-F1	7041-001	SOIL	102		96		100		93	
UG-1_10-10.5	7023-001	SOIL	104		98		104		96	
UG-2_9-9.5	7023-002	SOIL	100		100		101		98	
FO-1_6-6.5	7023-004	SOIL	99		100		120		99	
DF-1_10-10.5	7023-005	SOIL	100		97		102		97	
DF-2_6-6.5	7023-006	SOIL	100		102		143		103	
SANDBLAST_GR	7055-001	SOIL	104		99		108		100	
PMK101001	7070-001	SOIL	101		102		106		104	
DF-3_10-10.5	7023-007	SOIL	87		92		92		94	
DF-4_8.5-9	7023-008	SOIL	90		95		99		98	
DW-1_5-5.5	7023-010	SOIL	100		95		111		97	
DW-2_8-8.5	7023-011	SOIL	82		93		88		96	
PCB	7041-001-MS1	SOIL	101		103		109		106	
PCB	7041-001-MSD1	SOIL	101		99		109		103	
PCB	PBS1015-MS1	SOIL	100		102		109		107	

Surrogate QC Limits

TCMX = Tetrachloro-m-xylene

DCB = Decachlorobiphenyl

<u>Soil</u>	<u>Aqueous</u>
39-147	39-151
33-179	30-150

Column to be used to flag recovery values

* Values outside of QC limits

D Surrogate diluted out

M Matrix interference

SOIL PCB MATRIX SPIKE/SPIKE DUPLICATE RECOVERY

Matrix spike Lab sample ID: 7041-001-MSD1

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
Aroclor-1016	5.0	0.0	3.8	76	58 - 140
Aroclor-1260	5.0	0.0	3.8	76	56 - 161

Compound	SAMPLE CONC. (ug/Kg)	MSD CONC. (ug/Kg)	MSD % # REC	% RPD #	QC LIMITS	
					RPD	REC.
Aroclor-1016	0.0	3.9	78	3	14	58 - 140
Aroclor-1260	0.0	3.8	76	0	18	56 - 161

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

RPD: 0 out of 2 outside limits

Spike Recovery: 0 out of 4 outside limits

SOIL PCB BLANK SPIKE RECOVERY

Matrix spike Lab sample ID: PBS1015-MS1

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
Aroclor-1016	5.0	0.0	3.9	78	58 - 140
Aroclor-1260	5.0	0.0	4.0	80	56 - 161

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

Spike Recovery: 0 out of 2 outside limits

PCB RETENTION TIME SHIFT SUMMARY

Instrument ID: GC-V

Column: DB-5/DB-1701P

Surrogate RT from initial calibration :

TCMX 1 4.92 DCB 1 22.58 TCMX 2 4.76 DCB 2 22.63

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed	TCMX 1 RT #	DCB 1 RT #	TCMX 2 RT #	DCB 2 RT #
(7023..7055)	1015-BLK1	10/16/2001	13:06	4.92	22.58	4.76	22.63
ABSEGAMI	7035-001	10/16/2001	13:38	4.92	22.58	4.76	22.64
5-F1	7041-001	10/16/2001	14:10	4.92	22.57	4.75	22.63
UG-1_10-10.5	7023-001	10/16/2001	14:42	4.92	22.57	4.75	22.62
UG-2_9-9.5	7023-002	10/16/2001	15:14	4.92	22.57	4.75	22.63
FO-1_6-6.5	7023-004	10/16/2001	15:46	4.92	22.58	4.76	22.63
DF-1_10-10.5	7023-005	10/16/2001	16:18	4.92	22.58	4.76	22.64
DF-2_6-6.5	7023-006	10/16/2001	16:50	4.92	22.57	4.75	22.63
SANDBLAST_GR	7055-001	10/16/2001	17:23	4.92	22.57	4.76	22.63
PMK101001	7070-001	10/16/2001	17:55	4.92	22.57	4.75	22.63
DF-3_10-10.5	7023-007	10/16/2001	18:27	4.92	22.58	4.76	22.64
DF-4_8.5-9	7023-008	10/16/2001	18:59	4.92	22.57	4.76	22.63
DW-1_5-5.5	7023-010	10/16/2001	19:31	4.92	22.58	4.76	22.64
DW-2_8-8.5	7023-011	10/16/2001	20:03	4.93	22.59	4.76	22.64
PCB	7041-001-MS1	10/16/2001	20:35	4.92	22.58	4.76	22.64
PCB	7041-001-MSD1	10/16/2001	21:07	4.92	22.57	4.75	22.63
PCB	PBS1015-MS1	10/16/2001	21:39	4.93	22.58	4.76	22.64

Surrogate QC Limits

TCMX = Tetrachloro-m-xylene

(± 0.05 Minutes)

DCB = Decachlorobiphenyl

(± 0.05 Minutes)

Column to be used to flag recovery values

* Values outside of QC limits

D Surrogate diluted out

M Matrix interference

Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6530.D\ADC1A.CH Vial: 3
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6530.D\ADC1B.CH
 Acq On : 16 Oct 2001 14:10 Operator: MJ
 Sample : 5-F1,7041-001,S,5.72g,9.47,10/15/01,4 Inst : V_3400
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 07:22:15 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Initial Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

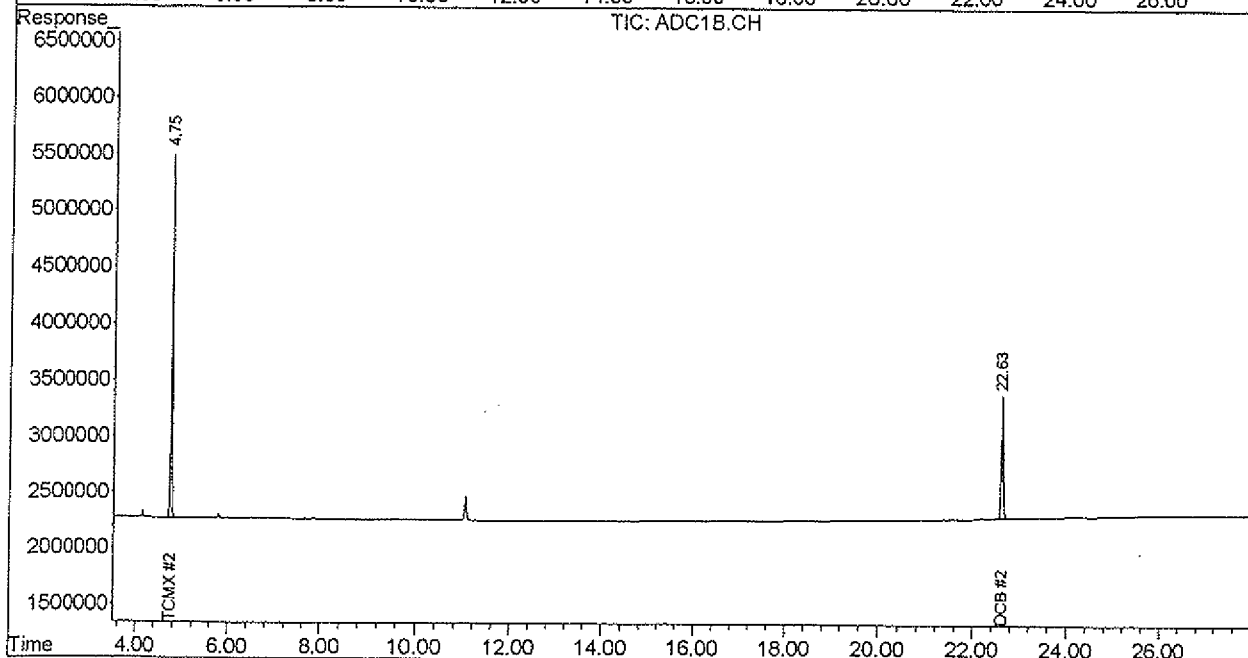
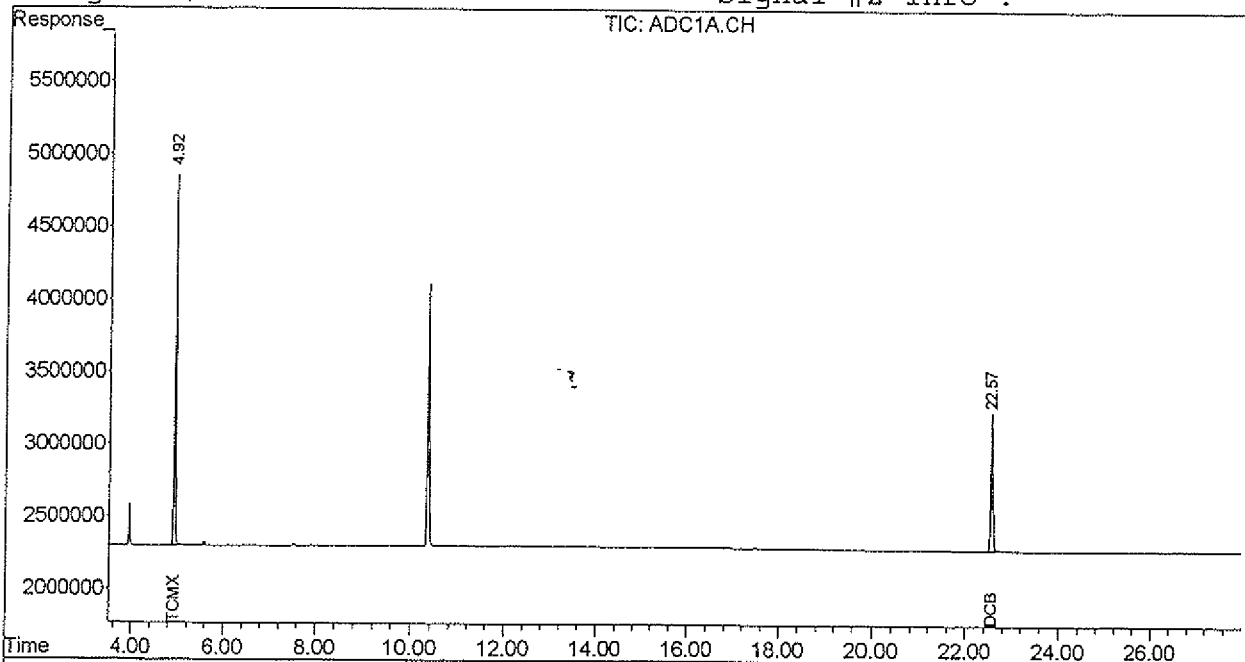
System Monitoring Compounds						
1) S TCMX	4.92	4.75	47042755	57818402	102.166	100.011
Spiked Amount	100.000			Recovery	=	102.17%
2) S DCB	22.57	22.63	30320478	35265059	95.977	92.635
Spiked Amount	100.000			Recovery	=	95.98%
Target Compounds						
Sum Aroclor-1016			0	0	N.D.	N.D.
Average Aroclor-1016					0.000	0.000
Sum Aroclor-1221			0	0	N.D.	N.D.
Average Aroclor-1221					0.000	0.000
Sum Aroclor-1232			0	0	N.D.	N.D.
Average Aroclor-1232					0.000	0.000
Sum Aroclor-1242			0	0	N.D.	N.D.
Average Aroclor-1242					0.000	0.000
Sum Aroclor-1248			0	0	N.D.	N.D.
Average Aroclor-1248					0.000	0.000
Sum Aroclor-1254			0	0	N.D.	N.D.
Average Aroclor-1254					0.000	0.000
Sum Aroclor-1260			0	0	N.D.	N.D.
Average Aroclor-1260					0.000	0.000

Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6530.D\ADC1A.CH Vial: 3
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6530.D\ADC1B.CH
 Acq On : 16 Oct 2001 14:10 Operator: MJ
 Sample : 5-F1,7041-001,S,5.72g,9.47,10/15/01,4 Inst : V_3400
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 7:28 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :



Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6528.D\ADC1A.CH Vial: 1
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6528.D\ADC1B.CH
 Acq On : 16 Oct 2001 13:06 Operator: MJ
 Sample : (7023..7055),1015-BLK1,S,5.93g,0,10/15/0 Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 07:22:05 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Initial Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

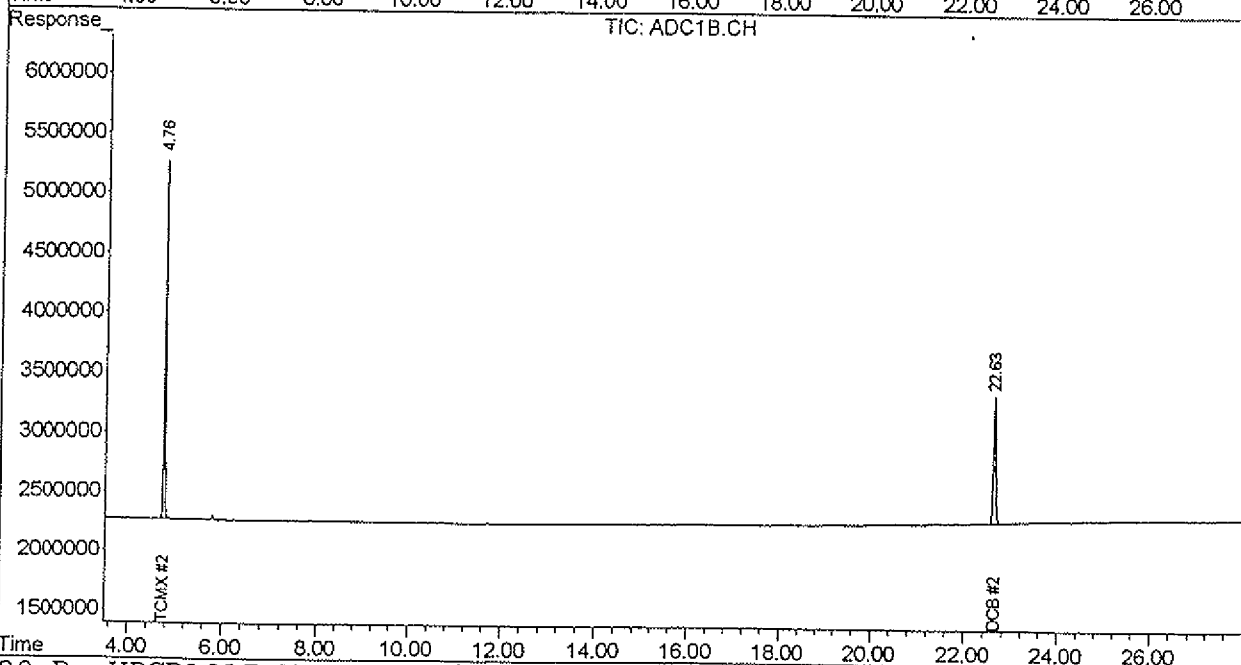
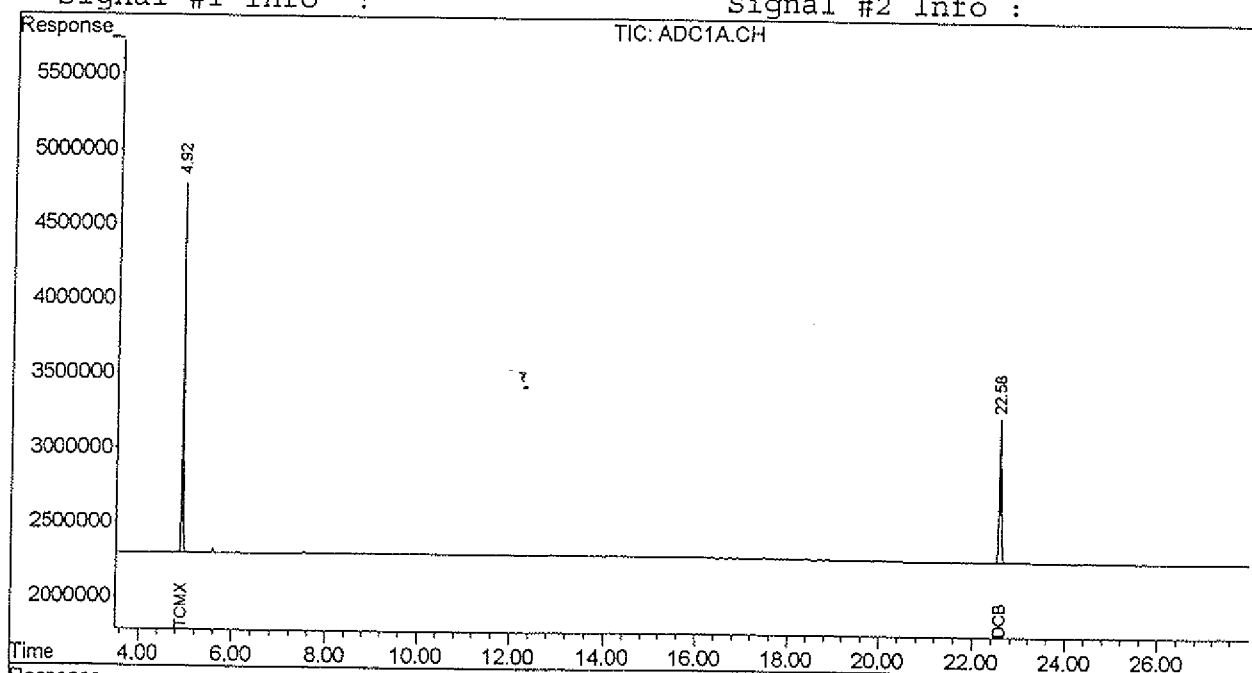
Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2
System Monitoring Compounds						
1) S TCMX	4.92	4.76	47293817	56164306	102.711	97.150
Spiked Amount	100.000		Recovery	=	102.71%	97.15%
2) S DCB	22.58	22.63	30868105	35020161	97.710	91.991
Spiked Amount	100.000		Recovery	=	97.71%	91.99%
Target Compounds						
Sum Aroclor-1016			0	0	N.D.	N.D.
Average Aroclor-1016					0.000	0.000
Sum Aroclor-1221			0	0	N.D.	N.D.
Average Aroclor-1221					0.000	0.000
Sum Aroclor-1232			0	0	N.D.	N.D.
Average Aroclor-1232					0.000	0.000
Sum Aroclor-1242			0	0	N.D.	N.D.
Average Aroclor-1242					0.000	0.000
Sum Aroclor-1248			0	0	N.D.	N.D.
Average Aroclor-1248					0.000	0.000
Sum Aroclor-1254			0	0	N.D.	N.D.
Average Aroclor-1254					0.000	0.000
Sum Aroclor-1260			0	0	N.D.	N.D.
Average Aroclor-1260					0.000	0.000

Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6528.D\ADC1A.CH Vial: 1
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6528.D\ADC1B.CH
 Acq On : 16 Oct 2001 13:06 Operator: MJ
 Sample : (7023..7055),1015-BLK1,S,5.93g,0,10/15/0 Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 7:27 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :



Quantitation Report (Not Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6527.D\ADC1A.CH Vial: 14
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6527.D\ADC1B.CH
 Acq On : 16 Oct 2001 12:02 Operator: MJ
 Sample : 8082_C_IAS1574,1_PPM Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 07:21:58 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Initial Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	4.92	4.76	45088682	50229407	97.922	86.884
Spiked Amount 100.000			Recovery	=	97.92%	86.88%
2) S DCB	22.58	22.63	31760958	34329163	100.536	90.176
Spiked Amount 100.000			Recovery	=	100.54%	90.18%
Target Compounds						
3) L2 Aroclor-1016	5.76	6.25	5625942	6449490	1036.527	927.461
4) L2 Aroclor-1016 {2}	7.29	7.31	7498730	11456522	1037.221	932.422
5) L2 Aroclor-1016 {3}	8.31	8.66	10200325	26811558	1053.562	908.023
6) L2 Aroclor-1016 {4}	9.16	9.04	4403027	9552570	1032.078	966.064
7) L2 Aroclor-1016 {5}	9.64	9.36	6160315	9866811	1035.072	931.823
Sum Aroclor-1016			33888338	64136951	5194.460	4665.794
Average Aroclor-1016					1038.892	933.159
Sum Aroclor-1221			0	0	N.D.	N.D.
Average Aroclor-1221					0.000	0.000
Sum Aroclor-1232			0	0	N.D.	N.D.
Average Aroclor-1232					0.000	0.000
Sum Aroclor-1242			0	0	N.D.	N.D.
Average Aroclor-1242					0.000	0.000
Sum Aroclor-1248			0	0	N.D.	N.D.
Average Aroclor-1248					0.000	0.000
Sum Aroclor-1254			0	0	N.D.	N.D.
Average Aroclor-1254					0.000	0.000
3) L8 Aroclor-1260	15.29	15.03	26314389	18613389	1020.046	931.403
4) L8 Aroclor-1260 {2}	15.85	16.28	25494609	15861652	1031.425	937.167
5) L8 Aroclor-1260 {3}	17.49	17.29	53289695	17537726	1003.805	934.412
6) L8 Aroclor-1260 {4}	18.42	18.26	24316750	45544712	1040.312	911.234
7) L8 Aroclor-1260 {5}	20.46	20.53	13808162	5929257	1045.522	948.640
Sum Aroclor-1260			143.2E6	103.5E6	5141.111	4662.856
Average Aroclor-1260					1028.222	932.571

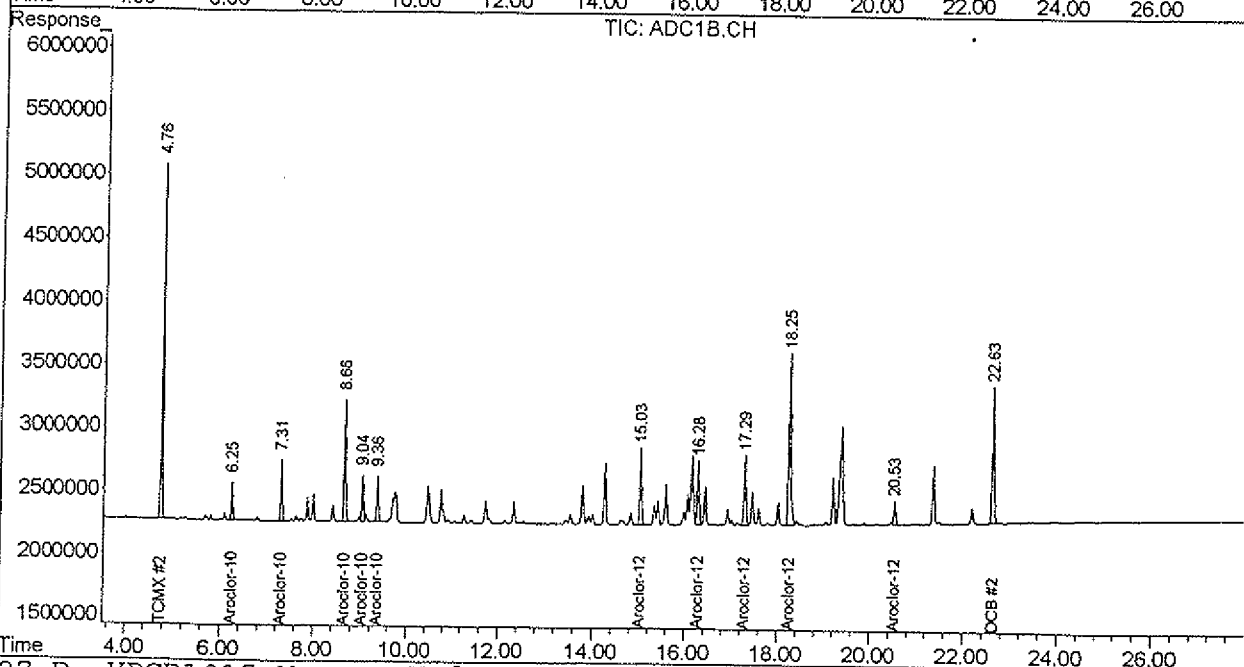
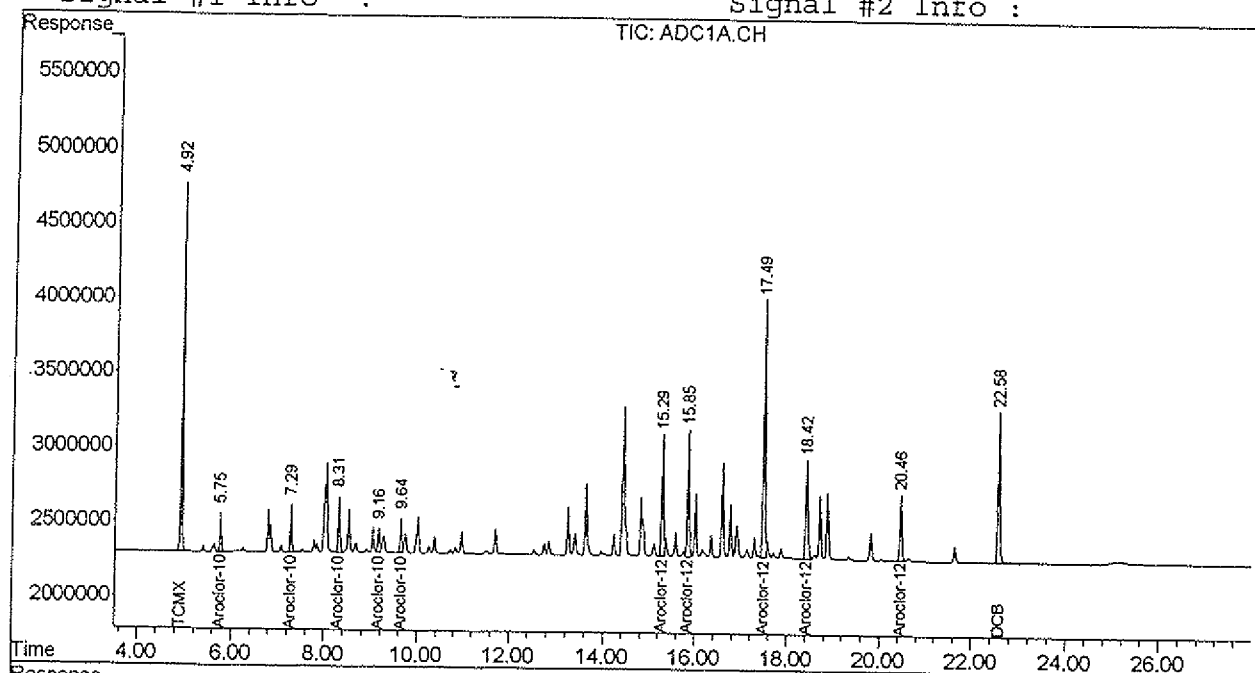
(f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.
 V6527.D VPCB1015.M Wed Oct 17 07:50:22 2001 V3400 Page 1

Quantitation Report (Not Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6527.D\ADC1A.CH Vial: 14
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6527.D\ADC1B.CH
 Acq On : 16 Oct 2001 12:02 Operator: MJ
 Sample : 8082_C_IAS1574,1_PPM Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 7:27 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6545.D\ADC1A.CH Vial: 14
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6545.D\ADC1B.CH
 Acq On : 16 Oct 2001 22:11 Operator: MJ
 Sample : 8082_C_IAS1574,1_PPM Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 07:23:46 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Initial Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	4.92	4.76	43353813	59320484	94.154	102.609
Spiked Amount 100.000			Recovery	=	94.15%	102.61%
2) S DCB	22.58	22.64	31975698	40101198	101.216	105.338
Spiked Amount 100.000			Recovery	=	101.22%	105.34%
Target Compounds						
3) L2 Aroclor-1016	5.76	6.25	5431076	7408986	1000.625	1065.441
4) L2 Aroclor-1016 {2}	7.29	7.31	7323125	13102559	1012.931	1066.389
5) L2 Aroclor-1016 {3}	8.31	8.66	9903276	30689978	1022.881	1039.372
6) L2 Aroclor-1016 {4}	9.16	9.04	4284872	10925767	1004.383	1104.938
7) L2 Aroclor-1016 {5}	9.63	9.36	5997486	11279229	1007.713	1065.212
Sum Aroclor-1016			32939835	73406519	5048.532	5341.352
Average Aroclor-1016					1009.706	1068.270
Sum Aroclor-1221			0	0	N.D.	N.D.
Average Aroclor-1221					0.000	0.000
Sum Aroclor-1232			0	0	N.D.	N.D.
Average Aroclor-1232					0.000	0.000
Sum Aroclor-1242			0	0	N.D.	N.D.
Average Aroclor-1242					0.000	0.000
Sum Aroclor-1248			0	0	N.D.	N.D.
Average Aroclor-1248					0.000	0.000
Sum Aroclor-1254			0	0	N.D.	N.D.
Average Aroclor-1254					0.000	0.000
33) L8 Aroclor-1260	15.29	15.03	25764332	21173867	998.723	1059.528
34) L8 Aroclor-1260 {2}	15.85	16.28	24870436	18080488	1006.173	1068.264
35) L8 Aroclor-1260 {3}	17.49	17.29	52324078	20007904	985.616	1066.024
36) L8 Aroclor-1260 {4}	18.42	18.26	24030754	52657760	1028.077	1053.548
37) L8 Aroclor-1260 {5}	20.46	20.53	13717396	6873789	1038.650	1099.759
Sum Aroclor-1260			140.7E6	118.8E6	5057.240	5347.122
Average Aroclor-1260					1011.448	1069.424

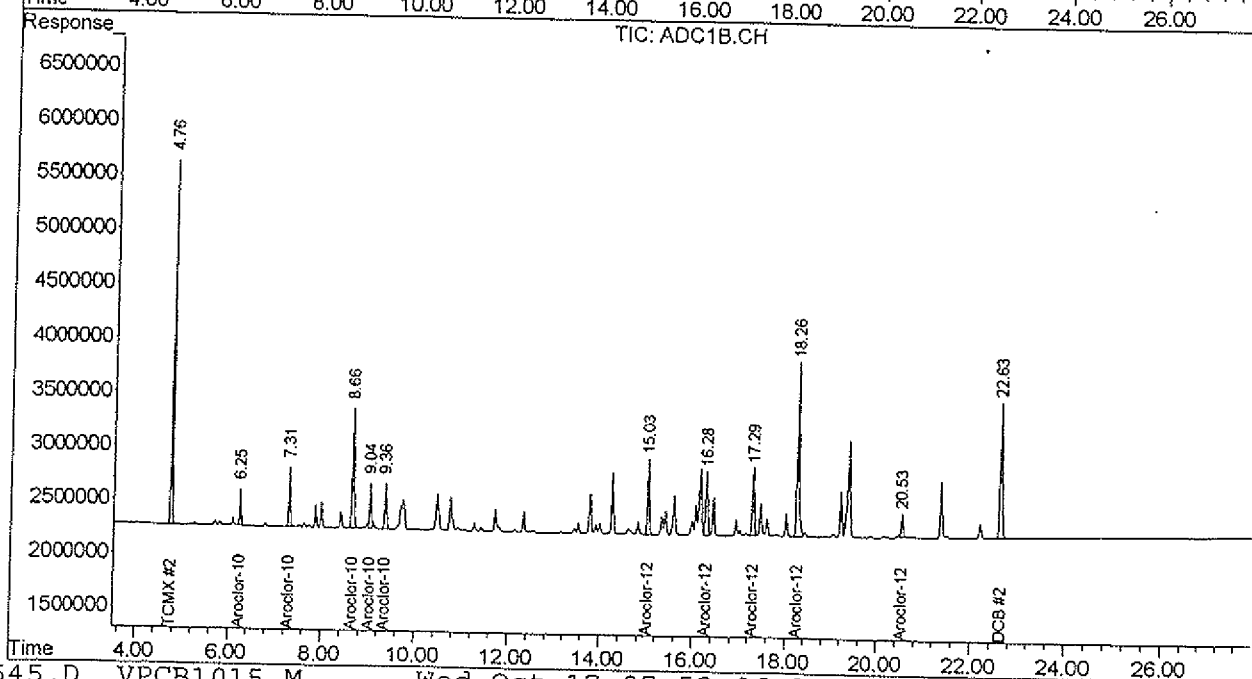
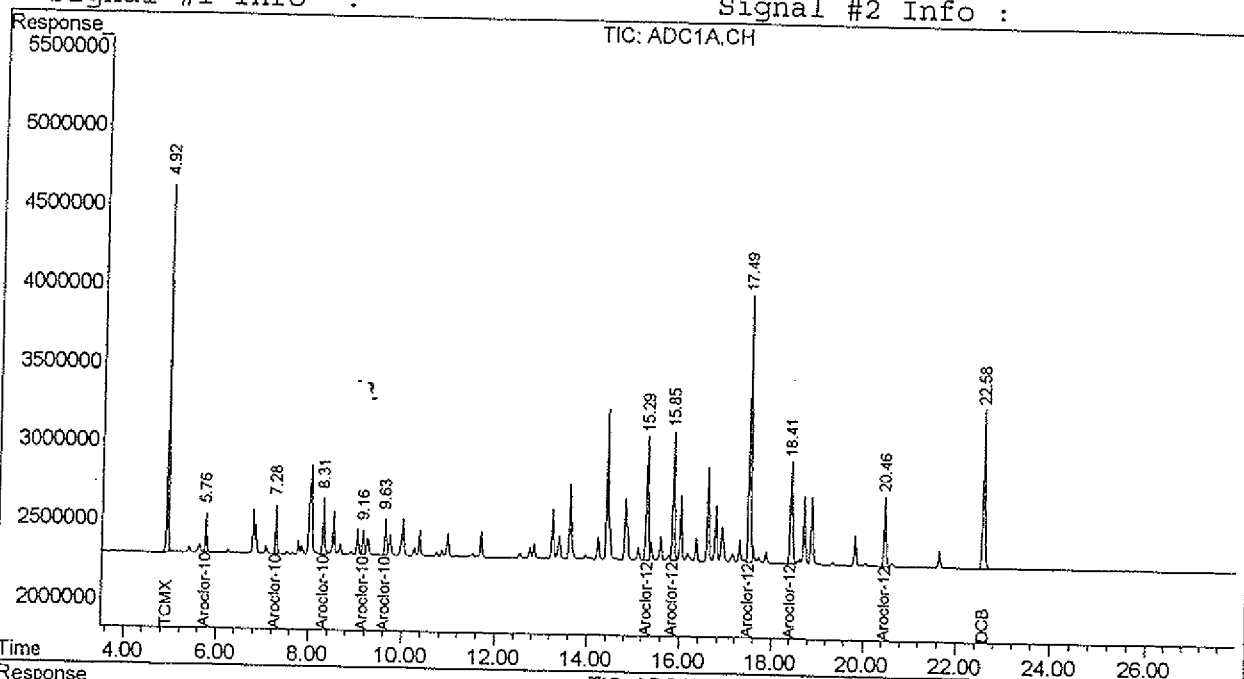
(f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.
 V6545.D VPCB1015.M Wed Oct 17 07:52:02 2001 V3400 Page 1

Quantitation Report (QT Reviewed)

Signal #1 : C:\MSDCHEM\1\DATA\10-16-01\V6545.D\ADC1A.CH Vial: 14
 Signal #2 : C:\MSDCHEM\1\DATA\10-16-01\V6545.D\ADC1B.CH
 Acq On : 16 Oct 2001 22:11 Operator: MJ
 Sample : 8082_C_IAS1574,1_PPM Inst : V_3400
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: events.e IntFile Signal #2: events2.e
 Quant Time: Oct 17 7:38 2001 Quant Results File: VPCB1015.RES

Quant Method : C:\MSDCHEM\1\METHODS\VPCB1015.M (Chemstation Integrator)
 Title :
 Last Update : Tue Oct 16 12:18:24 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : VPCB1015.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



PESTICIDE METHOD BLANK SUMMARY

Lab File ID: W8926.D

Instrument ID: GC-W

Date Extracted: 10/15/2001

Matrix: SOIL

Date Analyzed: 10/16/2001

Time Analyzed: 14:33

THIS METHOD BLANK APPLIES TO THE FOLLOWING SAMPLES, MS & MSD:

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed
ABSEGAMI	7035-001	10/16/2001	14:58
5-F1	7041-001	10/16/2001	15:33
UG-1_10-10.5	7023-001	10/16/2001	15:58
UG-2_9-9.5	7023-002	10/16/2001	16:23
FO-1_6-6.5	7023-004	10/16/2001	16:48
DF-1_10-10.5	7023-005	10/16/2001	17:13
DF-2_6-6.5	7023-006	10/16/2001	17:38
DF-3_10-10.5	7023-007	10/16/2001	18:03
DF-4_8.5-9	7023-008	10/16/2001	18:28
DW-1_5-5.5	7023-010	10/16/2001	18:53
DW-2_8-8.5	7023-011	10/16/2001	19:18
PEST	7041-001-MS1	10/16/2001	19:43
PEST	7041-001-MSD1	10/16/2001	20:08
PEST	PBS1015-MS1	10/16/2001	20:33

INTEGRATED ANALYTICAL LABORATORIES

PESTICIDES

Client/Project: NA

Lab ID: 1015-BK1

Client ID: (7023.7041)

Date Received: NA

Date Extracted: 10/15/2001

Date Analyzed: 10/16/2001

Data file: W8926.D

GC Column: DB-5/DB1701P

Sample wt/vol: 5.9g

Matrix-Units: Soil- μ g/Kg (ppb)

Dilution Factor: 1

% Moisture: 0.0

Compound	Concentration	Q	MDL
alpha-BHC	ND		3.39
beta-BHC	ND		3.39
gamma-BHC	ND		3.39
delta-BHC	ND		3.39
Heptachlor	ND		3.39
Aldrin	ND		3.39
Heptachlor epoxide	ND		3.39
Endosulfan I	ND		3.39
4,4'-DDE	ND		3.39
Dieldrin	ND		3.39
Endrin	ND		3.39
Endosulfan II	ND		3.39
4,4'-DDD	ND		3.39
Endrin aldehyde	ND		3.39
Endosulfan sulfate	ND		3.39
4,4'-DDT	ND		3.39
Chlordane	ND		17
Toxaphene	ND		17

PESTICIDE INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/03/2001

Instrument ID: GC-W

GC Column (1st): DB-5

Data File: W8772.D W8771.D W8770.D W8769.D W8768.D

Compound	RT OF STANDARDS					MEAN RT	RT WINDOW	
	10	50	100	200	300		FROM	TO
alpha-BHC	4.36	4.36	4.36	4.36	4.35	4.36	4.29	4.43
beta-BHC	4.84	4.85	4.85	4.84	4.83	4.84	4.77	4.91
gamma-BHC	4.97	4.97	4.97	4.97	4.96	4.97	4.90	5.04
delta-BHC	5.44	5.44	5.44	5.44	5.43	5.44	5.37	5.51
Heptachlor	6.52	6.52	6.52	6.52	6.51	6.52	6.45	6.59
Aldrin	7.33	7.33	7.33	7.33	7.32	7.33	7.26	7.40
Heptachlor epoxide	8.33	8.33	8.33	8.33	8.32	8.33	8.26	8.40
Endosulfan I	9.23	9.24	9.24	9.24	9.23	9.23	9.16	9.30
4,4'-DDE	10.00	10.00	10.01	10.00	10.00	10.00	9.93	10.07
Dieldrin	9.92	9.92	9.92	9.92	9.92	9.92	9.85	9.99
Endrin	10.47	10.47	10.47	10.47	10.46	10.47	10.40	10.54
Endosulfan II	10.73	10.73	10.73	10.72	10.72	10.72	10.65	10.79
4,4'-DDD	11.11	11.11	11.11	11.11	11.10	11.11	11.04	11.18
Endrin aldehyde	11.25	11.25	11.25	11.25	11.24	11.25	11.18	11.32
Endosulfan sulfate	11.90	11.90	11.90	11.90	11.89	11.90	11.83	11.97
4,4'-DDT	12.13	12.13	12.13	12.13	12.12	12.13	12.06	12.20
Endrin ketone	13.09	13.10	13.10	13.10	13.09	13.09	13.02	13.16
Methoxychlor	13.80	13.80	13.80	13.80	13.80	13.80	13.73	13.87
alpha-Chlordane	9.34	9.34	9.34	9.34	9.33	9.34	9.27	9.41
gamma-Chlordane	8.94	8.94	8.94	8.94	8.93	8.93	8.86	9.00
Chlordane 500 ppb			8.94				8.87	9.01
Chlordane {2}			9.14				9.07	9.21
Chlordane {3}			9.34				9.27	9.41
Chlordane {4}			9.49				9.42	9.56
Chlordane {5}			11.32				11.25	11.39
Toxaphene 500 ppb			11.67				11.60	11.74
Toxaphene {2}			12.18				12.11	12.25
Toxaphene {3}			13.03				12.96	13.10
Toxaphene {4}			13.25				13.18	13.32
Toxaphene {5}			14.27				14.20	14.34

PESTICIDE INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/03/2001

Instrument ID: GC-W

GC Column (1st): DB-5

Data File: W8772.D W8771.D W8770.D W8769.D W8768.D

Compound	CALIBRATION FACTORS					MEAN	%RSD
	10	50	100	200	300		
alpha-BHC	41735923	47428777	48592042	48074152	49053223	46976823	6.37
beta-BHC	18079867	17559304	17330508	17050118	17264821	17456924	2.25
gamma-BHC	38461694	42150505	42520097	42456746	42756006	41669009	4.33
delta-BHC	37345093	42531667	42938864	43182033	44174232	42034378	6.40
Heptachlor	32526297	35958364	36204423	35259571	34724085	34934548	4.20
Aldrin	37072827	40724640	40709504	39533769	39208840	39449916	3.79
Heptachlor epoxide	34494618	36236063	35464363	33460908	32581608	34447512	4.28
Endosulfan I	32808899	34243194	33399666	31565782	30810948	32565698	4.25
4,4'-DDE	32821078	36560871	36744403	35877059	35923476	35585378	4.47
Dieldrin	34167758	36995894	36554297	34892507	34059424	35333976	3.86
Endrin	29077991	31686476	30617693	30073232	29696874	30230453	3.27
Endosulfan II	30981860	32241740	31422974	29974691	29259748	30776202	3.83
4,4'-DDD	29987289	32054022	31080325	30692766	30214058	30805692	2.65
Endrin aldehyde	22442860	23056745	22052336	20947595	20811242	21862156	4.42
Endosulfan sulfate	27673717	29067543	28322094	27109035	26862766	27807031	3.24
4,4'-DDT	14951120	19636203	21620620	22394924	23690007	20458575	16.67
Endrin ketone	35263764	35766336	34223870	31790272	30651531	33539155	6.63
Methoxychlor	8983698	10543821	10828881	10726104	10780163	10372533	7.56
alpha-Chlordane	34618561	36634036	36622976	35939999	36066893	35976493	2.29
gamma-Chlordane	34900440	38496765	38824701	38228482	38310425	37752163	4.27
Chlordane 500 ppb			4171440				
Chlordane {2}			887533				
Chlordane {3}			3922407				
Chlordane {4}			2442430				
Chlordane {5}			1050809				
Toxaphene 500 ppb			485936				
Toxaphene {2}			822837				
Toxaphene {3}			917851				
Toxaphene {4}			763741				
Toxaphene {5}			479631				

PESTICIDE INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/03/2001

Instrument ID: GC-W
GC Column (2nd): DB-1701P

Data File: W8772.C W8771.C W8770.C W8769.C W8768.C

Compound	RT OF STANDARDS					MEAN RT	RT WINDOW	
	10	50	100	200	300		FROM	TO
alpha-BHC	5.18	5.19	5.19	5.19	5.18	5.18	5.11	5.25
beta-BHC	8.16	8.16	8.16	8.16	8.16	8.16	8.09	8.23
gamma-BHC	6.10	6.10	6.10	6.10	6.10	6.10	6.03	6.17
delta-BHC	8.70	8.71	8.70	8.70	8.70	8.70	8.63	8.77
Heptachlor	6.56	6.56	6.56	6.56	6.55	6.56	6.49	6.63
Aldrin	7.20	7.20	7.20	7.20	7.20	7.20	7.13	7.27
Heptachlor epoxide	9.01	9.01	9.01	9.01	9.01	9.01	8.94	9.08
Endosulfan I	9.68	9.68	9.68	9.68	9.68	9.68	9.61	9.75
4,4'-DDE	10.46	10.46	10.46	10.46	10.46	10.46	10.39	10.53
Dieldrin	10.70	10.70	10.70	10.70	10.70	10.70	10.63	10.77
Endrin	11.21	11.21	11.21	11.21	11.21	11.21	11.14	11.28
Endosulfan II	12.64	12.64	12.64	12.64	12.64	12.64	12.57	12.71
4,4'-DDD	12.69	12.69	12.69	12.69	12.69	12.69	12.62	12.76
Endrin aldehyde	13.78	13.78	13.78	13.78	13.78	13.78	13.71	13.85
Endosulfan sulfate	14.68	14.68	14.68	14.68	14.68	14.68	14.61	14.75
4,4'-DDT	13.11	13.11	13.11	13.12	13.11	13.11	13.04	13.18
Endrin ketone	15.68	15.68	15.68	15.68	15.68	15.68	15.61	15.75
Methoxychlor	14.98	14.98	14.98	14.98	14.98	14.98	14.91	15.05
alpha-Chlordane	10.07	10.07	10.07	10.07	10.07	10.07	10.00	10.14
gamma-Chlordane	9.92	9.92	9.92	9.92	9.92	9.92	9.85	9.99
Chlordane 500 ppb			7.88				7.81	7.95
Chlordane {2}			9.53				9.46	9.60
Chlordane {3}			9.92				9.85	9.99
Chlordane {4}			10.07				10.00	10.14
Chlordane {5}			10.13				10.06	10.20
Toxaphene 500 ppb			12.71				12.64	12.78
Toxaphene {2}			13.42				13.35	13.49
Toxaphene {3}			14.41				14.34	14.48
Toxaphene {4}			14.60				14.53	14.67
Toxaphene {5}			15.04				14.97	15.11

PESTICIDE INITIAL CALIBRATION SUMMARY

Date Analyzed: 10/03/2001

Instrument ID: GC-W

GC Column (2nd): DB-1701P

Data File: W8772.C W8771.C W8770.C W8769.C W8768.C

Compound	CALIBRATION FACTORS					MEAN	%RSD
	10	50	100	200	300		
alpha-BHC	12385165	14791668	15426108	15714013	15821036	14827598	9.60
beta-BHC	6336455	6088434	6075295	6000261	5985710	6097231	2.31
gamma-BHC	11681418	13542169	13996884	14224266	14289408	13546829	8.00
delta-BHC	11372621	13285253	13761392	14045936	14143642	13321769	8.55
Heptachlor	10889392	12154587	12529820	12570865	12456521	12120237	5.83
Aldrin	10994720	12428148	12866154	12872917	12788406	12390069	6.47
Heptachlor epoxide	10614434	11389226	11497101	11365746	11221822	11217666	3.13
Endosulfan I	9804666	10610519	10834070	10637802	10555166	10488445	3.78
4,4'-DDE	9746868	11233236	11647128	11802215	11845030	11254895	7.79
Dieldrin	10328290	11504511	11776248	11775902	11714108	11419812	5.43
Endrin	9580852	10782043	10790090	11007670	10998107	10631753	5.62
Endosulfan II	9659839	10058289	10111595	10300761	10234850	10073067	2.48
4,4'-DDD	9193669	10221000	10291838	9978695	9793268	9895694	4.44
Endrin aldehyde	7038357	6556987	7029676	7007187	7109833	6948408	3.20
Endosulfan sulfate	8496880	8955067	9004746	9005302	9029495	8898298	2.54
4,4'-DDT	4937829	6721398	7449230	7921891	8384717	7083013	19.03
Endrin ketone	10387205	10966109	10964387	10736416	10602968	10731417	2.30
Methoxychlor	3271752	3813643	3909466	3957344	4051896	3800820	8.10
alpha-Chlordane	10701120	11490133	11746399	11775383	11769701	11496547	4.00
gamma-Chlordane	10610298	11704350	12058834	12190322	12225209	11757803	5.73
Chlordane 500 ppb			214161				
Chlordane {2}			286639				
Chlordane {3}			1309951				
Chlordane {4}			1142675				
Chlordane {5}			723821				
Toxaphene 500 ppb			298191				
Toxaphene {2}			281068				
Toxaphene {3}			291954				
Toxaphene {4}			143654				
Toxaphene {5}			147283				

PESTICIDE CALIBRATION VERIFICATION SUMMARY

Date Analyzed: 10/16/01

Instrument ID: GC-W

Data File: W8923.D

GC Column (1st): DB-5

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
alpha-BHC	4.35	4.28	4.42	46976823	44523067	5.22
beta-BHC	4.84	4.77	4.91	17456924	15860580	9.14
gamma-BHC	4.97	4.90	5.04	41669009	39600256	4.96
delta-BHC	5.44	5.37	5.51	42034378	40019176	4.79
Heptachlor	6.52	6.45	6.59	34934548	35589155	1.87
Aldrin	7.33	7.26	7.40	39449916	37100385	5.96
Heptachlor epoxide	8.33	8.26	8.40	34447512	32957215	4.33
Endosulfan I	9.23	9.16	9.30	32565698	30735097	5.62
4,4'-DDE	10.00	9.93	10.07	35585378	32874162	7.62
Dieldrin	9.92	9.85	9.99	35333976	33753531	4.47
Endrin	10.47	10.40	10.54	30230453	30439365	0.69
Endosulfan II	10.72	10.65	10.79	30776202	28447037	7.57
4,4'-DDD	11.11	11.04	11.18	30805692	27524042	10.65
Endrin aldehyde	11.24	11.17	11.31	21862156	21275811	2.68
Endosulfan sulfate	11.90	11.83	11.97	27807031	26557424	4.49
4,4'-DDT	12.13	12.06	12.20	20458575	22236926	8.69
Endrin ketone	13.10	13.03	13.17	33539155	28565119	14.83
Methoxychlor	13.80	13.73	13.87	10372533	10767745	3.81
alpha-Chlordane	9.34	9.27	9.41	35976493	32894183	8.57
gamma-Chlordane	8.93	8.86	9.00	37752163	34752231	7.95
Average %D						6.20
Chlordane 500 ppb	8.94	8.87	9.01	4171440	3923841	5.94
Chlordane {2}	9.13	9.06	9.20	887533	848649	4.38
Chlordane {3}	9.34	9.27	9.41	3922407	3767105	3.96
Chlordane {4}	9.48	9.41	9.55	2442430	2257732	7.56
Chlordane {5}	11.32	11.25	11.39	1050809	1048469	0.22
Toxaphene 500 ppb	11.67	11.60	11.74	485936	533852	9.86
Toxaphene {2}	12.18	12.11	12.25	822837	803456	2.36
Toxaphene {3}	13.03	12.96	13.10	917851	777246	15.32
Toxaphene {4}	13.25	13.18	13.32	763741	755590	1.07
Toxaphene {5}	14.28	14.21	14.35	479631	477121	0.52
Average %D						5.12

PESTICIDE CALIBRATION VERIFICATION SUMMARY

Date Analyzed: 10/16/01

Instrument ID: GC-W

Data File: W8923.C

GC Column (2nd): DB-1701P

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
alpha-BHC	5.18	5.11	5.25	14827598	15469447	4.33
beta-BHC	8.16	8.09	8.23	6097231	6066596	0.50
gamma-BHC	6.10	6.03	6.17	13546829	14187021	4.73
delta-BHC	8.70	8.63	8.77	13321769	13966681	4.84
Heptachlor	6.56	6.49	6.63	12120237	13181412	8.76
Aldrin	7.20	7.13	7.27	12390069	13026534	5.14
Heptachlor epoxide	9.01	8.94	9.08	11217666	11631074	3.69
Endosulfan I	9.68	9.61	9.75	10488445	10773521	2.72
4,4'-DDE	10.46	10.39	10.53	11254895	11399185	1.28
Dieldrin	10.70	10.63	10.77	11419812	11751939	2.91
Endrin	11.21	11.14	11.28	10631753	11217130	5.51
Endosulfan II	12.64	12.57	12.71	10073067	10081928	0.09
4,4'-DDD	12.69	12.62	12.76	9895694	9237954	6.65
Endrin aldehyde	13.78	13.71	13.85	6948408	6739933	3.00
Endosulfan sulfate	14.68	14.61	14.75	8898298	8866614	0.36
4,4'-DDT	13.11	13.04	13.18	7083013	6812720	3.82
Endrin ketone	15.68	15.61	15.75	10731417	10161140	5.31
Methoxychlor	14.98	14.91	15.05	3800820	4099976	7.87
alpha-Chlordane	10.07	10.00	10.14	11496547	11551923	0.48
gamma-Chlordane	9.92	9.85	9.99	11757803	11919945	1.38
Average %D						3.67
Chlordane 500 ppb	7.88	7.81	7.95	214161	218768	2.15
Chlordane {2}	9.53	9.46	9.60	286639	293843	2.51
Chlordane {3}	9.92	9.85	9.99	1309951	1342124	2.46
Chlordane {4}	10.07	10.00	10.14	1142675	1165236	1.97
Chlordane {5}	10.13	10.06	10.20	723821	737938	1.95
Toxaphene 500 ppb	12.72	12.65	12.79	298191	298847	0.22
Toxaphene {2}	13.42	13.35	13.49	281068	265851	5.41
Toxaphene {3}	14.41	14.34	14.48	291954	267957	8.22
Toxaphene {4}	14.60	14.53	14.67	143654	131903	8.18
Toxaphene {5}	15.04	14.97	15.11	147283	134305	8.81
Average %D						4.19

PESTICIDE CALIBRATION VERIFICATION SUMMARY

Date Analyzed: 10/16/01

Instrument ID: GC-W

Data File: W8941.D

GC Column (1st): DB-5

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
alpha-BHC	4.36	4.29	4.43	46976823	50389675	7.26
beta-BHC	4.84	4.77	4.91	17456924	17905035	2.57
gamma-BHC	4.97	4.90	5.04	41669009	44180960	6.03
delta-BHC	5.44	5.37	5.51	42034378	45631651	8.56
Heptachlor	6.52	6.45	6.59	34934548	38088745	9.03
Aldrin	7.33	7.26	7.40	39449916	41917543	6.26
Heptachlor epoxide	8.33	8.26	8.40	34447512	37022831	7.48
Endosulfan I	9.24	9.17	9.31	32565698	34696256	6.54
4,4'-DDE	10.00	9.93	10.07	35585378	37606821	5.68
Dieldrin	9.92	9.85	9.99	35333976	38261464	8.29
Endrin	10.47	10.40	10.54	30230453	31863003	5.40
Endosulfan II	10.73	10.66	10.80	30776202	32487619	5.56
4,4'-DDD	11.11	11.04	11.18	30805692	31918069	3.61
Endrin aldehyde	11.25	11.18	11.32	21862156	23747233	8.62
Endosulfan sulfate	11.90	11.83	11.97	27807031	30018216	7.95
4,4'-DDT	12.13	12.06	12.20	20458575	24200771	18.29
Endrin ketone	13.10	13.03	13.17	33539155	35556458	6.01
Methoxychlor	13.81	13.74	13.88	10372533	12008260	15.77
alpha-Chlordane	9.34	9.27	9.41	35976493	37683983	4.75
gamma-Chlordane	8.94	8.87	9.01	37752163	39863792	5.59
Average.%D						7.46

PESTICIDE CALIBRATION VERIFICATION SUMMARY

Date Analyzed: 10/16/01

Instrument ID: GC-W

Data File: W8941.C

GC Column (2nd): DB-1701P

Compound	RT	RT WI NDOW		Avg CF	CC CF	%D
		FROM	TO			
alpha-BHC	5.18	5.11	5.25	14827598	16519318	11.41
beta-BHC	8.16	8.09	8.23	6097231	6480915	6.29
gamma-BHC	6.10	6.03	6.17	13546829	15063967	11.20
delta-BHC	8.70	8.63	8.77	13321769	15112169	13.44
Heptachlor	6.56	6.49	6.63	12120237	13608581	12.28
Aldrin	7.20	7.13	7.27	12390069	13730616	10.82
Heptachlor epoxide	9.01	8.94	9.08	11217666	12330256	9.92
Endosulfan I	9.68	9.61	9.75	10488445	11456875	9.23
4,4'-DDE	10.46	10.39	10.53	11254895	12344824	9.68
Dieldrin	10.70	10.63	10.77	11419812	12596225	10.30
Endrin	11.21	11.14	11.28	10631753	11259849	5.91
Endosulfan II	12.64	12.57	12.71	10073067	11001291	9.21
4,4'-DDD	12.69	12.62	12.76	9895694	10387267	4.97
Endrin aldehyde	13.78	13.71	13.85	6948408	7577796	9.06
Endosulfan sulfate	14.68	14.61	14.75	8898298	9754556	9.62
4,4'-DDT	13.11	13.04	13.18	7083013	8153197	15.11
Endrin ketone	15.68	15.61	15.75	10731417	11674239	8.79
Methoxychlor	14.98	14.91	15.05	3800820	4357271	14.64
alpha-Chlordane	10.07	10.00	10.14	11496547	12512231	8.83
gamma-Chlordane	9.92	9.85	9.99	11757803	12858065	9.36
Average %D						10.00

PESTICIDE SURROGATE PERCENT RECOVERY SUMMARY

Date Analyzed: 10/16/2001

Client ID	Lab Sample ID	Matrix	TCMX 1		DCB 1		TCMX 2		DCB 2	
			% rec	#	% rec	#	% rec	#	% rec	#
(7023.7041)	1015-BK1	SOIL	94		66		103		76	
ABSEGAMI	7035-001	SOIL	90		63		98		72	
5-F1	7041-001	SOIL	96		66		103		76	
UG-1_10-10.5	7023-001	SOIL	98		69		107		79	
UG-2_9-9.5	7023-002	SOIL	94		66		102		76	
FO-1_6-6.5	7023-004	SOIL	63		64		81		75	
DF-1_10-10.5	7023-005	SOIL	97		68		103		74	
DF-2_6-6.5	7023-006	SOIL	83		63		96		75	
DF-3_10-10.5	7023-007	SOIL	84		65		88		72	
DF-4_8.5-9	7023-008	SOIL	84		64		91		74	
DW-1_5-5.5	7023-010	SOIL	62		67		81		74	
DW-2_8-8.5	7023-011	SOIL	71		68		79		74	
PEST	7041-001-MS1	SOIL	97		72		100		77	
PEST	7041-001-MSD1	SOIL	101		75		103		80	
PEST	PBS1015-MS1	SOIL	102		77		105		82	

Surrogate QC Limits

TCMX = Tetrachloro-m-xylene

DCB = Decachlorobiphenyl

Soil

39-147

33-179

Aqueous

39-151

30-150

Column to be used to flag recovery values

* Values outside of QC limits

D Surrogate diluted out

M Matrix interference

SOIL PESTICIDES MATRIX SPIKE/SPIKE DUPLICATE RECOVERY

Matrix spike Lab sample ID: 7041-001-MSD1

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
gamma-BHC	100.0	0.0	113.2	113	44 - 149
Heptachlor	100.0	0.0	111.2	111	50 - 157
Aldrin	100.0	0.0	114.0	114	40 - 150
Dieldrin	100.0	0.0	112.9	113	56 - 134
Endrin	100.0	0.0	118.9	119	65 - 148
4,4'-DDT	100.0	0.0	112.7	113	42 - 175

Compound	SAMPLE CONC. (ug/Kg)	MSD CONC. (ug/Kg)	MSD % # REC	% RPD #	QC LIMITS	
					RPD	REC.
gamma-BHC	0.0	115.5	116	3	18	44 - 149
Heptachlor	0.0	115.8	116	4	18	50 - 157
Aldrin	0.0	116.3	116	2	18	40 - 150
Dieldrin	0.0	114.8	115	2	13	56 - 134
Endrin	0.0	120.7	121	2	14	65 - 148
4,4'-DDT	0.0	116.9	117	3	27	42 - 175

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

RPD: 0 out of 6 outside limits

Spike Recovery: 0 out of 12 outside limits

SOIL PESTICIDES BLANK SPIKE RECOVERY

Matrix spike Lab sample ID: PBS1015-MS1

Compound	SPIKE ADDED (ug/Kg)	SAMPLE CONC. (ug/Kg)	MS CONC. (ug/Kg)	MS % REC #	QC LIMITS REC.
gamma-BHC	100.0	0.0	116.9	117	44 - 149
Heptachlor	100.0	0.0	121.7	122	50 - 157
Aldrin	100.0	0.0	121.4	121	40 - 150
Dieldrin	100.0	0.0	118.8	119	56 - 134
Endrin	100.0	0.0	119.5	120	65 - 148
4,4'-DDT	100.0	0.0	124.1	124	42 - 175

Column to be used to flag recovery and RPD values with an asterisk

* Values outside of QC limits

NC Non calculable

Spike Recovery: 0 out of 6 outside limits

PESTICIDE RETENTION TIME SHIFT SUMMARY

Instrument ID: GC-W

Column: DB-5/DB-1701P

Surrogate RT from initial calibration :

TCMX 1 3.67 DCB 1 18.40 TCMX 2 3.38 DCB 2 18.02

Client ID	Lab Sample ID	Date Analyzed	Time Analyzed	TCMX 1 RT	DCB 1 RT	TCMX 2 RT	DCB 2 RT
(7023.7041)	1015-BK1	10/16/2001	14:33	3.67	18.40	3.38	18.02
ABSEGAMI	7035-001	10/16/2001	14:58	3.69	18.41	3.38	18.02
5-F1	7041-001	10/16/2001	15:33	3.68	18.40	3.38	18.02
UG-1_10-10.5	7023-001	10/16/2001	15:58	3.69	18.41	3.38	18.02
UG-2_9-9.5	7023-002	10/16/2001	16:23	3.69	18.41	3.38	18.02
FO-1_6-6.5	7023-004	10/16/2001	16:48	3.69	18.41	3.38	18.02
DF-1_10-10.5	7023-005	10/16/2001	17:13	3.69	18.41	3.38	18.02
DF-2_6-6.5	7023-006	10/16/2001	17:38	3.69	18.40	3.39	18.02
DF-3_10-10.5	7023-007	10/16/2001	18:03	3.69	18.41	3.38	18.02
DF-4_8.5-9	7023-008	10/16/2001	18:28	3.68	18.41	3.38	18.02
DW-1_5-5.5	7023-010	10/16/2001	18:53	3.69	18.41	3.38	18.02
DW-2_8-8.5	7023-011	10/16/2001	19:18	3.69	18.41	3.38	18.02
PEST	7041-001-MS1	10/16/2001	19:43	3.69	18.41	3.38	18.02
PEST	7041-001-MSD1	10/16/2001	20:08	3.69	18.41	3.38	18.02
PEST	PBS1015-MS1	10/16/2001	20:33	3.69	18.41	3.38	18.02

Surrogate QC Limits

TCMX = Tetrachloro-m-xylene

($\pm$ 0.05 Minutes)

DCB = Decachlorobiphenyl

($\pm$ 0.05 Minutes)

Column to be used to flag recovery values

* Values outside of QC limits

D Surrogate diluted out

M Matrix interference

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8928.D\ECD1B.CH Vial: 7
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8928.D\ECD2A.CH
 Acq On : 10-16-01 15:33:18 Operator: LAG
 Sample : 5-F1,7041-001,S,5.7g,9.4,10/15/01,4 Inst : GC_W
 Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:30 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

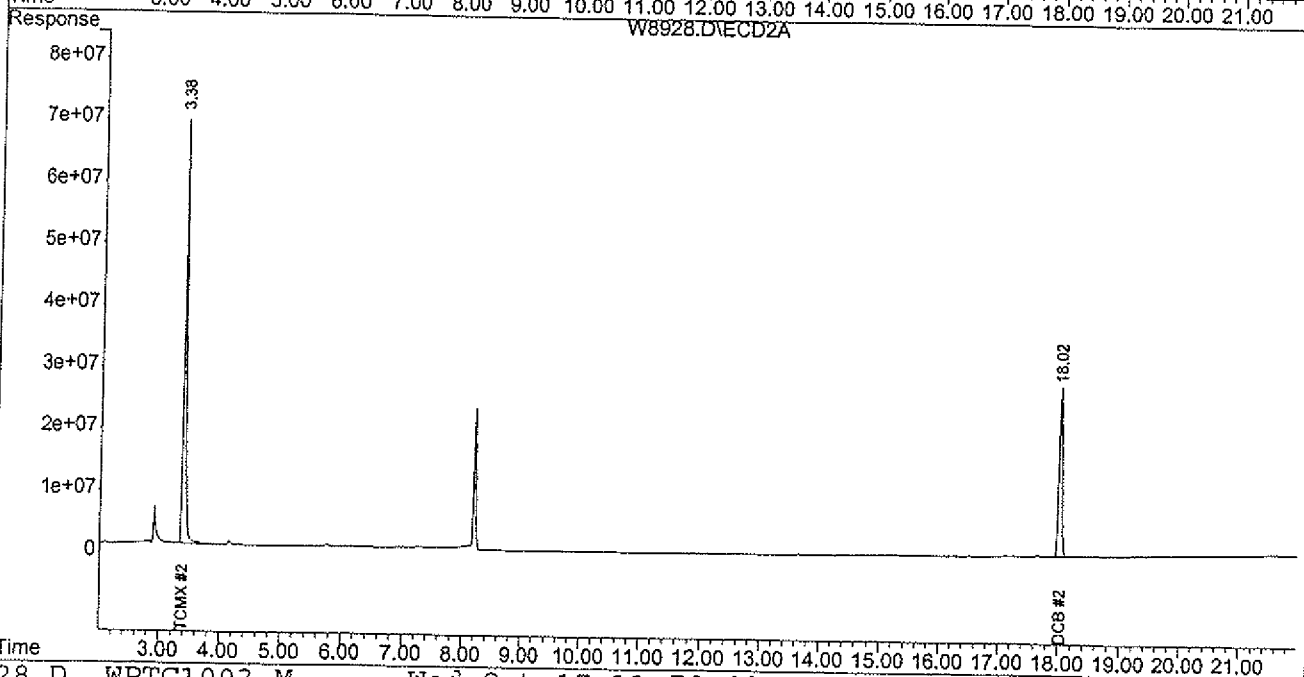
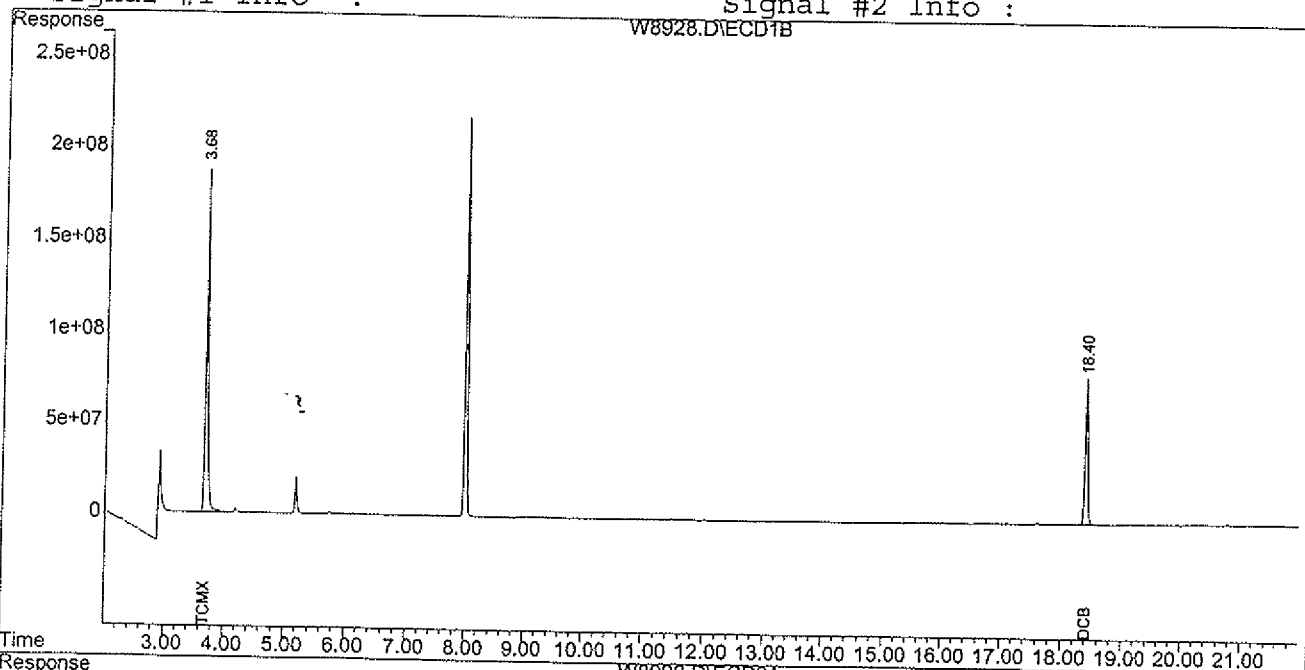
Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	3.68	3.38	5256.6E6	1903.6E6	95.529	102.464
Spiked Amount	100.000		Recovery	=	95.53%	102.46%
2) S DCB	18.40	18.02	2664.5E6	968.4E6	66.190	75.691
Spiked Amount	100.000		Recovery	=	66.19%	75.69%
Target Compounds						
Sum Chlordane			0	0	N.D.	N.D.
Average Chlordane					0.000	0.000
Sum Toxaphene			0	0	N.D.	N.D.
Average Toxaphene					0.000	0.000

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8928.D\ECD1B.CH Vial: 7
Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8928.D\ECD2A.CH
Acq On : 10-16-01 15:33:18 Operator: LAG
Sample : S-F1,7041-001,S,5.7g,9.4,10/15/01,4 Inst : GC_W
Misc : BRENNAN/99704_FINETEX,10/11/01,10/11/01, Multiplr: 1.00
IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
Quant Time: Oct 17 11:30 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
Title :
Last Update : Thu Oct 04 10:37:06 2001
Response via : Multiple Level Calibration
DataAcq Meth : WPTC1003.M

Volume Inj. :
Signal #1 Phase :
Signal #1 Info :
Signal #2 Phase :
Signal #2 Info :



Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8926.D\ECD1B.CH Vial: 5
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8926.D\ECD2A.CH
 Acq On : 10-16-01 14:33:45 Operator: LAG
 Sample : (7023.7041),1015-BK1,S,5.9g,0.0,10/15/01 Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:30 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	3.67	3.38	5182.6E6	1910.8E6	94.184	102.855
Spiked Amount	100.000			Recovery	=	94.18%
2) S DCB	18.40	18.02	2670.3E6	972.7E6	66.333	76.029
Spiked Amount	100.000			Recovery	=	66.33%
Target Compounds						
Sum Chlordane			0	0	N.D.	N.D.
Average Chlordane					0.000	0.000
Sum Toxaphene			0	0	N.D.	N.D.
Average Toxaphene					0.000	0.000

 (f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.

W8926.D WPTC1003.M

Wed Oct 17 11:58:15 2001

GC_W

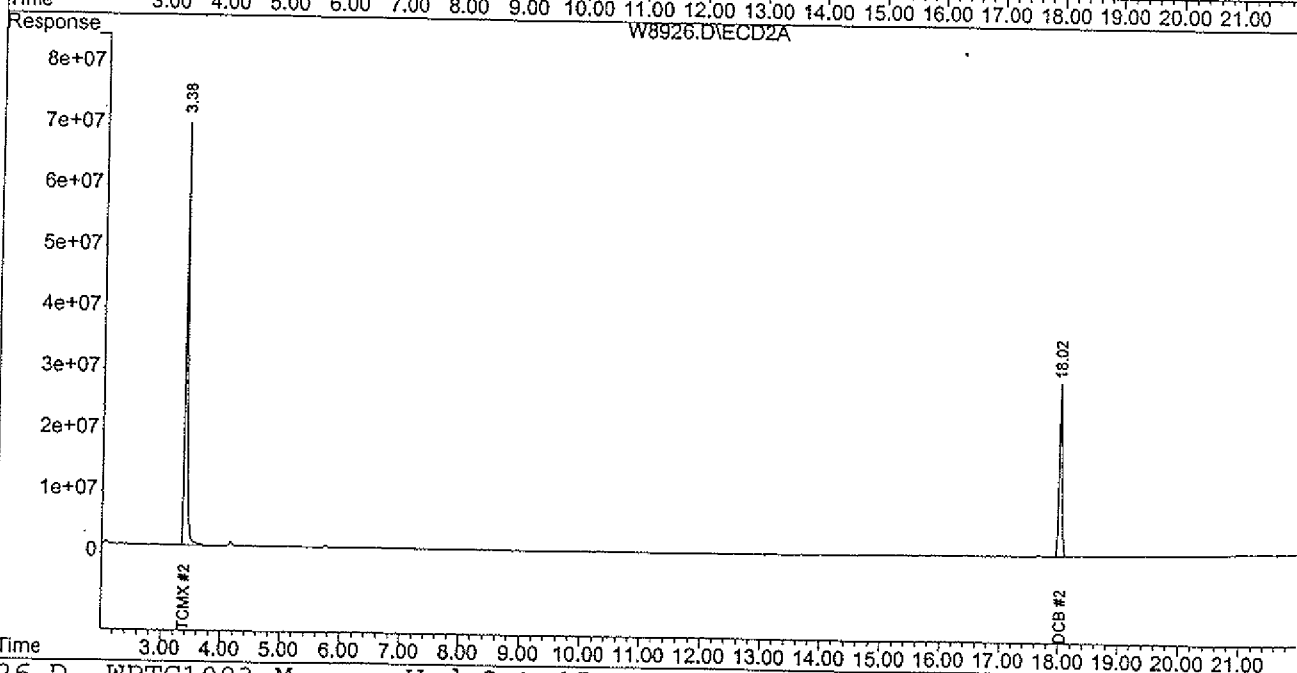
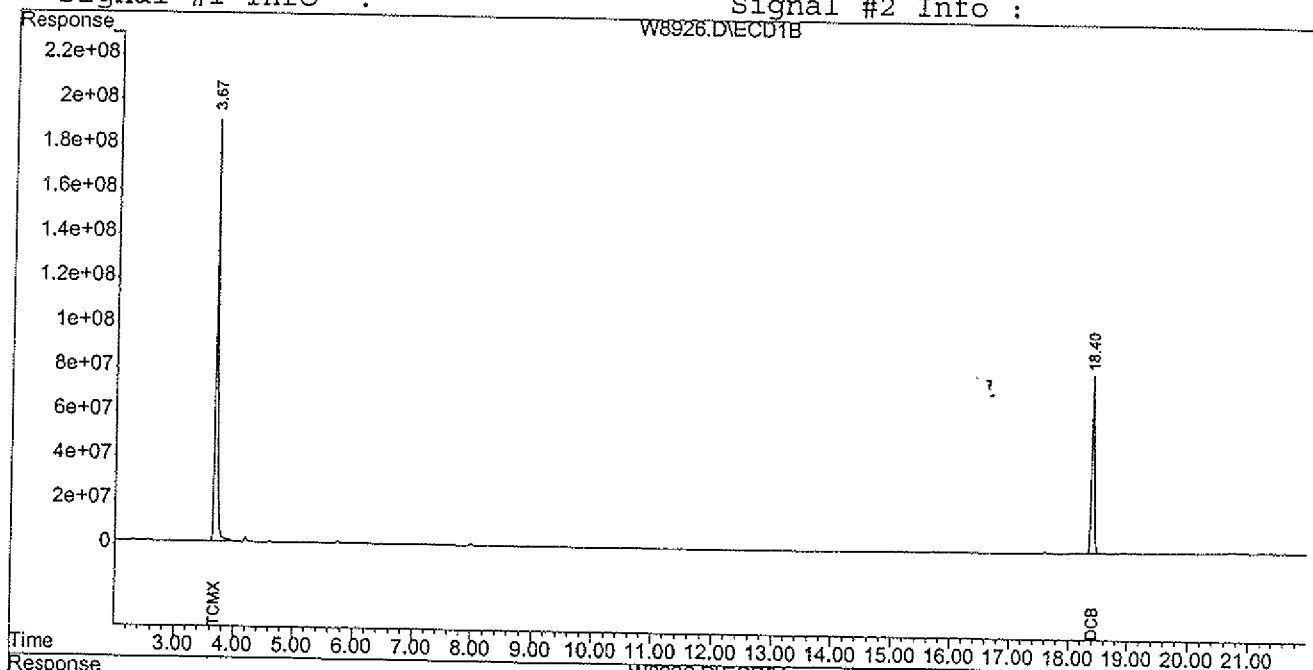
Page 1

Quantitation Report (QT Reviewed)

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8926.D\ECD1B.CH Vial: 5
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8926.D\ECD2A.CH
 Acq On : 10-16-01 14:33:45 Operator: LAG
 Sample : (7023.7041),1015-BK1,S,5.9g,0.0,10/15/01 Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:30 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8923.D\ECD1B.CH Vial: 2
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8923.D\ECD2A.CH
 Acq On : 10-16-01 12:46:04 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 16 14:28 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	3.69	3.39	5141.1E6	1909.4E6	93.430	102.776
Spiked Amount	100.000		Recovery	=	93.43%	102.78%
2) S DCB	18.40	18.02	3100.4E6	1137.9E6	77.018	88.943
Spiked Amount	100.000		Recovery	=	77.02%	88.94%
Target Compounds						
3) T alpha-BHC	4.35	5.18	4452.3E6	1546.9E6	94.777	104.329
4) T beta-BHC	4.84	8.16	1586.1E6	606.7E6	90.856	99.498
5) T gamma-BHC	4.97	6.10	3960.0E6	1418.7E6	95.035	104.726
6) T delta-BHC	5.44	8.70	4001.9E6	1396.7E6	95.206	104.841
7) T Heptachlor	6.52	6.56	3558.9E6	1318.1E6	101.874	108.755
8) T Aldrin	7.33	7.20	3710.0E6	1302.7E6	94.044	105.137
9) T Heptachlor epoxi	8.33	9.01	3295.7E6	1163.1E6	95.674	103.685
10) T Endosulfan I	9.23	9.68	3073.5E6	1077.4E6	94.379	102.718
11) T 4,4'-DDE	10.00	10.46	3287.4E6	1139.9E6	92.381	101.282
12) T Dieldrin	9.92	10.70	3375.4E6	1175.2E6	95.527	102.908
13) T Endrin	10.47	11.21	3043.9E6	1121.7E6	100.691	105.506
14) T Endosulfan II	10.72	12.64	2844.7E6	1008.2E6	92.432	100.088
15) T 4,4'-DDD	11.11	12.69	2752.4E6	923.8E6	89.347	93.353
16) T Endrin aldehyde	11.24	13.78	2127.6E6	674.0E6	97.318	97.000
17) T Endosulfan sulfa	11.90	14.68	2655.7E6	886.7E6	95.506	99.644
18) T 4,4'-DDT	12.13	13.11	2223.7E6	681.3E6	108.692	96.184
19) T Endrin ketone	13.10	15.68	2856.5E6	1016.1E6	85.169	94.686
20) T Methoxychlor	13.80	14.98	1076.8E6	410.0E6	103.810	107.871
21) T alpha-Chlordane	9.34	10.07	3289.4E6	1155.2E6	91.432	100.482
22) T gamma-Chlordane	8.93	9.92	3475.2E6	1192.0E6	92.054	101.379
Sum Chlordane			0	0	N.D.	N.D.
Average Chlordane					0.000	0.000
Sum Toxaphene			0	0	N.D.	N.D.
Average Toxaphene					0.000	0.000

(f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.

W8923.D WPTC1003.M

Wed Oct 17 11:24:17 2001

GC_W

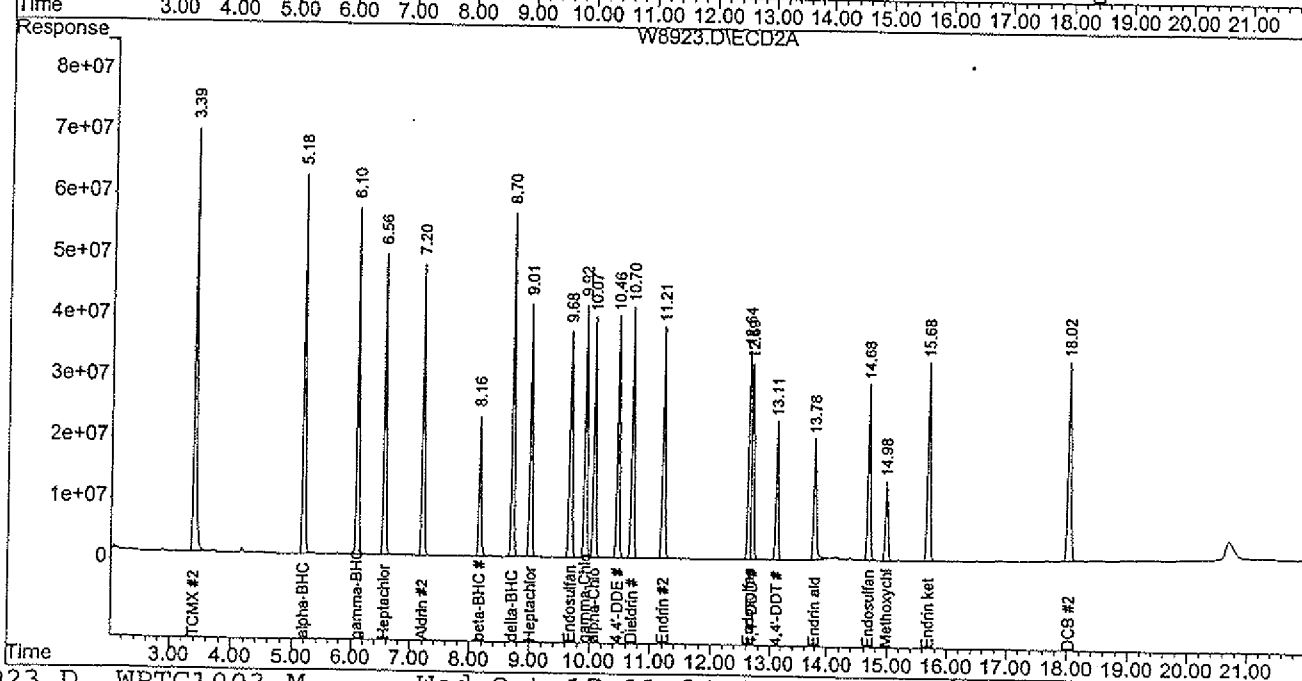
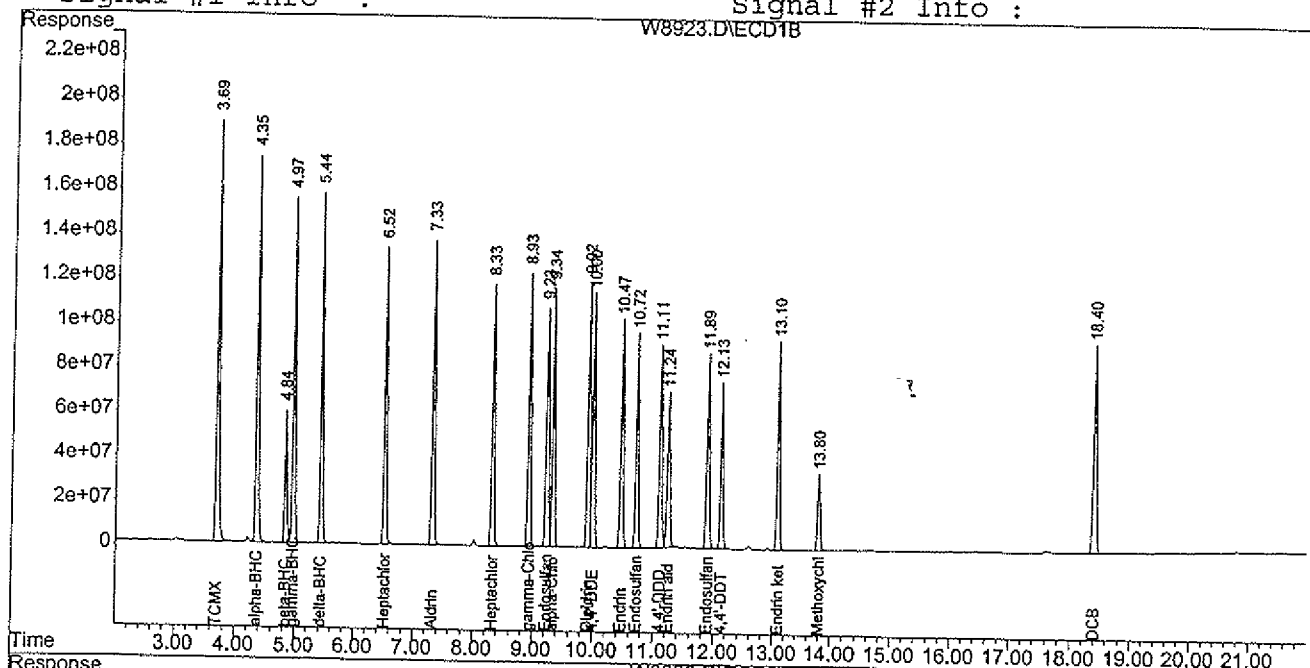
Page 1

Quantitation Report (QT Reviewed)

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8923.D\ECD1B.CH Vial: 2
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8923.D\ECD2A.CH
 Acq On : 10-16-01 12:46:04 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 16 14:28 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD1B.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint1.e

Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD2A.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint2.e

Quant Time: Oct 17 9:29 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	3.70	3.40	5687.7E6	1988.0E6	103.362	107.010
Spiked Amount	100.000		Recovery	=	103.36%	107.01%
2) S DCB	18.41	18.02	3807.1E6	1298.6E6	94.573	101.500
Spiked Amount	100.000		Recovery	=	94.57%	101.50%
Target Compounds						
3) T alpha-BHC	4.36	5.18	5039.0E6	1651.9E6	107.265	111.409
4) T beta-BHC	4.84	8.16	1790.5E6	648.1E6	102.567	106.293
5) T gamma-BHC	4.97	6.10	4418.1E6	1506.4E6	106.028	111.199
6) T delta-BHC	5.44	8.70	4563.2E6	1511.2E6	108.558	113.440
7) T Heptachlor	6.52	6.56	3808.9E6	1360.9E6	109.029	112.280
8) T Aldrin	7.33	7.20	4191.8E6	1373.1E6	106.255	110.820
9) T Heptachlor epoxi	8.33	9.01	3702.3E6	1233.0E6	107.476	109.918
10) T Endosulfan I	9.24	9.68	3469.6E6	1145.7E6	106.542	109.233
11) T 4,4'-DDE	10.00	10.46	3760.7E6	1234.5E6	105.681	109.684
12) T Dieldrin	9.92	10.70	3826.1E6	1259.6E6	108.285	110.302
13) T Endrin	10.47	11.21	3186.3E6	1126.0E6	105.400	105.908
14) T Endosulfan II	10.73	12.64	3248.8E6	1100.1E6	105.561	109.215
15) T 4,4'-DDD	11.11	12.69	3191.8E6	1038.7E6	103.611	104.968
16) T Endrin aldehyde	11.25	13.78	2374.7E6	757.8E6	108.623	109.058
17) T Endosulfan sulfa	11.90	14.68	3001.8E6	975.5E6	107.952	109.623
18) T 4,4'-DDT	12.13	13.11	2420.1E6	815.3E6	118.292	115.109
19) T Endrin ketone	13.10	15.68	3555.6E6	1167.4E6	106.015	108.786
20) T Methoxychlor	13.81	14.98	1200.8E6	435.7E6	115.770	114.640
21) T alpha-Chlordane	9.34	10.07	3768.4E6	1251.2E6	104.746	108.835
22) T gamma-Chlordane	8.94	9.92	3986.4E6	1285.8E6	105.593	109.358
Sum Chlordane			0	0	N.D.	N.D.
Average Chlordane					0.000	0.000
Sum Toxaphene			0	0	N.D.	N.D.
Average Toxaphene					0.000	0.000

(f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.

W8941.D WPTC1003.M

Wed Oct 17 11:57:07 2001

GC_W

Page 1

Quantitation Report (QT Reviewed)

Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD1B.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint1.e

Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD2A.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint2.e

Quant Time: Oct 17 9:29 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase : Signal #2 Phase:
 Signal #1 Info : Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2
----------	------	------	--------	--------	------	------

Quantitation Report (QT Reviewed)

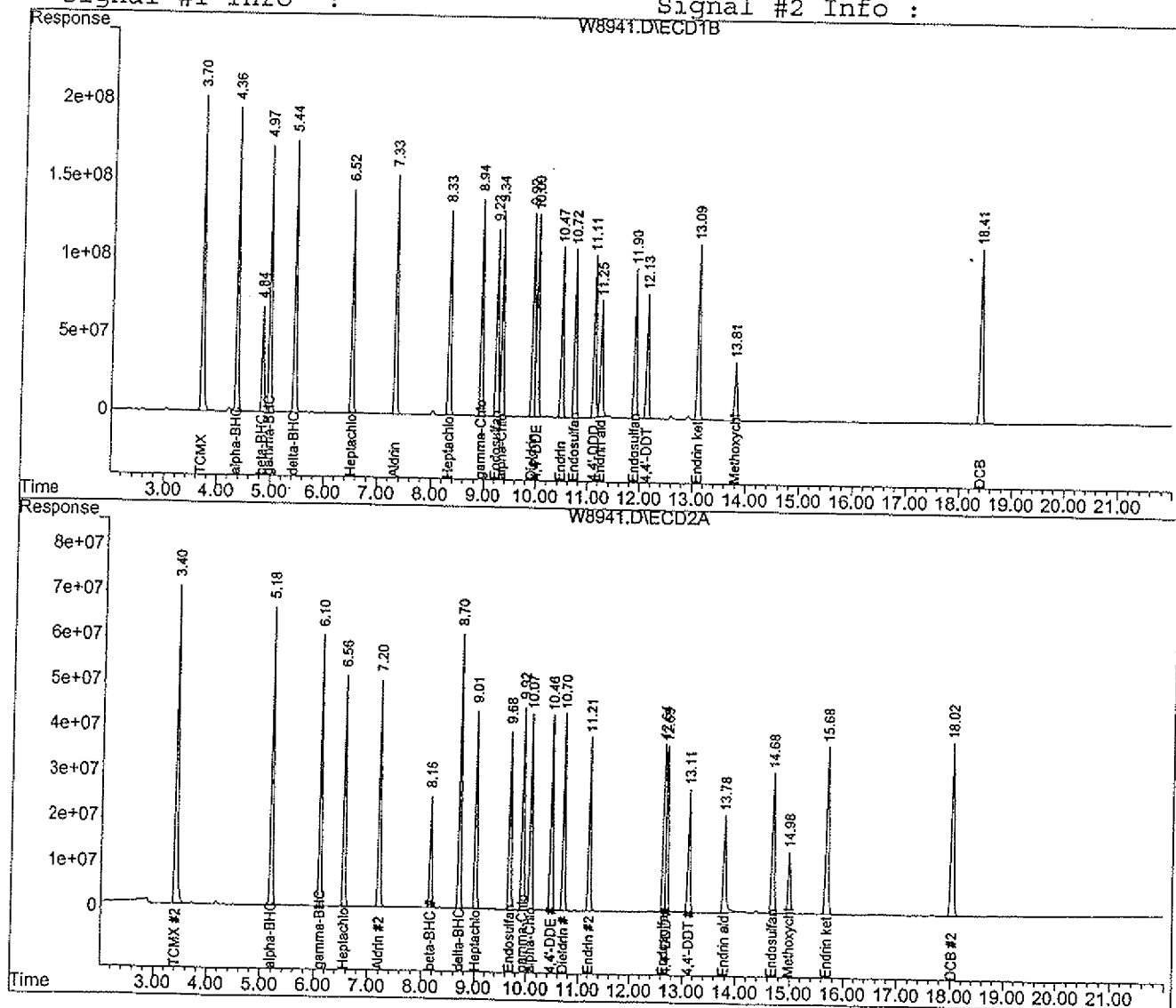
Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD1B.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint1.e

Data File : C:\HPCHEM\1\DATA\10-16-01\W8941.D\ECD2A.CH Vial: 20
 Acq On : 10-16-01 20:58:34 Operator: LAG
 Sample : PEST_C_IAS_1554,100_PPB Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile : autoint2.e

Quant Time: Oct 17 9:29 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



Quantitation Report (QT Reviewed)

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8924.D\ECD1B.CH Vial: 3
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8924.D\ECD2A.CH
 Acq On : 10-16-01 13:11:08 Operator: LAG
 Sample : TOX_C_IAS_1533,0.5_PPM Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:25 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase:
 Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

System Monitoring Compounds						
1) S TCMX	3.70	3.39	5562.3E6	2029.4E6	101.085	109.238
Spiked Amount	100.000					
				Recovery	=	101.09%
2) S DCB	18.41	18.02	2500.0E6	884.0E6	62.103	69.094m
Spiked Amount	100.000					
				Recovery	=	62.10%
Target Compounds						
Sum Chlordane			0	0	N.D.	N.D.
Average Chlordane					0.000	0.000
28) L2 Toxaphene	11.67	12.72	266.9E6	149.4E6	549.302m	501.099
29) L2 Toxaphene {2}	12.18	13.42	401.7E6	132.9E6	488.223	472.930
30) L2 Toxaphene {3}	13.03	14.41	388.6E6	134.0E6	423.405	458.902
31) L2 Toxaphene {4}	13.25	14.60	377.8E6	65951530	494.663	459.100
32) L2 Toxaphene {5}	14.28	15.04	238.6E6	67152370	497.384	455.941
Sum Toxaphene			1673.6E6	549.4E6	2452.978	2347.973
Average Toxaphene					490.596	469.595

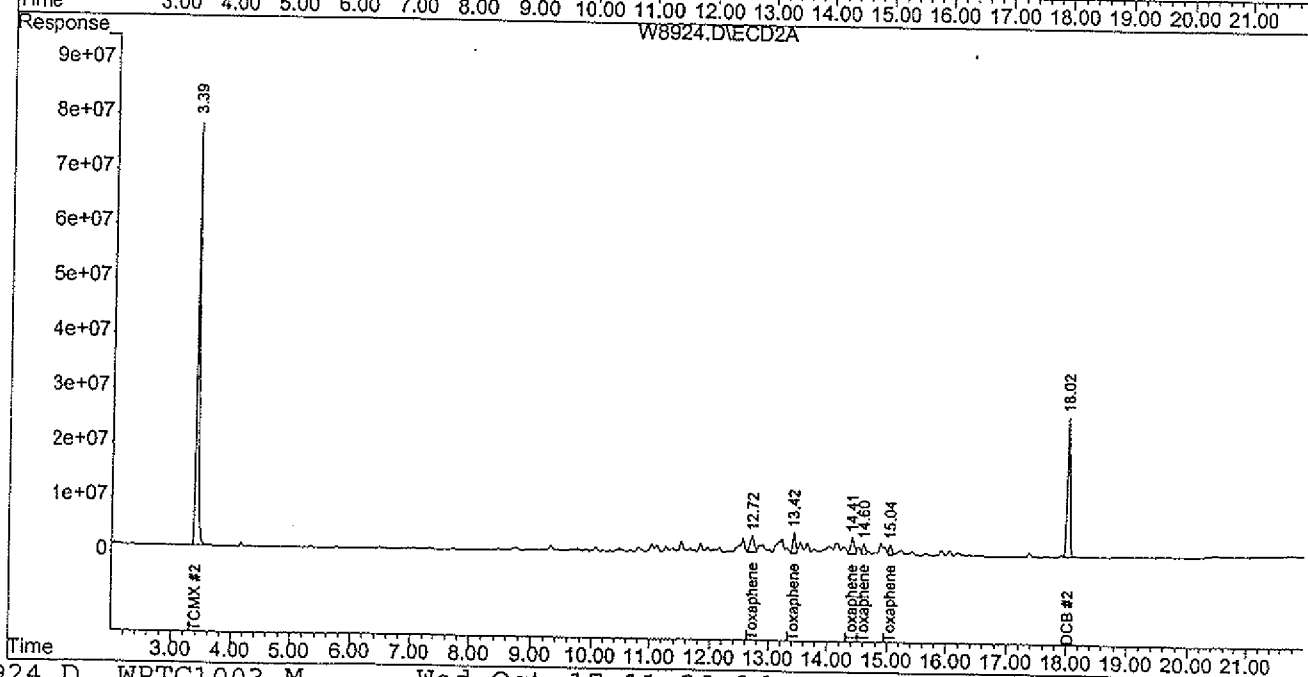
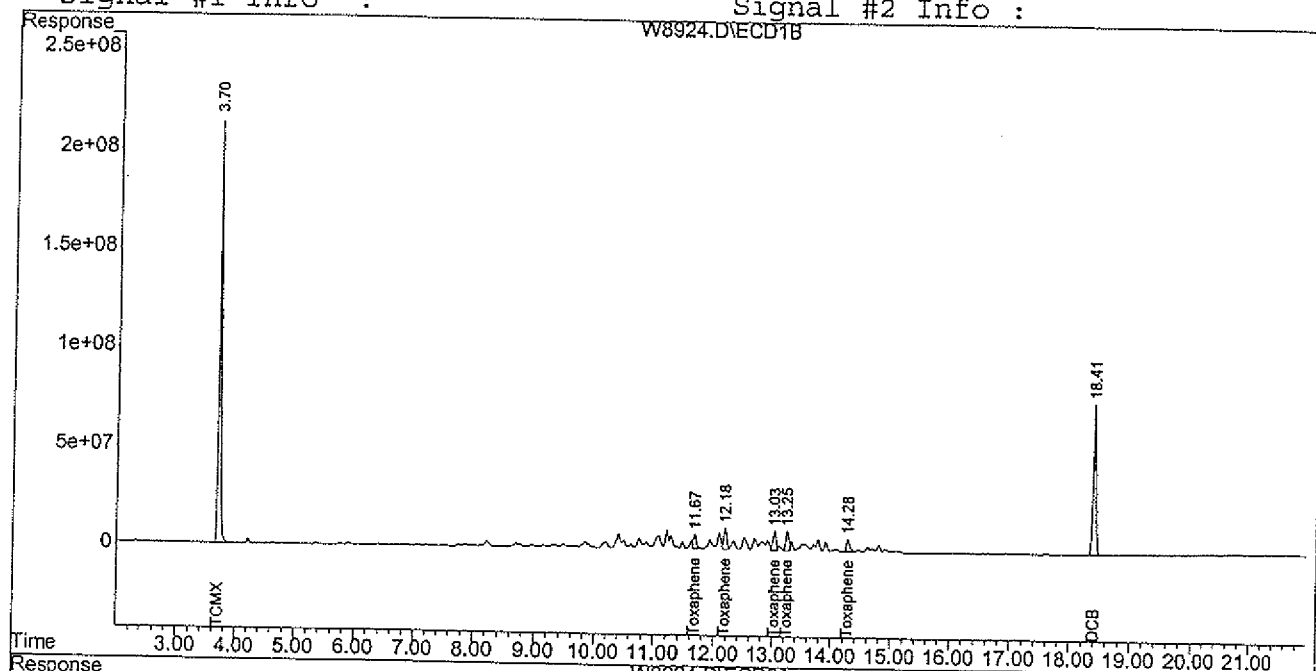
 (f)=RT Delta > 1/2 Window (#)=Amounts differ by > 25% (m)=manual int.
 W8924.D WPTC1003.M Wed Oct 17 11:28:34 2001 GC_W Page 1

Quantitation Report (QT Reviewed)

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8924.D\ECD1B.CH Vial: 3
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8924.D\ECD2A.CH
 Acq On : 10-16-01 13:11:08 Operator: LAG
 Sample : TOX_C_IAS_1533,0.5_PPM Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:25 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8925.D\ECD1B.CH Vial: 4
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8925.D\ECD2A.CH
 Acq On : 10-16-01 13:36:22 Operator: LAG
 Sample : CHLOR_C_IAS_1534,0.5_PPM Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:29 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Initial Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :

Compound	RT#1	RT#2	Resp#1	Resp#2	ng#1	ng#2

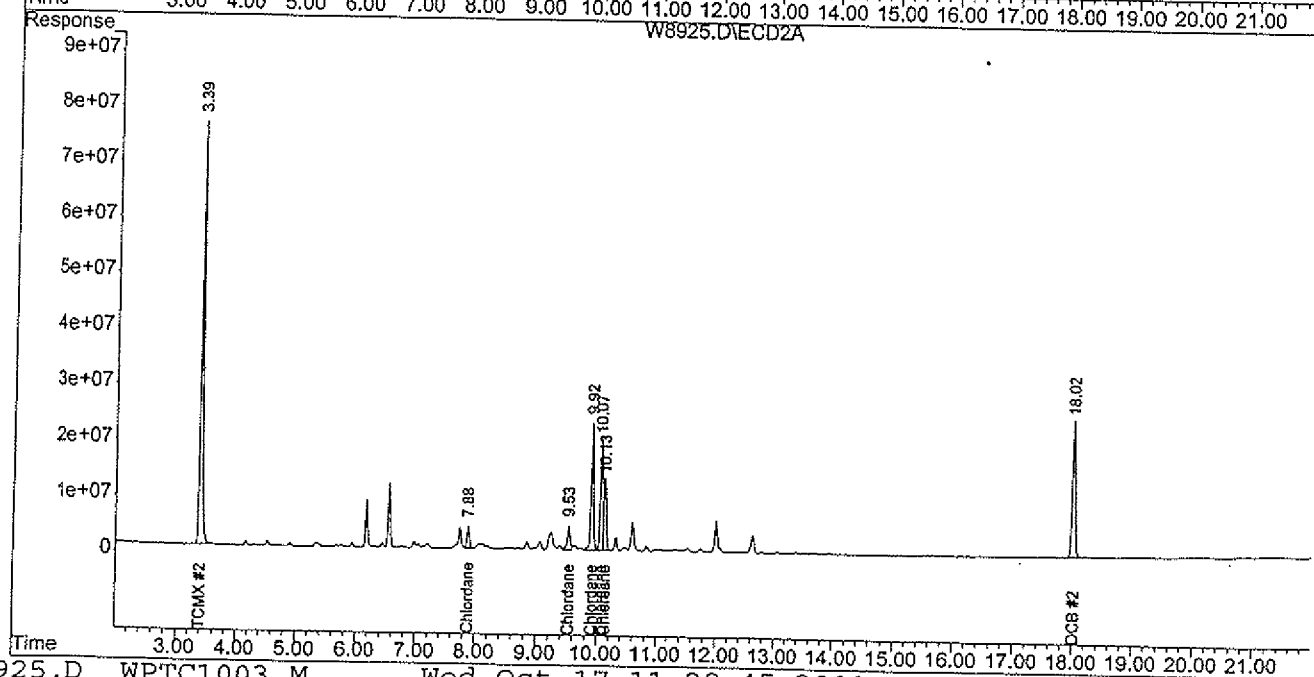
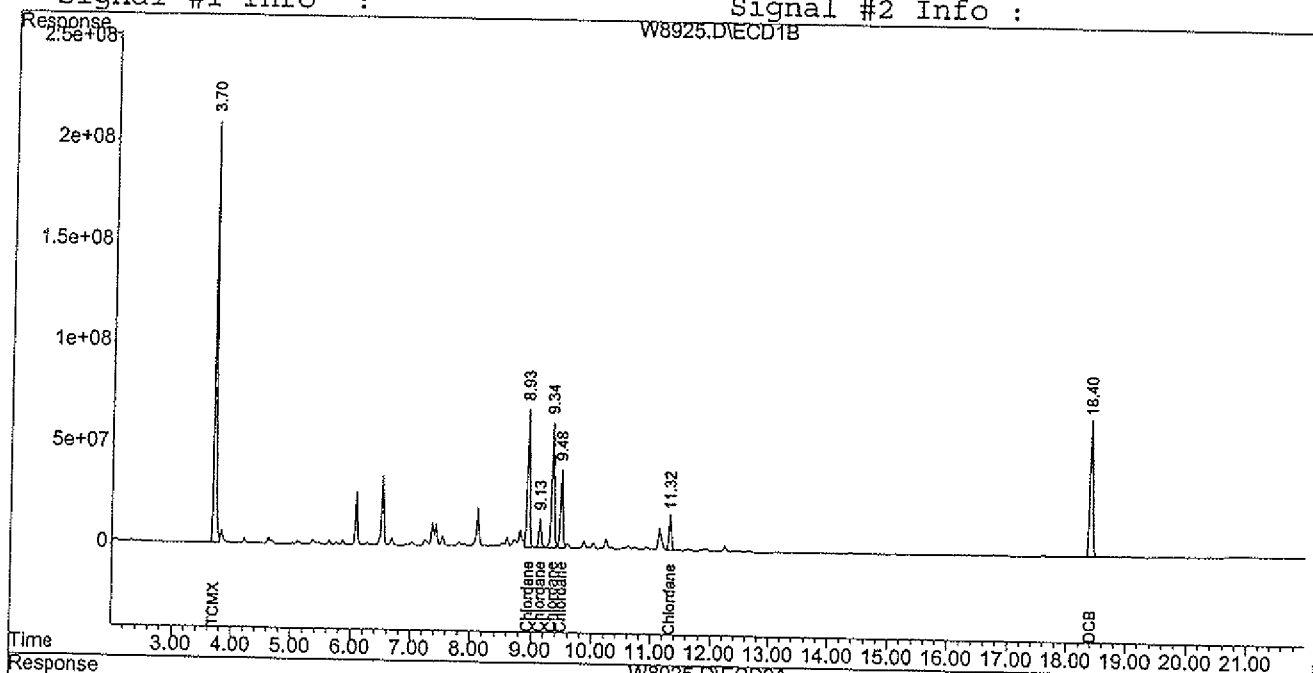
System Monitoring Compounds						
1) S TCMX	3.70	3.39	5232.2E6	1948.4E6	95.085	104.880
Spiked Amount	100.000		Recovery	=	95.08%	104.88%
2) S DCB	18.41	18.02	2311.9E6	864.0E6	57.430	67.533
Spiked Amount	100.000		Recovery	=	57.43%	67.53%
Target Compounds						
23) L1 Chlordane	8.94	7.88	1961.9E6	109.4E6	470.322	510.757
24) L1 Chlordane {2}	9.13	9.53	424.3E6	146.9E6	478.094	512.567
25) L1 Chlordane {3}	9.34	9.92	1883.6E6	671.1E6	480.203	512.280
26) L1 Chlordane {4}	9.48	10.07	1128.9E6	582.6E6	462.190	509.872
27) L1 Chlordane {5}	11.32	10.13	524.2E6	369.0E6	498.887	509.752
Sum Chlordane			5922.9E6	1879.0E6	2389.696	2555.228
Average Chlordane					477.939	511.046
Sum Toxaphene			0	0	N.D.	N.D.
Average Toxaphene					0.000	0.000

Quantitation Report (QT Reviewed)

Signal #1 : C:\HPCHEM\1\DATA\10-16-01\W8925.D\ECD1B.CH Vial: 4
 Signal #2 : C:\HPCHEM\1\DATA\10-16-01\W8925.D\ECD2A.CH
 Acq On : 10-16-01 13:36:22 Operator: LAG
 Sample : CHLOR_C_IAS_1534,0.5_PPM Inst : GC_W
 Misc : NA,NA,NA,1 Multiplr: 1.00
 IntFile Signal #1: autoint1.e IntFile Signal #2: autoint2.e
 Quant Time: Oct 17 11:29 2001 Quant Results File: WPTC1003.RES

Quant Method : C:\HPCHEM\1\METHODS\WPTC1003.M (Chemstation Integrator)
 Title :
 Last Update : Thu Oct 04 10:37:06 2001
 Response via : Multiple Level Calibration
 DataAcq Meth : WPTC1003.M

Volume Inj. :
 Signal #1 Phase :
 Signal #1 Info :
 Signal #2 Phase :
 Signal #2 Info :



INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

BLANK RESULTS SUMMARY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil

Concentration/Units: ppm (mg/kg)

ANALYTE	INITIAL CALIBRATION	CONTINUING CALIBRATION		INST. MDL	REAGENT BLANK	MDL	IDL
		1	2				
Aluminum	ND	ND	ND	0.010	ND	10.0	
Antimony	ND	ND	ND	0.001	ND	1.00	
Arsenic	ND	ND	ND	0.001	ND	1.00	
Barium	ND	ND	ND	0.010	ND	10.0	
Beryllium	ND	ND	ND	0.0005	ND	0.500	
Cadmium	ND	ND	ND	0.00025	ND	0.250	
Calcium	ND	ND	ND	0.050	ND	50.0	
Chromium	ND	ND	ND	0.002	ND	2.00	
Cobalt	ND	ND	ND	0.002	ND	2.00	
Copper	ND	ND	ND	0.002	ND	2.00	
Iron	ND	ND	ND	0.025	ND	25.0	
Lead	ND	ND	ND	0.0005	ND	0.500	
Magnesium	ND	ND	ND	0.050	ND	50.0	
Manganese	ND	ND	ND	0.005	ND	5.00	
Mercury	ND	ND	ND	0.00025	ND	0.013	
Nickel	ND	ND	ND	0.001	ND	1.00	
Potassium	ND	ND	ND	0.050	ND	50.0	
Selenium	ND	ND	ND	0.002	ND	2.00	
Silver	ND	ND	ND	0.0005	ND	0.500	
Sodium	ND	ND	ND	0.100	ND	100	
Thallium	ND	ND	ND	0.0001	ND	0.100	
Vanadium	ND	ND	ND	0.002	ND	2.00	
Zinc	ND	ND	ND	0.002	ND	2.00	

This QC Blank applies to the following samples:

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1
6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1
6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1
7056-1; 7083-1-8; 7085-3; 7051-1

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

INITIAL & CONTINUING CALIBRATION VERIFICATION

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Concentration/Units: ppb ($\mu\text{g/L}$)

ANALYTE	INITIAL CALIBRATION			CONTINUING CALIBRATION					MDL	IDL
	TRUE	FOUND	%R (1)	TRUE	FOUND	%R (1)	FOUND	%R (1)		
Aluminum	400	436	109	400	428	107	416	104	10.0	
Antimony	120	132	110	120	131	109	114	95.0	1.00	
Arsenic	20.0	20.3	102	20.0	20.1	101	19.8	99.0	1.00	
Barium	400	421	105	400	427	107	365	91.3	10.0	
Beryllium	10.0	10.6	106	10.0	10.3	103	9.32	93.2	0.500	
Cadmium	10.0	10.2	102	10.0	10.2	102	10.1	101	0.250	
Calcium	10000	10300	103	10000	10400	104	10100	101	50.0	
Chromium	20.0	21.2	106	20.0	21.4	107	21.2	106	2.00	
Cobalt	100	102	102	100	103	103	102	102	2.00	
Copper	50.0	51.8	104	50.0	51.4	103	51.4	103	2.00	
Iron	200	208	104	200	214	107	183	91.5	25.0	
Lead	10.0	10.6	106	10.0	10.6	106	10.4	104	0.500	
Magnesium	10000	9920	99.2	10000	10100	101	10900	109	50.0	
Manganese	30.0	31.7	106	30.0	32.1	107	32.9	110	5.00	
Mercury	5.00	5.11	102	5.00	5.21	104	4.81	96.2	0.250	
Nickel	80.0	83.8	105	80.0	83.5	104	83.8	105	1.00	
Potassium	10000	10400	104	10000	10500	105	11000	110	50.0	
Selenium	10.0	10.5	105	10.0	10.6	106	11.0	110	2.00	
Silver	20.0	20.1	101	20.0	20.1	101	19.0	95.0	0.500	
Sodium	10000	10500	105	10000	10300	103	10900	109	100	
Thallium	20.0	21.1	106	20.0	21.5	108	20.9	105	0.100	
Vanadium	100	108	108	100	110	110	99.7	99.7	2.00	
Zinc	40.0	43.1	108	40.0	42.3	106	42.4	106	2.00	

(1) Control Limits: Mercury 80-120, Other Metals 90-110

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

ICP-MS ICSAB RESULTS SUMMARY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Aqueous

Concentration/Units: ppb(ug/L)

ANALYTE / MASS	INITIAL		
	SOL A	SOL AB	%R
Arsenic / 75	0.48	9.49	90.1
Cobalt / 59	0.30	9.66	93.6
Copper / 65	2.08	11.3	92.2
Manganese / 55	3.61	12.9	92.9
Nickel / 60	1.33	10.2	88.7
Zinc / 66	8.14	16.9	87.6

FINAL		
SOL A	SOL AB	%R
0.56	9.28	87.2
0.31	9.36	90.5
2.01	10.8	87.9
3.56	12.6	90.4
1.44	10.6	91.6
7.94	16.2	82.6

TRUE VALUE SOLN. B = 10 ppb

THERE ARE NO CONTROL LIMITS FOR SOLUTION AB

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

SPIKE SAMPLE RECOVERY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil

Concentration/Units: ppm (mg/kg)

ANALYTE	SSR1	SR1	%R1	SA1	SSR2	SR2	%R2	SA2	CONTROL LIMIT %R
Aluminum	3850	3370	NC	214	12500	11500	NC	216	70-130
Antimony	50.9	ND	119	42.8	50.6	ND	117	43.2	70-130
Arsenic	44.7	ND	104	42.8	44.4	1.37	99.6	43.2	70-130
Barium	65.7	19.4	108	42.8	105	60.0	104	43.2	70-130
Beryllium	41.2	ND	96.3	42.8	42.3	0.770	96.1	43.2	70-130
Cadmium	41.8	ND	97.7	42.8	40.0	ND	92.6	43.2	70-130
Calcium	1380	1110	NC	214	6150	5730	NC	216	70-130
Chromium	49.9	5.73	103	42.8	53.1	9.25	102	43.2	70-130
Cobalt	45.4	2.76	99.6	42.8	48.9	6.59	97.9	43.2	70-130
Copper	51.1	8.56	99.4	42.8	57.1	13.9	100	43.2	70-130
Iron	6120	5660	NC	214	17700	16900	NC	216	70-130
Lead	50.1	3.12	110	42.8	49.1	3.62	105	43.2	70-130
Magnesium	1930	1520	NC	214	4380	3980	NC	216	70-130
Manganese	211	164	110	42.8	471	424	109	43.2	70-130
Mercury	0.272	ND	101	0.268	0.274	ND	101	0.270	75-125
Nickel	50.5	6.29	103	42.8	50.1	6.46	101	43.2	70-130
Potassium	1190	956	109	214	1020	767	117	216	70-130
Selenium	45.2	ND	106	42.8	44.3	ND	103	43.2	70-130
Silver	42.1	ND	98.4	42.8	40.6	ND	94.0	43.2	70-130
Sodium	368	113	119	214	483	231	117	216	70-130
Thallium	43.9	ND	103	42.8	43.2	ND	100	43.2	70-130
Vanadium	52.1	7.71	104	42.8	62.6	18.6	102	43.2	70-130
Zinc	57.7	15.2	99.3	42.8	86.5	44.8	96.5	43.2	70-130

SSR = Spike Sample Result
 SA = Spike Added

SR = Sample Result
 %R = Percent Recovery

NC = Non-calculable % R; Sample concentration > 4 x Spike Concentration.

QC Sample 1 6953-029

QC Sample 2 6942-001

QC Sample 1 for following samples:

QC Sample 2 for following samples:

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1
6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1
6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1
7056-1; 7083-1-8; 7085-3; 7051-1

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1
6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1
6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1
7056-1; 7083-1-8; 7085-3; 7051-1

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

SPIKE SAMPLE RECOVERY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil

Concentration/Units: ppm (mg/kg)

ANALYTE	SSR3	SR3	%R3	SA3	SSR4	SR4	%R4	SA4	CONTROL LIMIT %R
Aluminum	4430	4100	NC	224					70-130
Antimony	51.7	ND	115	44.8					70-130
Arsenic	45.8	ND	102	44.8					70-130
Barium	64.8	19.5	101	44.8					70-130
Beryllium	40.0	ND	89.3	44.8					70-130
Cadmium	42.2	ND	94.2	44.8					70-130
Calcium	2920	2650	121	224					70-130
Chromium	52.7	6.92	102	44.8					70-130
Cobalt	47.6	3.19	99.1	44.8					70-130
Copper	53.8	10.8	96.0	44.8					70-130
Iron	6970	6800	75.9	224					70-130
Lead	50.9	3.80	105	44.8					70-130
Magnesium	2140	1880	116	224					70-130
Manganese	129	82.0	105	44.8					70-130
Nickel	52.2	7.10	101	44.8					70-130
Potassium	1360	1110	112	224					70-130
Selenium	45.7	ND	102	44.8					70-130
Silver	43.0	ND	96.0	44.8					70-130
Sodium	450	209	108	224					70-130
Thallium	44.5	ND	99.3	44.8					70-130
Vanadium	55.0	9.53	101	44.8					70-130
Zinc	62.2	18.5	97.5	44.8					70-130

SSR = Spike Sample Result
 SA = Spike Added

SR = Sample Result
 %R = Percent Recovery

NC = Non-calculable % R; Sample concentration > 4 x Spike Concentration.

QC Sample 3 6996-014

QC Sample 4 _____

QC Sample 3 for following samples:

QC Sample 4 for following samples:

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1

6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1

6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1

7056-1; 7083-1-8; 7085-3; 7051-1

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

DUPLICATE SAMPLE RECOVERY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil

Concentration/Units: ppm (mg/kg)

ANALYTE	CONTROL LIMIT 1	S1	D1	RPD1	CONTROL LIMIT 2	S2	D2	RPD2
Aluminum	20	3370	3420	1.47	20	11500	11500	0
Antimony	NA	ND	ND	NC	NA	ND	ND	NC
Arsenic	NA	ND	ND	NC	20	1.37	1.34	2.21
Barium	20	19.4	20.3	4.53	20	60.0	59.0	1.68
Beryllium	NA	ND	ND	NC	20	0.770	0.782	1.55
Cadmium	NA	ND	ND	NC	NA	ND	ND	NC
Calcium	20	1110	1140	2.67	20	5730	5500	4.10
Chromium	20	5.73	5.87	2.41	20	9.25	9.39	1.50
Cobalt	20	2.76	2.78	0.722	20	6.59	5.55	17.1
Copper	20	8.56	8.47	1.06	20	13.9	14.6	4.91
Iron	20	5660	5880	3.81	20	16900	17300	2.34 ?
Lead	20	3.12	3.28	5.00	20	3.62	3.56	1.67
Magnesium	20	1520	1570	3.24	20	3980	4210	5.62
Manganese	20	164	167	1.81	20	424	420	0.948
Mercury	NA	ND	ND	NC	NA	ND	ND	NC
Nickel	20	6.29	6.49	3.13	20	6.46	6.55	1.38
Potassium	20	956	1010	5.49	20	767	870	12.6
Selenium	NA	ND	ND	NC	NA	ND	ND	NC
Silver	NA	ND	ND	NC	NA	ND	ND	NC
Sodium	20	113	129	13.2	20	231	257	10.7
Thallium	NA	ND	ND	NC	NA	ND	ND	NC
Vanadium	20	7.71	7.68	0.390	20	18.6	19.6	5.24
Zinc	20	15.2	15.7	3.24	20	44.8	45.3	1.11

S1 = Sample 1

D1 = Duplicate 1

S2 = Sample 2

D2 = Duplicate 2

NA = Not Applicable

NC = Non-calculable RPD due to result (s) less than the detection limit.

QC Sample 1 6953-029

QC Sample 2 6942-001

QC Sample 1 for following samples:

6879-1, 3, 5, 7, 9; 6942-1*4; 6967-1; 6968-1; 6969-1

6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1*4; 7035-1

6996-1, 4, 7, 11, 14, 17; 7034-1*3, 6, 9, 12; 7041-1

7056-1; 7083-1*8; 7085-3; 7051-1

QC Sample 2 for following samples:

6879-1, 3, 5, 7, 9; 6942-1*4; 6967-1; 6968-1; 6969-1

6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1*4; 7035-1

6996-1, 4, 7, 11, 14, 17; 7034-1*3, 6, 9, 12; 7041-1

7056-1; 7083-1*8; 7085-3; 7051-1

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

DUPLICATE SAMPLE RECOVERY

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil

Concentration/Units: ppm (mg/kg)

ANALYTE	CONTROL LIMIT 3	S3	D3	RPD3	CONTROL LIMIT 4	S4	D4	RPD4
Aluminum	20	4100	4280	4.30				
Antimony	NA	ND	ND	NC				
Arsenic	NA	ND	ND	NC				
Barium	20	19.5	20.5	5.00				
Beryllium	NA	ND	ND	NC				
Cadmium	NA	ND	ND	NC				
Calcium	20	2650	2610	1.52				
Chromium	20	6.92	7.10	2.57				
Cobalt	20	3.19	3.15	1.26				
Copper	20	10.8	10.9	0.922				
Iron	20	6800	6910	1.60				
Lead	20	3.80	3.97	4.38				
Magnesium	20	1880	1860	1.07				
Manganese	20	82.0	83.8	2.17				
Nickel	20	7.10	7.08	0.282				
Potassium	20	1110	1190	6.96				
Selenium	NA	ND	ND	NC				
Silver	NA	ND	ND	NC				
Sodium	20	209	219	4.67				
Thallium	NA	ND	ND	NC				
Vanadium	20	9.53	9.75	2.28				
Zinc	20	18.5	18.8	1.61				

S3 = Sample 3

D3 = Duplicate 3

S4 = Sample 4

D4 = Duplicate 4

NA = Not Applicable

NC = Non-calculable RPD due to result (s) less than the detection limit.

QC Sample 3 6996-014

QC Sample 4 _____

QC Sample 3 for following samples:

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1

6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1

6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1

7056-1; 7083-1-8; 7085-3; 7051-1

QC Sample 4 for following samples:

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

LABORATORY CONTROL SAMPLE

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

ANALYTE	AQUEOUS (ug/L)			SOLID (mg/kg)			
	TRUE	FOUND	%R(1)	TRUE	FOUND	LIMITS	%R(2)
Aluminum				5760	6060	3750-7760	105
Antimony				26.2	7.92	D.L.-53.1	30.2
Arsenic				36.2	38.2	26.9-45.4	106
Barium				117	123	90.2-144	105
Beryllium				59.4	74.3	60.4-93.7	125
Cadmium				35.6	33.9	27.4-43.8	95.2
Calcium				1320	1300	989-1660	98.5
Chromium				108	108	79.2-137	100
Cobalt				67.2	60.8	53.4-80.9	90.5
Copper				63.7	60.9	52.2-75.2	95.6
Iron				7540	6850	4290-10800	90.8
Lead				52.4	54.0	37.3-67.6	103
Magnesium				1190	1200	815-1570	101
Manganese				151	151	128-189	100
Mercury				1.00	1.03	.614-1.39	103
Nickel				49.8	49.3	39-60.6	99.0
Potassium				2130	1910	1660-2600	89.7
Selenium				56.8	47.3	41-72.6	83.3
Silver				34.1	36.3	25.4-42.9	106
Sodium				487	651	354-911	134
Thallium				68.2	69.4	39-97.3	102
Vanadium				80.0	78.6	54.6-105	98.3
Zinc				151	137	117-185	90.7

(1) Control Limits = 90-110%

(2) Manufacture's Recommended Limits

Aqueous LCS Source: MV Laboratories

Solid LCS Source: Environmental Resource Associates: PriorityPollutnT/CLP Inorganics Soils

INTEGRATED ANALYTICAL LABORATORIES

METALS QUALITY CONTROL

SERIAL DILUTIONS & POST SPIKES

Lab Case Number: E01- 6879, 6942, 6953, 6967, 6968, 6969, 6996, 7022, 7034, 7035, 7041
7051, 7056, 7083, 7085

Matrix: Soil Concentration/Units: ppm (mg/kg)

ANALYTE	SERIAL DILUTION		%	POST SPIKE		%
	SR	SDR		SPR	SA	
Aluminum	3370	3600	6.60			
Antimony	ND			41.5	42.8	97.0
Arsenic	ND			41.8	42.8	97.7
Barium	19.4	19.5	0.514			
Beryllium	ND			42.6	42.8	99.5
Cadmium	ND			40.8	42.8	95.3
Calcium	1110	1150	3.54			
Chromium	5.73	5.53	3.55			
Cobalt	2.76	2.82	2.15			
Copper	8.56	8.35	2.48			
Iron	5660	5700	0.704			
Lead	3.12	3.01	3.59			
Magnesium	1520	1630	6.98			
Manganese	164	166	1.21			
Mercury	ND			0.276	0.268	103
Nickel	6.29	6.46	2.67			
Potassium	956	916	4.27			
Selenium	ND			40.9	42.8	95.6
Silver	ND			42.3	42.8	98.8
Sodium	113			944	856	97.1
Thallium	ND			44.0	42.8	103
Vanadium	7.71	7.70	0.130			
Zinc	15.2	15.2	0			

SR = Sample Result

SDR = Sample Dilution Result

SPR = Sample Post Spike Result

SA = Spike Added

Control Limits: (+) or (-) 10% Difference or 85 - 115% Recovery

QC Sample : 6953-029

QC Sample for following samples:

6879-1, 3, 5, 7, 9; 6942-1-4; 6967-1; 6968-1; 6969-1
6953-1, 4, 7, 9, 11, 13, 15, 18, 27, 29; 7022-1-4; 7035-1
6996-1, 4, 7, 11, 14, 17; 7034-1-3, 6, 9, 12; 7041-1
7056-1; 7083-1-8; 7085-3; 7051-1

General Chemistry Quality Control

Matrix: Soil
Units: S-mg/Kg

Batch ID: AP024-0114

Blank

Parameter	Method Blank	MDL	Analysis Date
Total Recoverable Phenols	ND	2.50	10/12/01

Duplicate Recovery

Parameter	QC Sample	Result	Duplicate Result	RPD	RPD Limits
Total Recoverable Phenols	7034-001S	53.7	53.3	1	20

Spike Recovery

Parameter	QC Sample	Result	Spike Added	Spike Result	%Spike Recovery	%Rec. Limits
Total Recoverable Phenols	7034-001	ND	55.2	53.7	97.3	75 - 125

The above blank result applies to the follow samples E01-

7034-001	7023-004
7034-001D	7023-005
7034-001S	7023-006
7034-001SD	7023-007
7034-002	7023-008
7034-003	7023-010
7034-006	7023-011
7034-009	7023-013
7034-012	7023-014
7023-001	7023-015
7023-002	7023-016
	7041-001

++ = No Flash - Sample boiled at 100C
NA = Not Applicable
ND = Not Detected
NC = Non calculable RPD due to value less than the detection limit
No = Does Not Ignite
Yes = Sample Ignites

General Chemistry Quality Control

Matrix: Soil
Units: S-mg/Kg

Batch ID: AP013-0098

Blank

Parameter	Method Blank	MDL	Analysis Date
Total Cyanide	ND	1.00	10/23/01

Duplicate Recovery

Parameter	QC Sample	Result	Duplicate Result	RPD	RPD Limits
Total Cyanide	7041-001S	15.5	15.2	2	20

Spike Recovery

Parameter	QC Sample	Result	Spike Added	Spike Result	%Spike Recovery	%Rec. Limits
Total Cyanide	7041-001	ND	0	15.5	112	75 - 125

The above blank result applies to the follow samples E01-

7041-001	7289-002
7041-001D	7320-001
7041-001S	7320-006
7041-001SD	7314-001
7129-001	7314-002
7213-001	7314-003
7288-001	7314-004
7288-002	7314-005
7289-001	7314-006

++ = No Flash - Sample boiled at 100C
NA = Not Applicable
ND = Not Detected
NC = Non calculable RPD due to value less than the detection limit
No = Does Not Ignite
Yes = Sample Ignites

Turnaround Time

Turnaround Time					Report Format	
<u>Conditional / TPHC</u>					Results Only Reduced Regulatory SNP Disk** dlf gr wkl	
24 hr*	48 hr	72 hr	1 wk	NA		Other: Std
<u>Verbal/Fax</u>						
24 hr*	48 hr*	72 hr*	1 wk*	2 wk		Other: Std
<u>Hard Copy</u>						
72 hr*	1 wk*	2 wk*	3 wk		Other: Std	

*Prior to sample arrival, Lab notification is required.

ANALYTICAL PARAMETERS / PRESERVATIVES

[illegible]

Please print legibly and fill out completely. Samples cannot be processed and the turnaround time will not start until any ambiguities have been resolved.

Concentrations Expected	Known Hazard: yes no
LOW MED HIGH	Describe:

Lab Case #

1702

LAB COPIES - WHITE & YELLOW; CLIENT COPY - PINK

CHAIN OF CUSTODY

Case No.: <u>E01-7041</u>	P.O. #: _____
Project : <u>99704 FINETEX</u>	
Client/Project: <u>BRENNAN/99704 FINETEX</u>	
Client Address:	Billing Address:
<u>Brennan Environmental, Inc.</u>	<u>Brennan Environmental, Inc.</u>
<u>8 Great Meadow Lane</u>	<u>8 Great Meadow Lane</u>
<u>Unit D</u>	<u>Unit D</u>
<u>East Hanover, NJ 07936</u>	<u>East Hanover, NJ 07936</u>
Date Received: <u>10/11/01</u>	Verbal Due: <u>Oct 26</u>
Time Received: <u>19:10</u>	Report Due: <u>Nov 2</u>
Report Format: <u>Reduced</u>	

# of Containers	2
IAL ID #	7041-001
Client ID #	5-F1
Matrix	Soil
Sample Date	10/11/01
Sample Time	10:17
Methanol Kit	✓
VO+15, PP MP	✓
Pesticides	✓
PCB	✓
BNA + 25	✓
PP Metals-HIGH	✓
Cyanide, Total	✓
Ttl Rec Phenols	✓
% Solids	✓
Electronic Del.	✓

omments:

INTEGRATED ANALYTICAL LABORATORIES, LLC

SAMPLE RECEIPT VERIFICATION

CASE NO:

7041

CLIENT:

BEI

COOLER TEMPERATURE: 2° - 6°C: ☒ (See Chain of Custody)

CHAIN OF CUSTODY:

COMPLETE / INCOMPLETE

Comments:

Sample Bottles Intact:

☒

Comments:

Sample Labels Intact/ Correct:

☒

Sufficient Sample Volume:

☒

Correct bottles/ preservative:

☒

Samples received in

holding time/ prep time:

☒

Headspace/ bubbles in voa samples:

☒

Samples to be subcontracted:

☒

Preserved Sample pH checked:

(Excluding voa samples)

☒

KEY

☒ = YES

☒ = NO

☒ = N/A

ADDITIONAL COMMENTS:

SAMPLE(S) VERIFIED BY:

INITIAL

BE

DATE

10/11/01

CORRECTIVE ACTION REQUIRED:

YES

(SEE BELOW)

NO

CLIENT NOTIFIED:

YES

Date/ Time:

NO

PROJECT CONTACT:

SUBCONTRACTED LAB:

DATE SHIPPED:

ADDITIONAL COMMENTS:

VERIFIED/TAKEN BY:

INITIAL



DATE

Integrated Analytical Laboratories, LLC.

Laboratory Custody Chronicle

Case No : E01-7041
 Client : Brennan Environmental, Inc.
 Project : 99704 FINETEX

	GC/MS V		EXTRACT			ANALYSIS		
			DATE	TIME	INITIAL	DATE	TIME	INITIAL
Methanol Kit	7041-001	S				10/16		DK
VO+15, PP MP	7041-001	S						
GC EX								
Pesticides	7041-001	S	10/15	9:00	EDM2	10/16	3:30	UG
PCB	7041-001	S				10/16	4:00	M
GC/MS X								
BNA + 25	7041-001	S	10/15	7:00 - 3:00	DL	10/15		DL
METALS								
PP Metals-HIGH	7041-001	S	10/15	7:00	g - d	10/15	12:00	DL
WETCHEM								
Cyanide, Total	7041-001	S	10/22	8:00	LB	10/23	1:00	LB
Ttl Rec Phenols	7041-001	S	10/22	8:00	DL	10/23	1:00	DL
% Solids	7041-001	S	10/12		LC	10/18		
AIR								
Electronic Del.	7041-001	S						

REVIEW & APPROVAL: Indymex
 REMARKS :